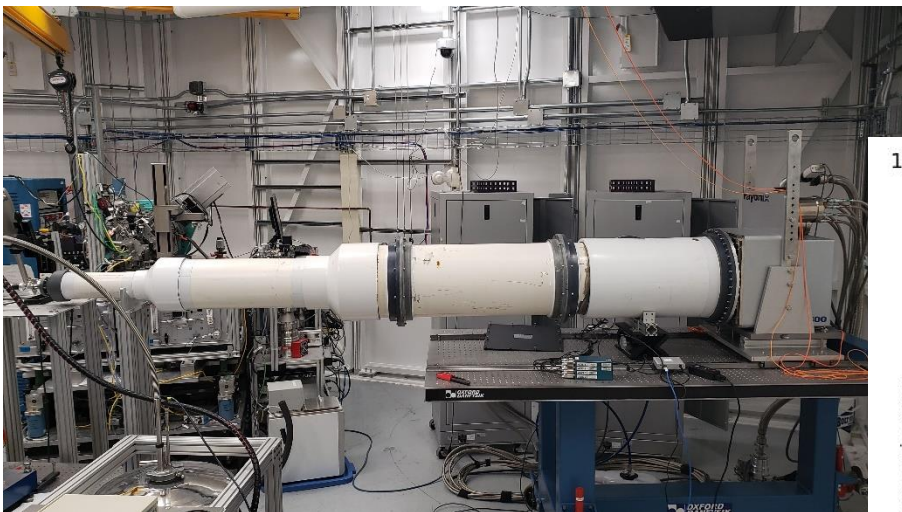
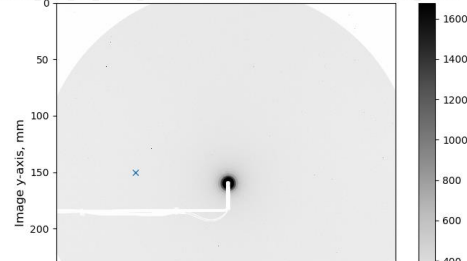


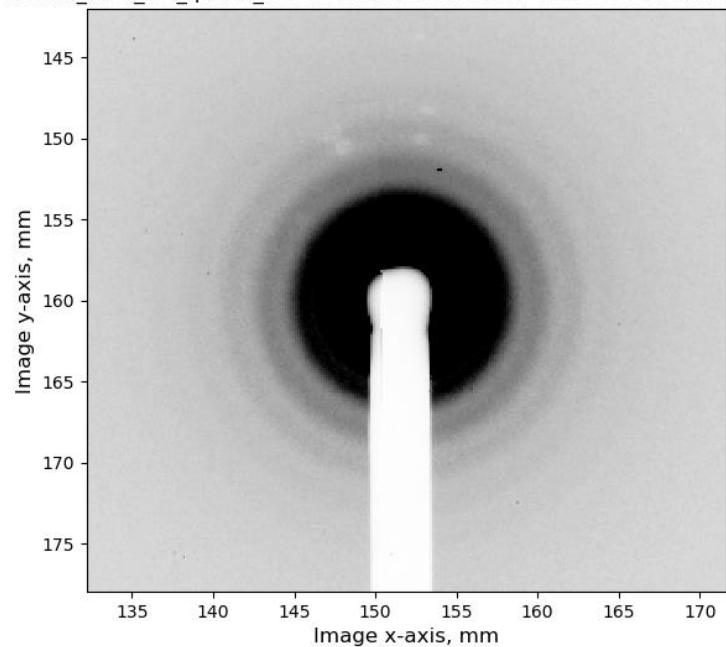
An Introduction to Small Angle X-ray Scattering (SAXS)



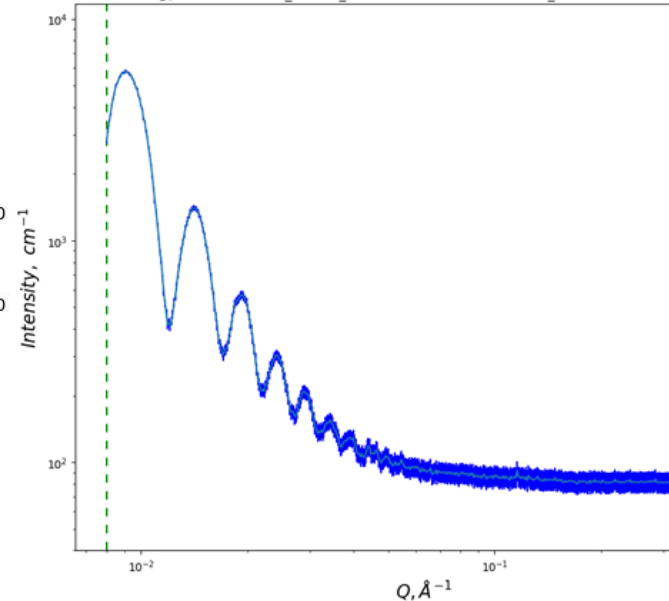
120nm_NIST_std_quartz_300s.S014.N051.4056907486906665264173_365.tif



120nm_NIST_std_quartz_300s.S014.N051.4056907486906665264173_365.tif



log(SASD NISTbeads_120nm_180s.S017.N051.1558669439_189.tif Azm= 270.1



Adam Leontowich
CLS XRD school, Friday, August 18, 2023



Canadian Light Source
Centre canadien de rayonnement synchrotron

The small angle X-ray scattering (SAXS) region

The instrumentation and sample prep

Data collection and reduction

Basic data processing with examples from BXDS



Brockhouse X-ray Diffraction and Scattering (BXDS) sector

Synchrotron
SR JOURNAL OF
SYNCHROTRON
RADIATION
ISSN 1600-5775

The lower energy diffraction and scattering side-bounce beamline for materials science at the Canadian Light Source

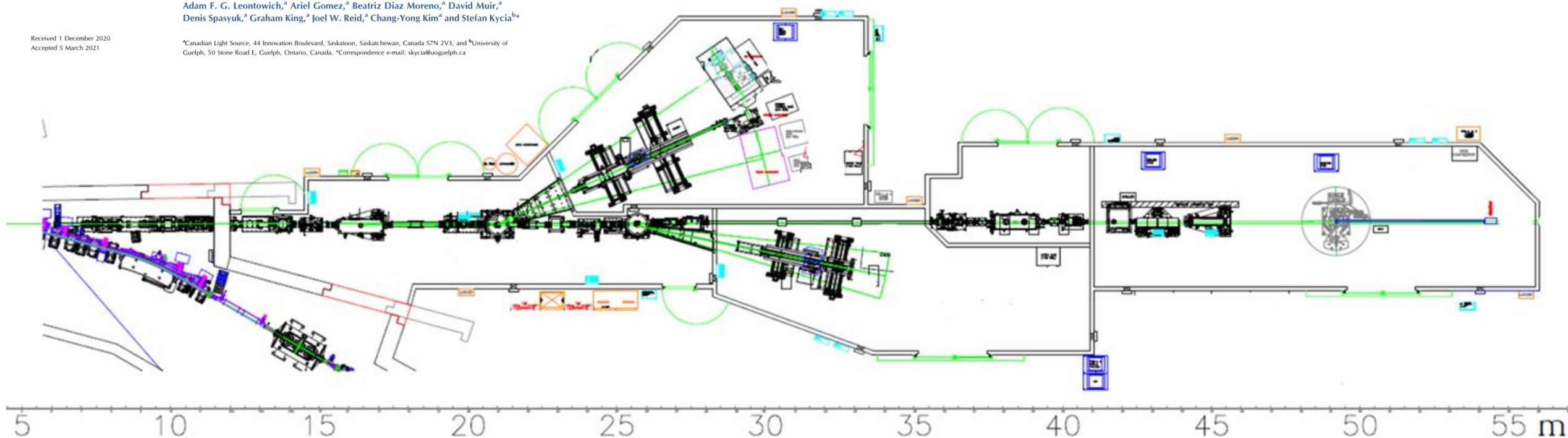
Adam F. G. Leontowich,^a Ariel Gomez,^a Beatriz Diaz Moreno,^a David Muir,^a Denis Spasyuk,^a Graham King,^a Joel W. Reid,^a Chang-Yong Kim^a and Stefan Kycia^{b*}

^aCanadian Light Source, 44 Innovation Boulevard, Saskatoon, Saskatchewan, Canada S7N 2V3, and ^bUniversity of Guelph, 50 Stone Road E, Guelph, Ontario, Canada. *Correspondence e-mail: skykia@uoguelph.ca

Received 1 December 2020
Accepted 5 March 2021



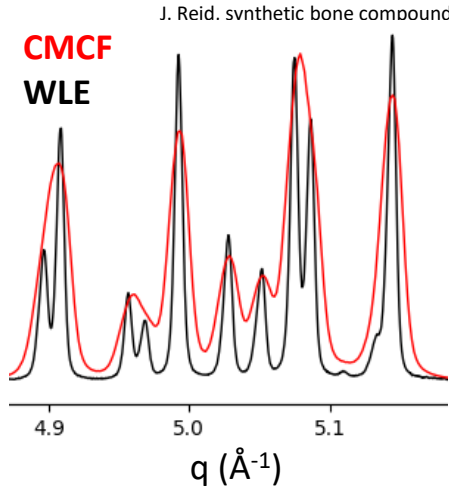
3 “hard X-ray” beamlines (5 – 100 keV)



Canadian Light Source
Centre canadien de rayonnement synchrotron

WLE beamline

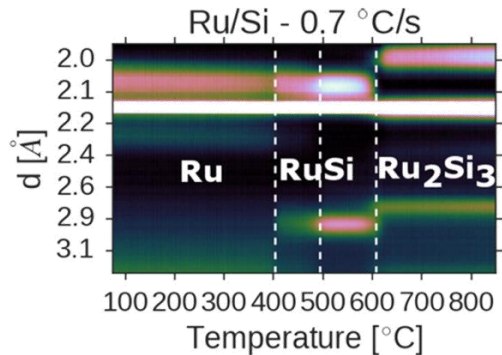
High-resolution powder diffraction



- Peak shape analysis
- Complex mixtures
- Complex structures

IBM (rapid thermal annealing to 1100 °C)

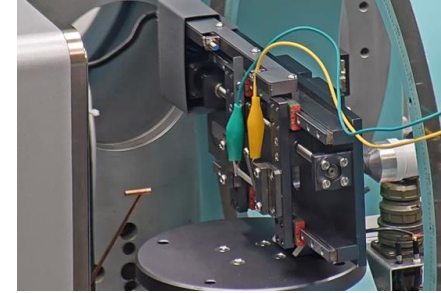
- Thin film studies: XRD, resistivity, roughness, under ultra-high purity gas



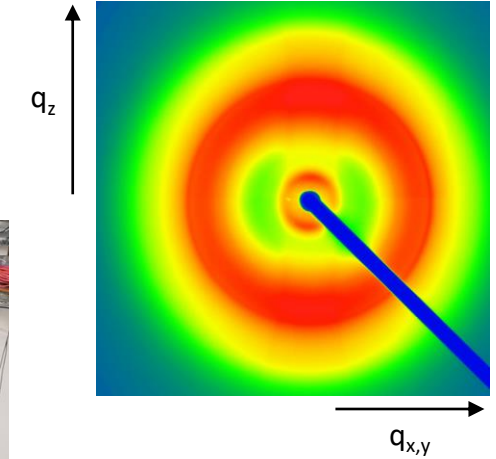
S. Dey et al., J. Vac. Sci. Technol. A 35, 03E109 (2017)

WAXS (XRD with area detector)

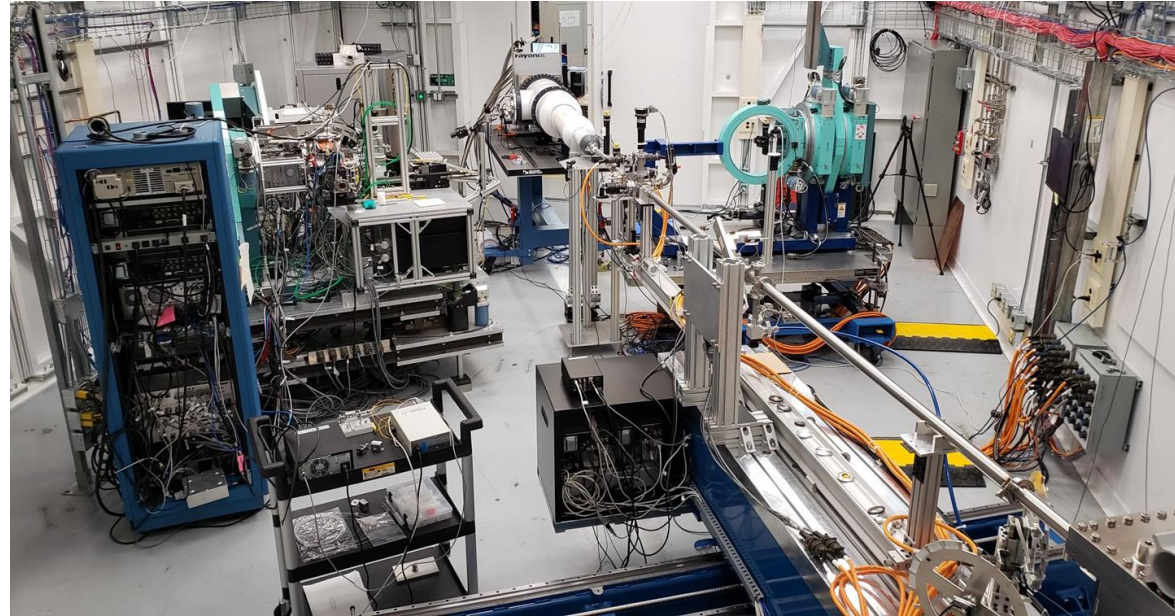
Biopolymers for bandages
Youchao Teng, Yimin Wu, U. Waterloo



Measure WAXS, voltage and current while stretching (0 – 100 N)



- Texture on surfaces
- Degree of orientation
- % crystallinity
- Speed/in-situ

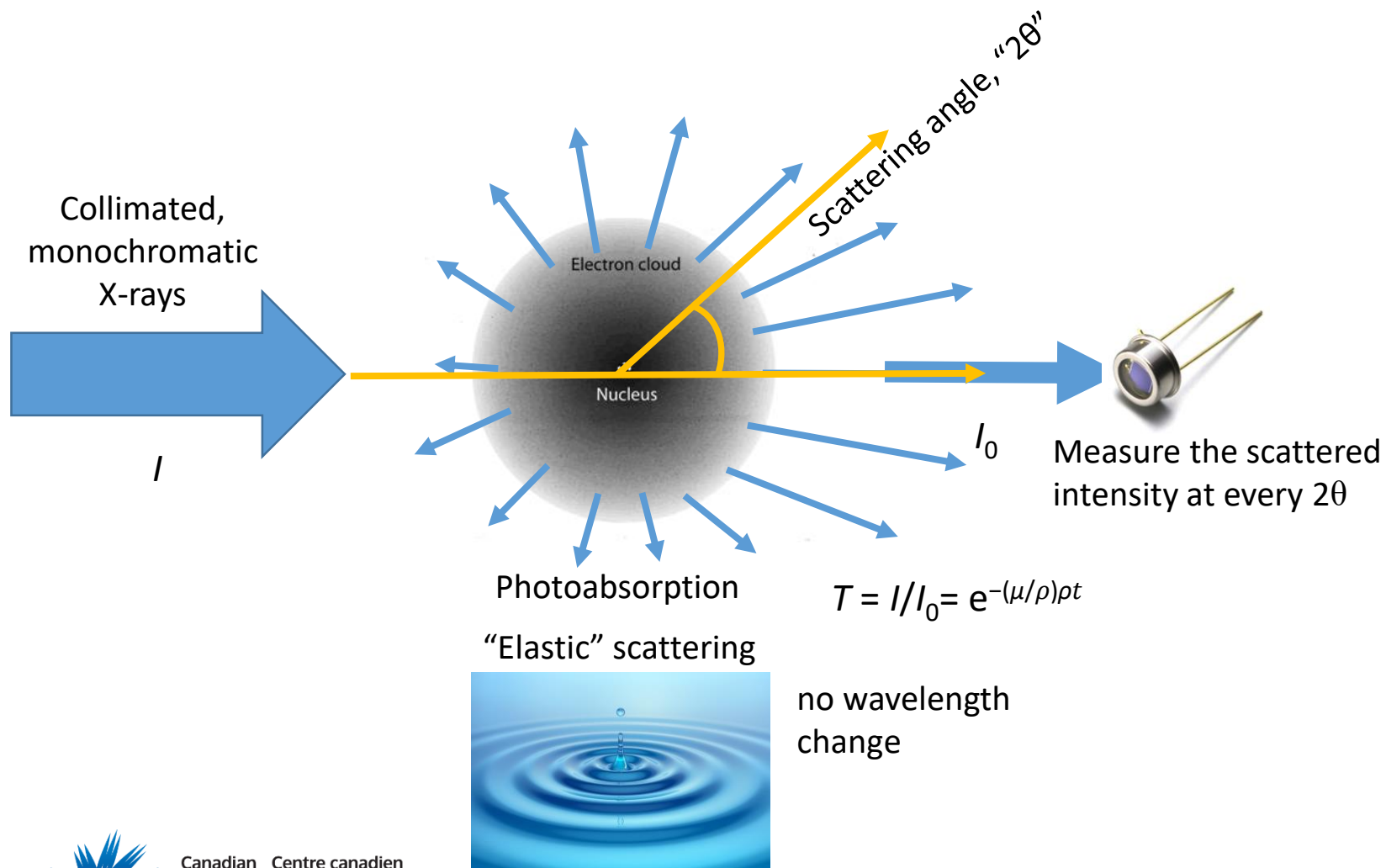


SAXS

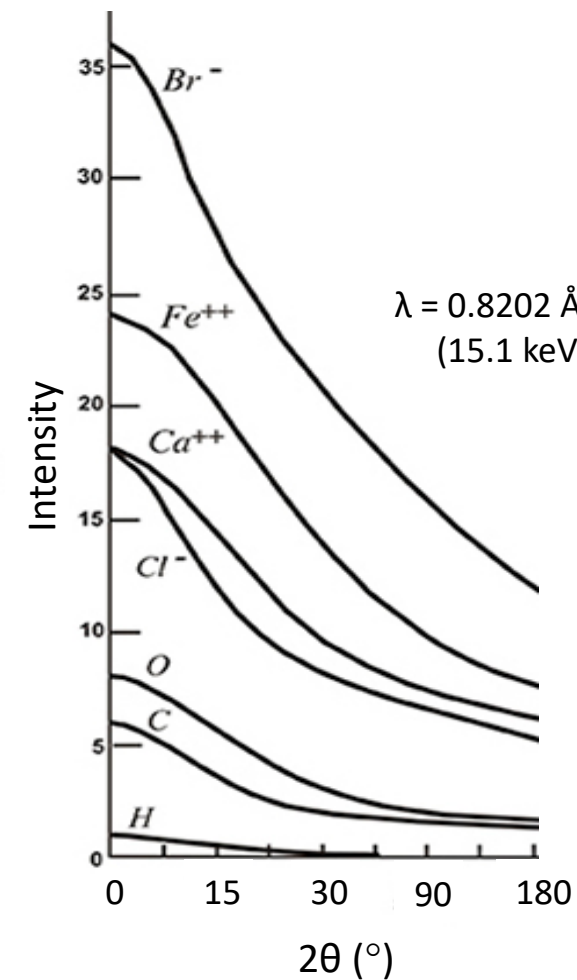
SAXS has been running since February 2021, fairly recent

No dedicated SAXS beamline at CLS

X-ray scattering



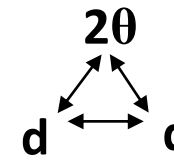
Atomic form factors
(or atomic scattering factors)



From scattering to diffraction

“Bragg” diffraction

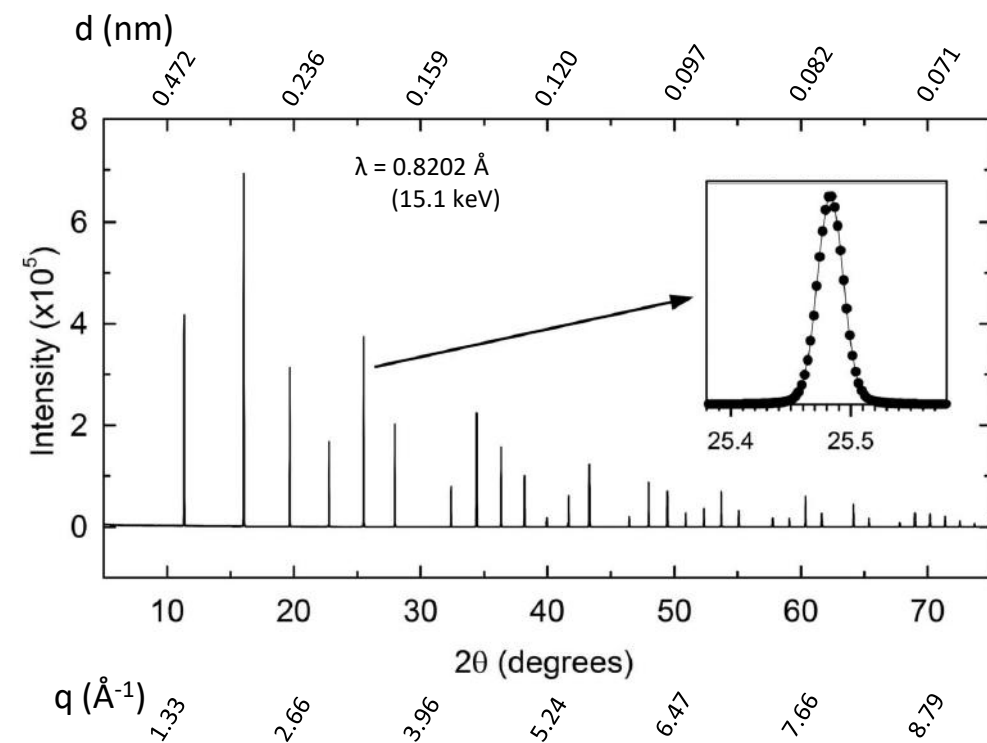
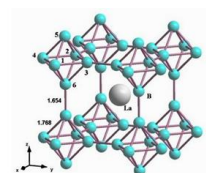
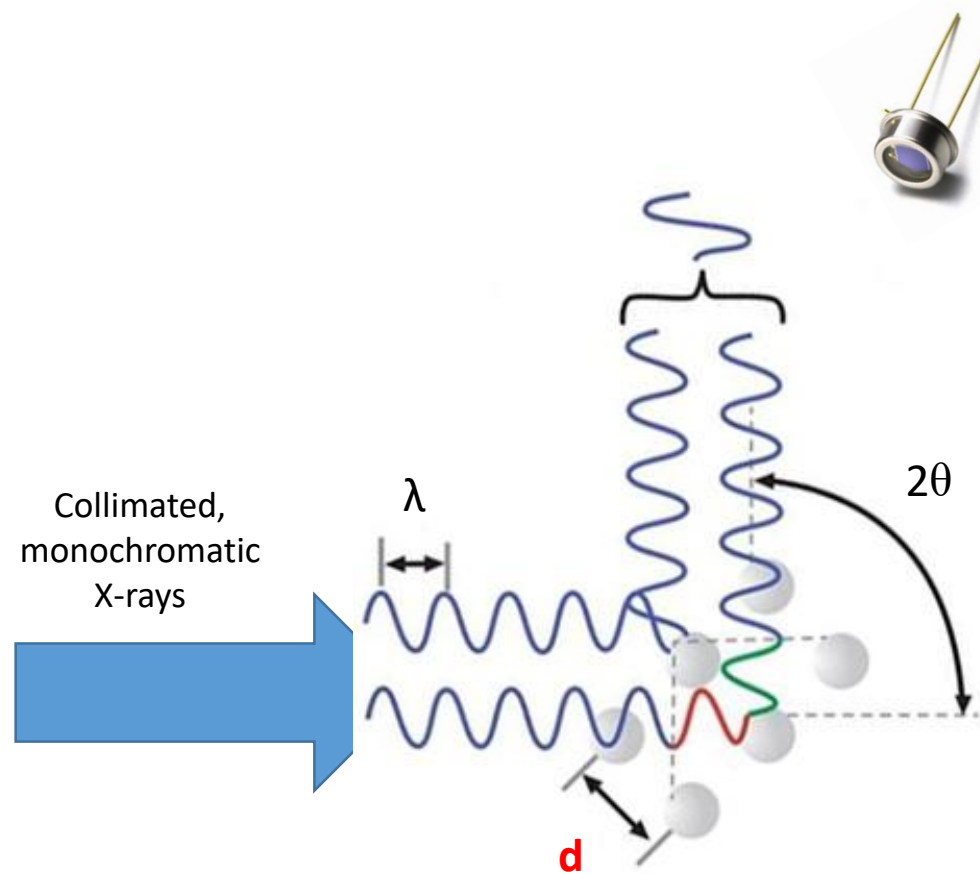
$$n\lambda = 2d\sin\theta$$



$$q = (4\pi\sin\theta)/\lambda$$

$$q = 2\pi/d$$

Constructive interference channels
 scattered intensity into specific directions

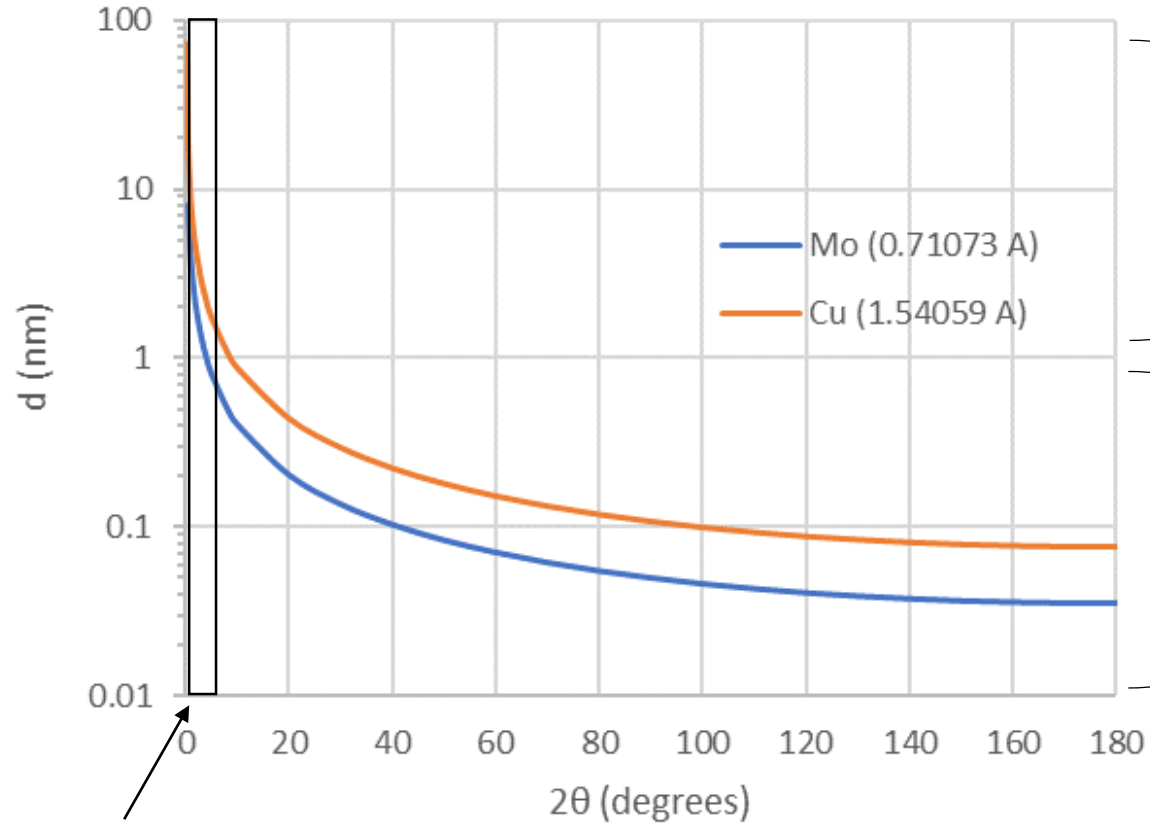
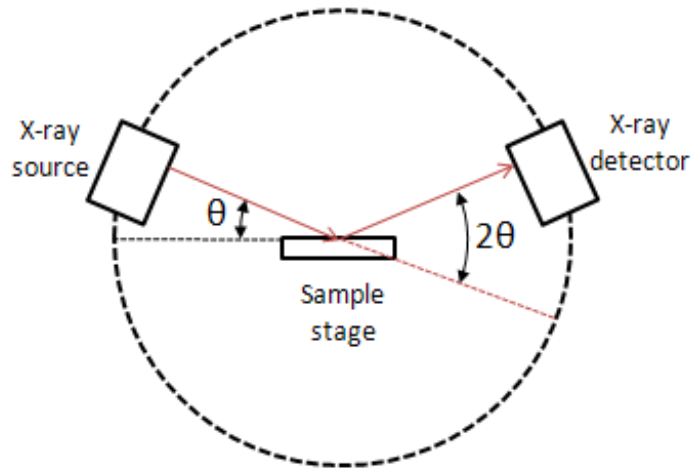


X-ray scattering and diffraction reveals structural order within materials
 on the atomic to >150 nm length scale

What is small angle X-ray scattering (SAXS)?

$$\lambda = 2d\sin\theta$$

Starting from powder diffraction...



The nano world

Atomic distances, spacing between planes of atoms, bond lengths

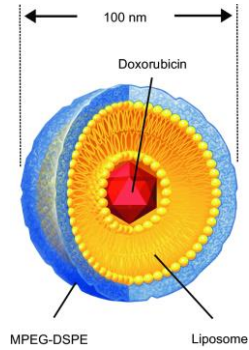
SAXS is generally $2\theta \leq 5^\circ$

Probe relatively big things (1 - 150 nm) by measuring elastic X-ray scattering at small angles

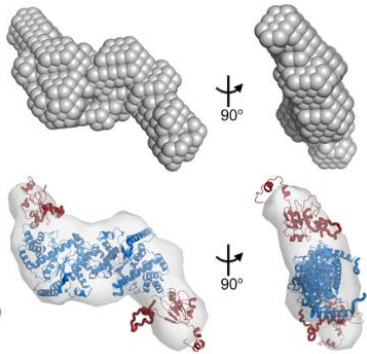


Applications

Drug delivery, Pharmaceuticals

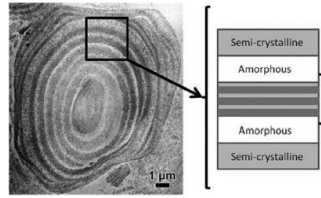


Proteins



define the global shape and conformation in solution

Food science

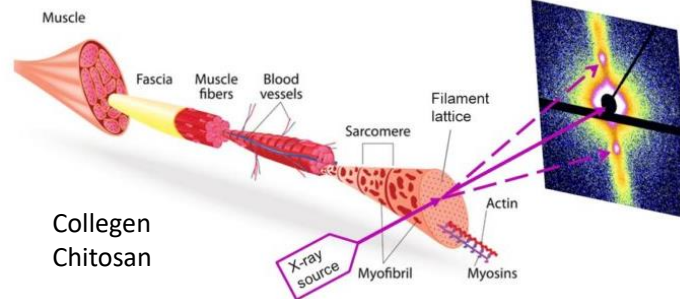


S. Perez and E. Bertoft, *Starch* 2010, 62, 389 - 420



Micelles, emulsions

Biomacromolecules

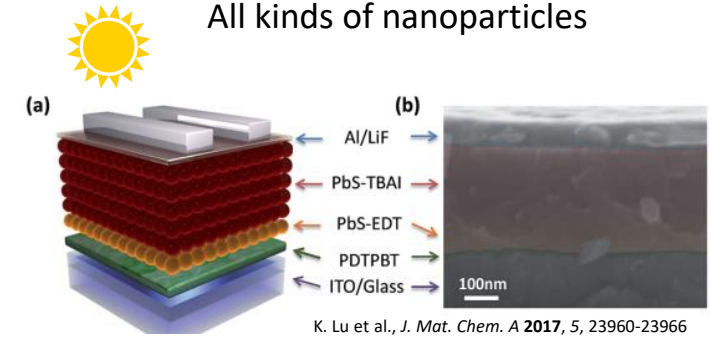


Collagen
Chitosan

<https://www.techexplorist.com/scientists-investigated-pork-fillet-x-ray-light/11572/>

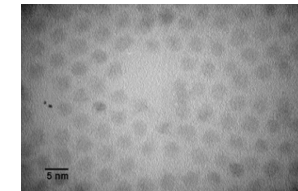
- 1) Packing, ordering
- 2) Size, size distribution
- 3) Shape

All kinds of nanoparticles



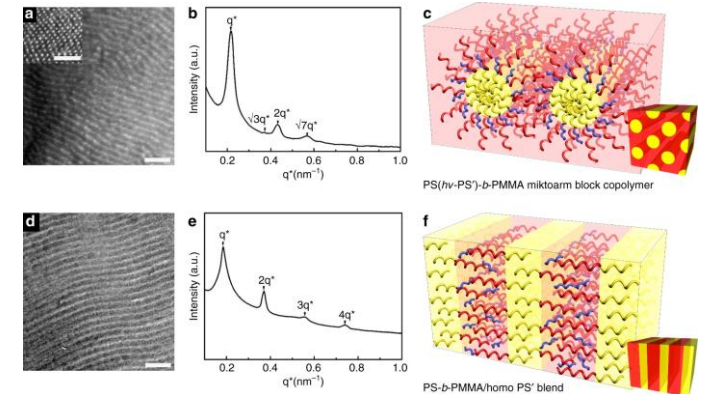
K. Lu et al., *J. Mat. Chem. A* 2017, 5, 23960-23966

Geology



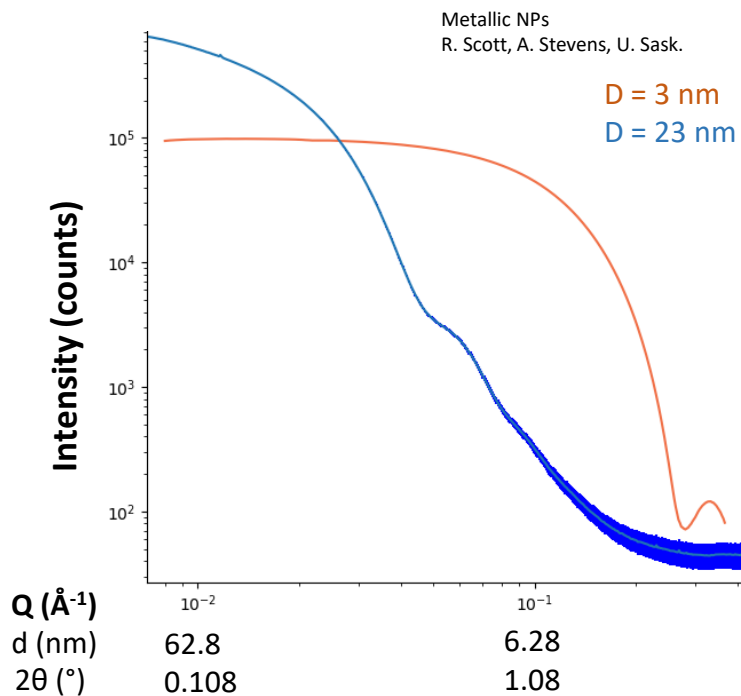
M. Yuan et al., *Adv. Mater.* 2014, 26, 3513-3519

Polymers

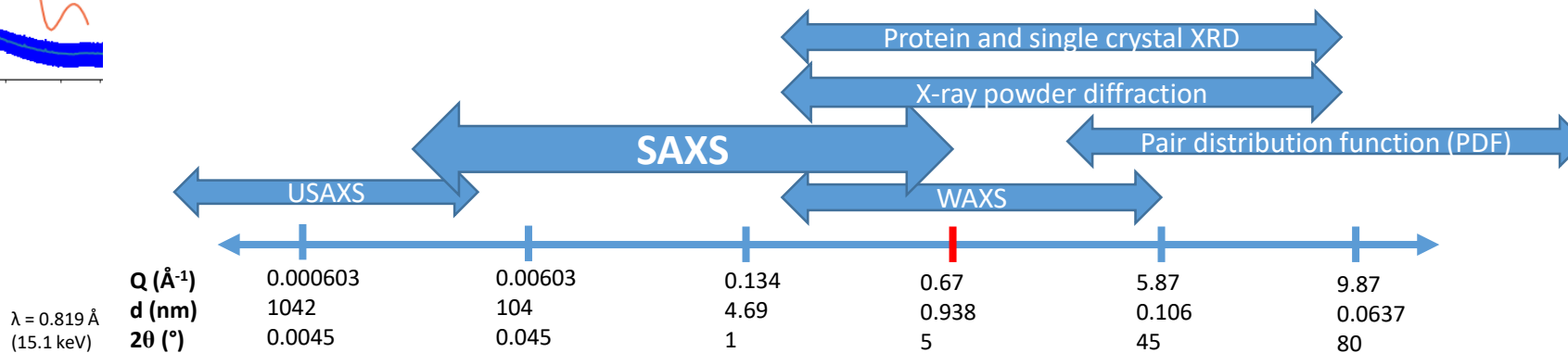
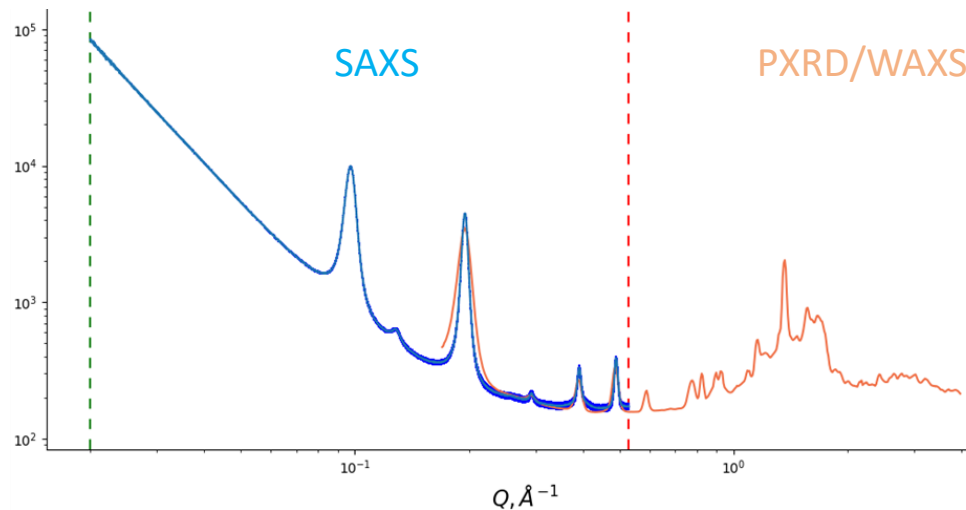


Nature Comm. 8, 1765 (2017)

How is SAXS related to other X-ray diffraction techniques?



- Intensity vs. Q
- log-log plot



Nano-scale

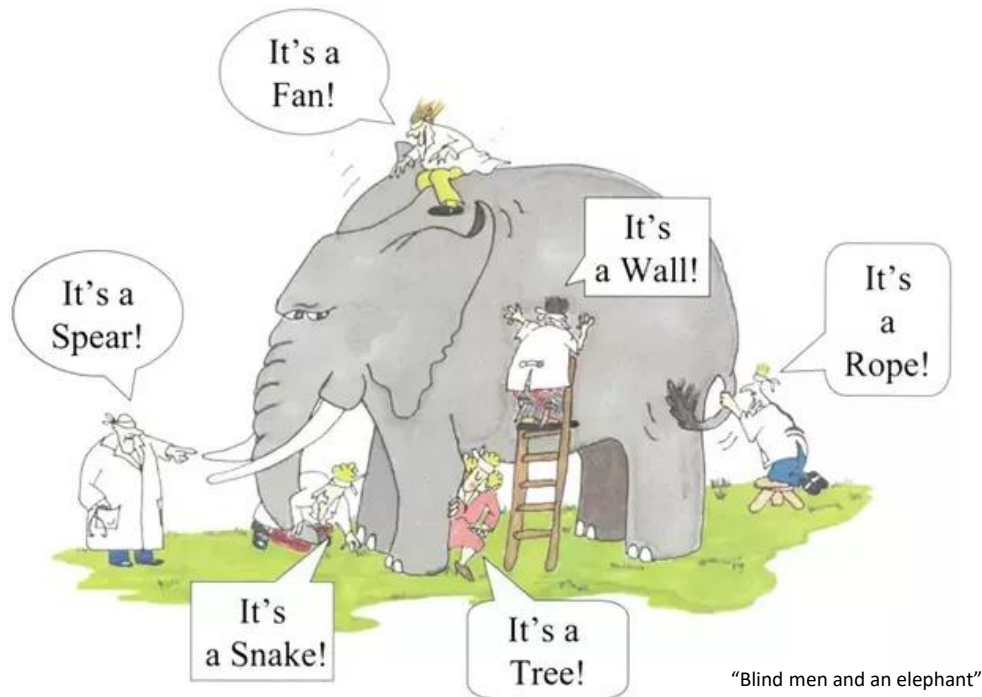
Atomic-scale

Inter-molecular distances, packing arrangement, particle shape, size,

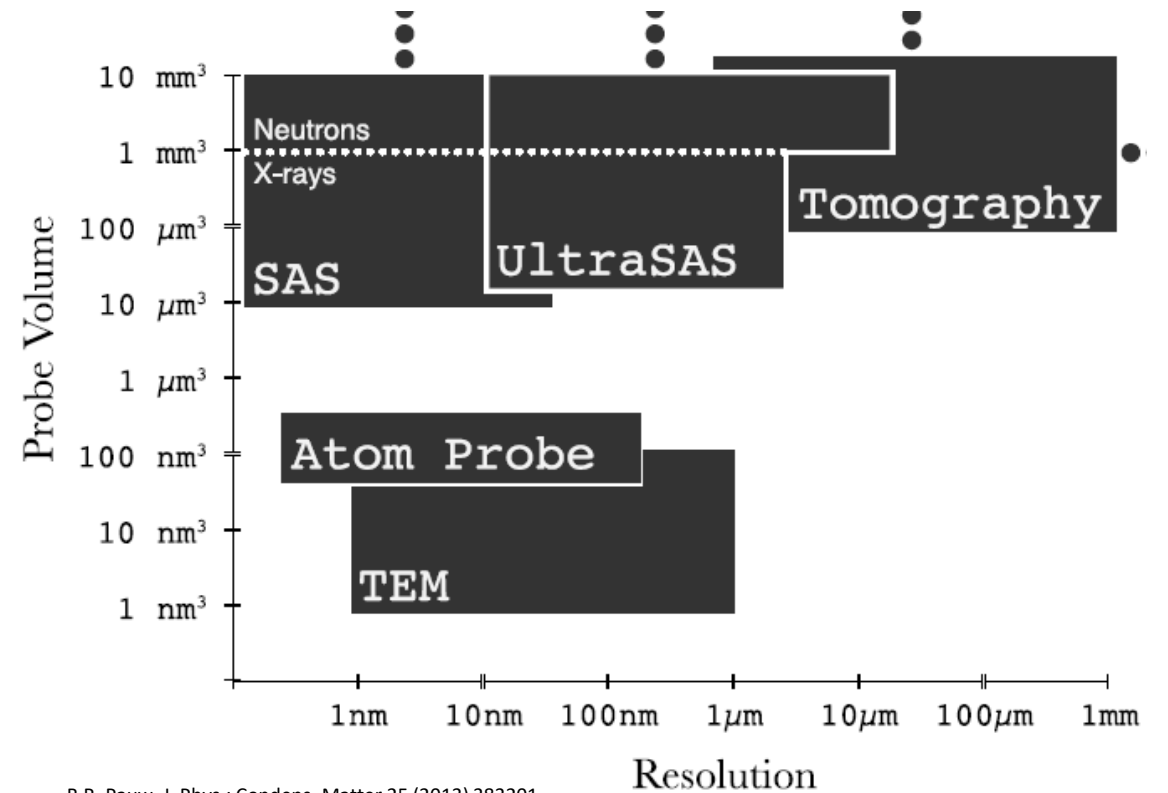
Inter-atomic distances, bond lengths, nearest neighbor atoms
Crystal structures with atomic resolution



How is SAXS complementary to microscopy?



- FOV decreases as resolution increases
- Observe a small fraction of the complete sample at a time
- Challenging sample prep at smaller size



B.R. Pauw, J. Phys.: Condens. Matter 25 (2013) 383201

- SAXS provides nanoscale information averaged over the beam volume (~mm³)
- Complementary info: shape, folding/unfolding, assembled state in solution

The instrumentation and sample prep



How to measure small angle X-ray scattering?

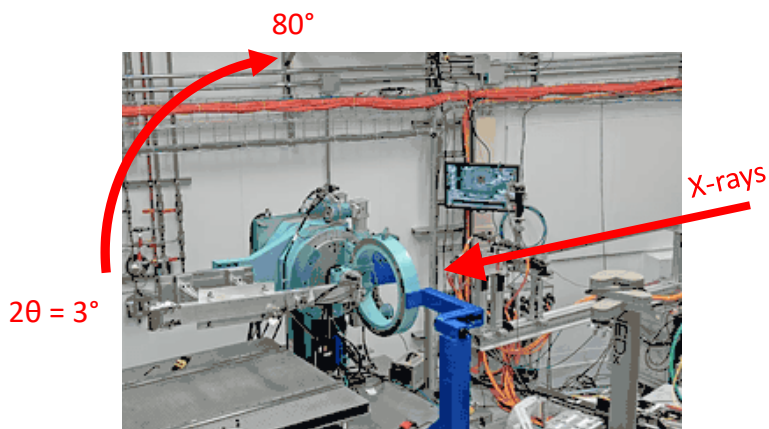
PXRD

SAXS

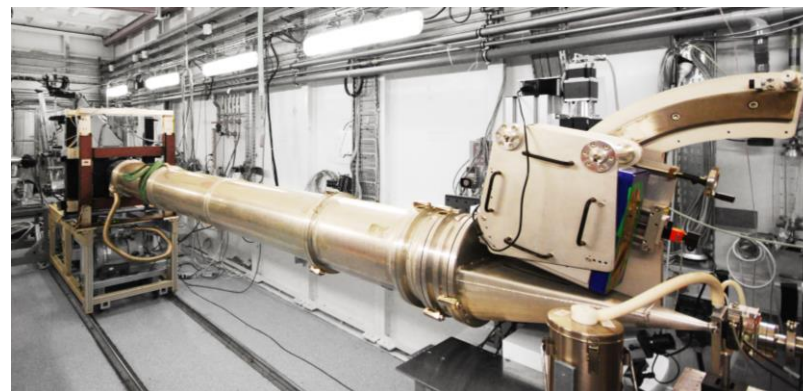
Lab source



Synchrotron



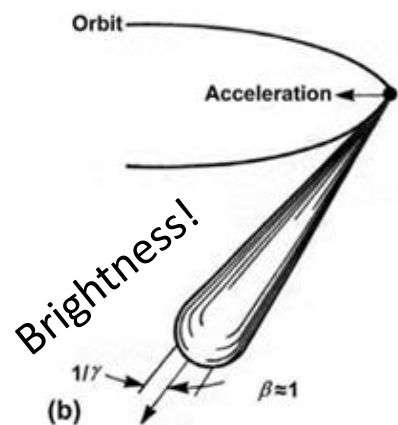
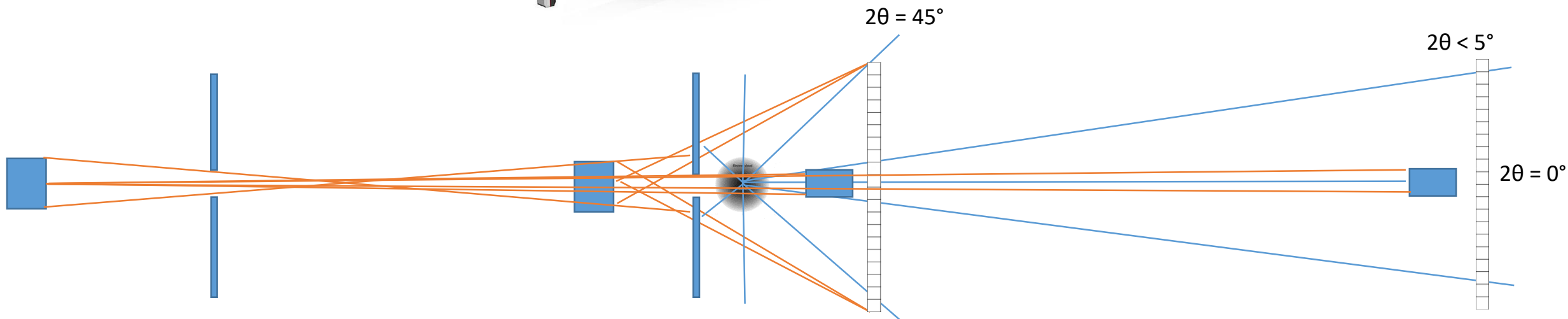
ESRF - BM26



- 1) Special instrumentation required for SAXS
- 2) SAXS instruments are long!



SAXS instrumentation basics



- More flux (better SNR), weakly scattering objects (organics/biology)
- Better q resolution
- Time resolved, SAXS mapping
- Choice of many wavelengths

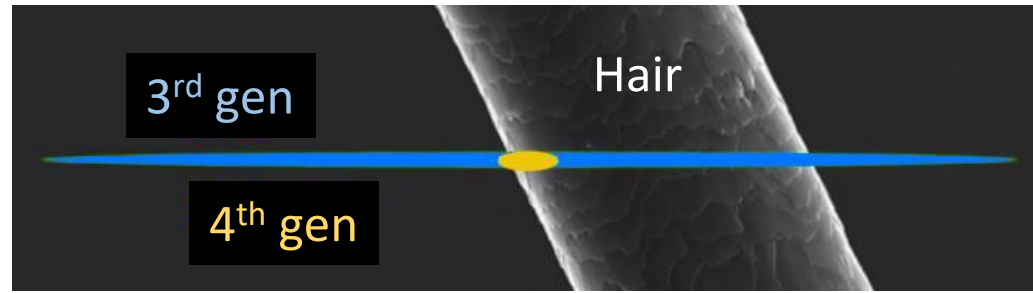
- Highly collimated hard X-ray beam (multiple slits)
- Large area detector with beamstop
- Space for >2 m sample to detector distance
- Some ability to change detector distance (SAXS/WAXS)
- All or most components in vacuum

State-of-the-art instrumentation

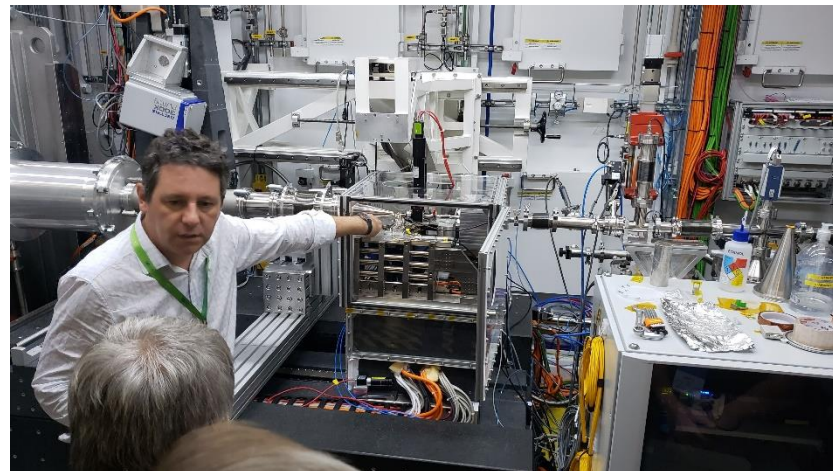
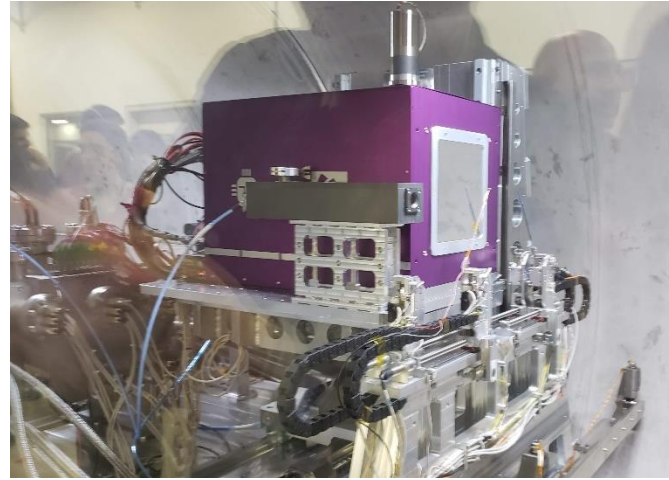
CATERETÊ beamline @ SIRIUS
Campinas, Brazil

Dedicated SAXS beamline, new for 2022

- 4th gen synchrotron, 88 m source to sample
- Modern large area detector with beamstop
- 0 - 28 m sample to detector distance
- All components in vacuum, no windows from source to detector



P. R. Willmott, 2022



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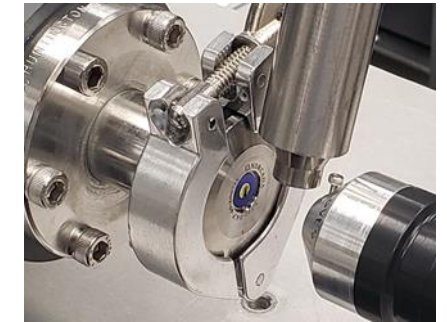
CLS SAXS instrument

1) Beam defining slit
+6.1 m
(1.00 mm x 1.00 mm)

2) Anti-scatter slit
+0.9 m
(0.25 mm V x 0.50 mm H)

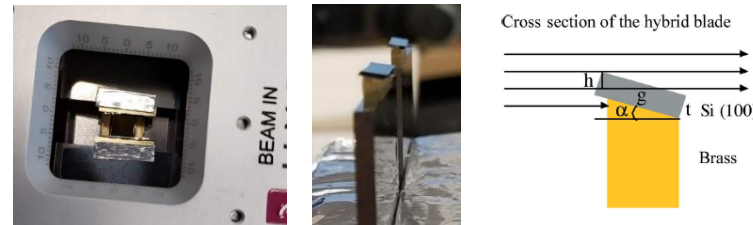
200 nm SiN window 3) Guard slit (0.80 mm)

~9 m long



Small air gap for sample (~35 mm)

low parasitic scatter



Y. Li et al., J. Appl. Cryst. (2008). 41, 1134–1139
N.M. Kirby et al., J. Appl. Cryst. 46, 1670–1680 (2013)

3 pinhole layout

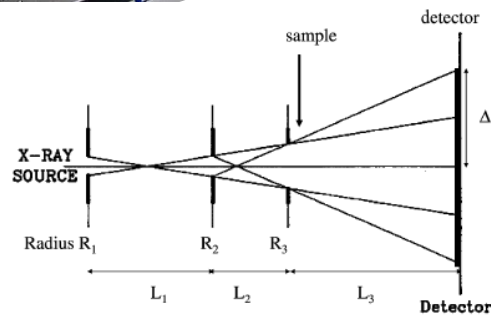


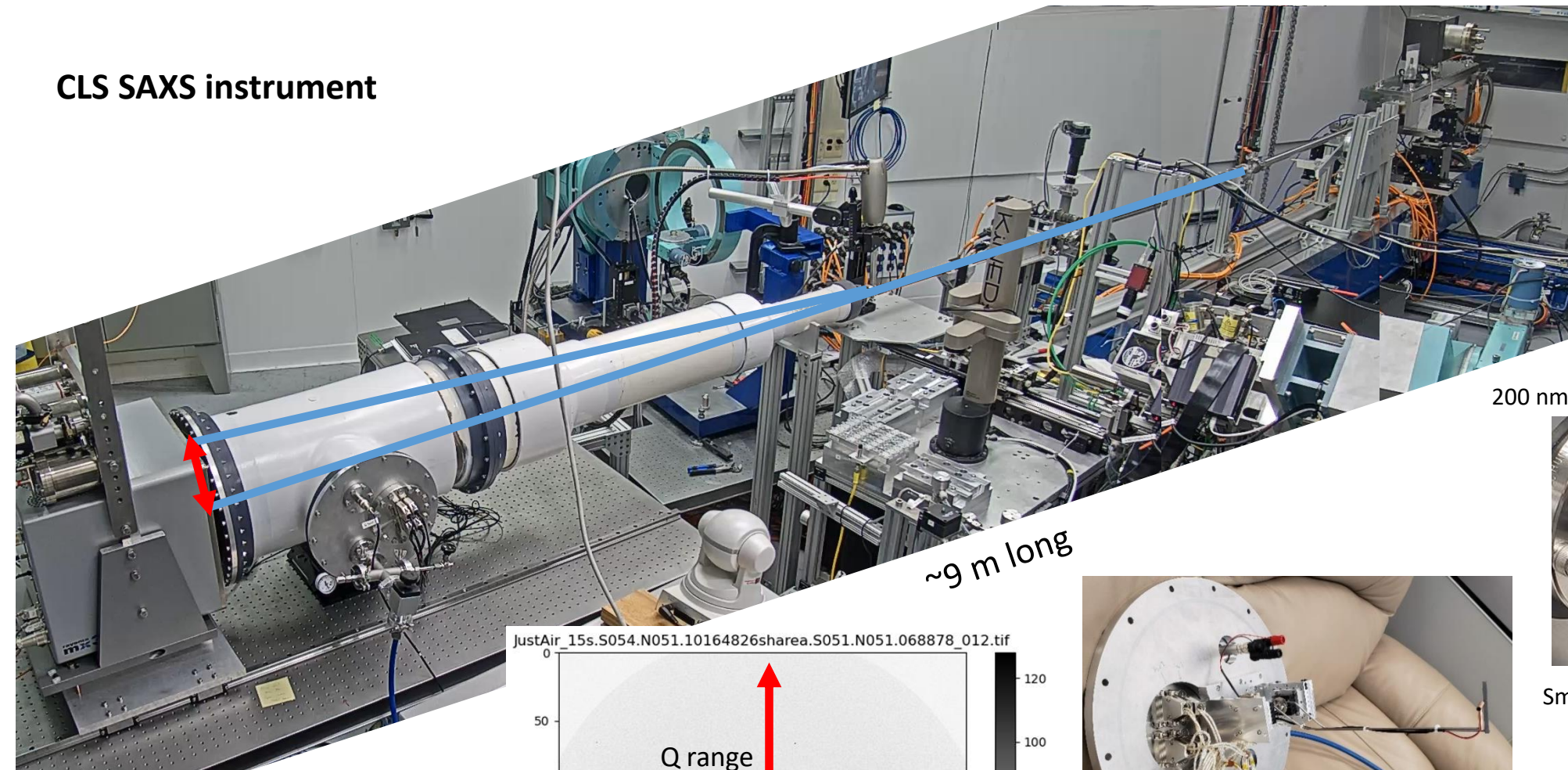
Figure 4
Schematic layout of a SAXS pinhole camera.

J. Appl. Cryst. (2004). 37, 369-380



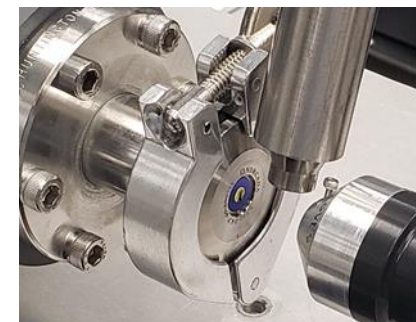
Canadian Light Source
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CLS SAXS instrument

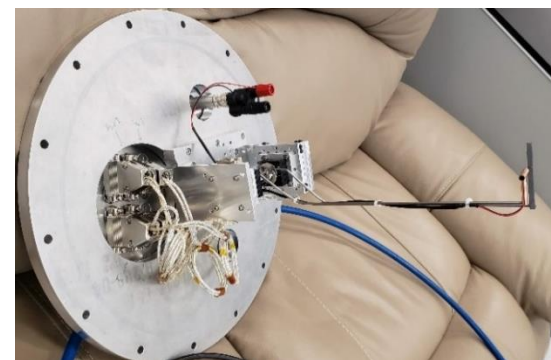


~9 m long

200 nm SiN window 3) Guard slit (0.80 mm)



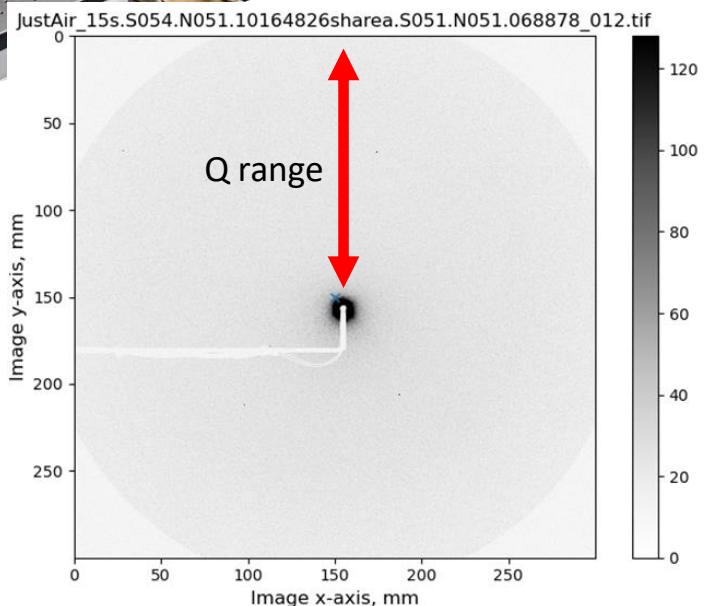
Small air gap for sample (~35 mm)



Vacuum tube ($\leq 10^{-2}$ Torr)
Beamstop with photodiode
-2.0 m
(4.0 mm)



13.5" Kapton window (125 μm)
~2100 pounds



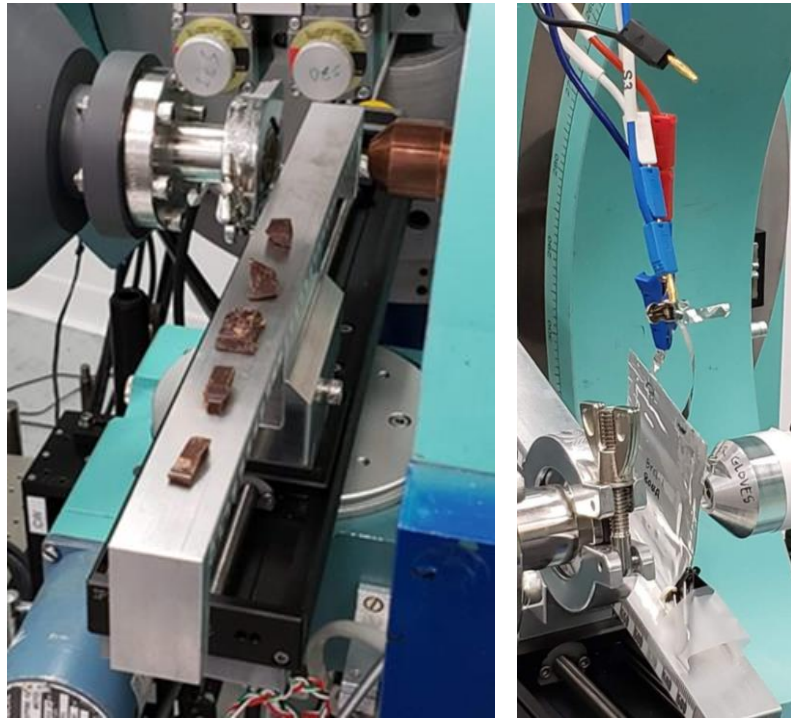
$\lambda = 1.26 \text{ \AA} \text{ (9.85 keV)}$
 $0.658 \text{ \AA} \text{ (18.8 keV)}$

$Q \sim 0.005 - 0.5 \text{ \AA}^{-1}$
 $d \sim 125 - 1.25 \text{ nm}$
 $2\theta \sim 0.05 - 3.7^\circ$

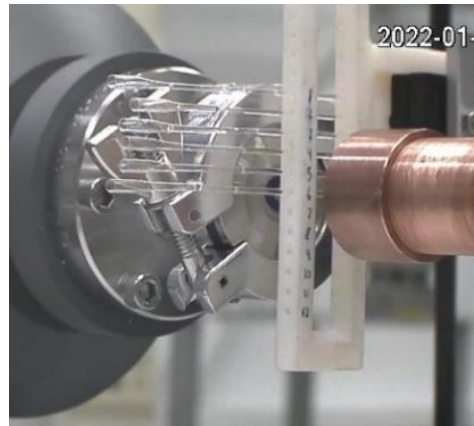
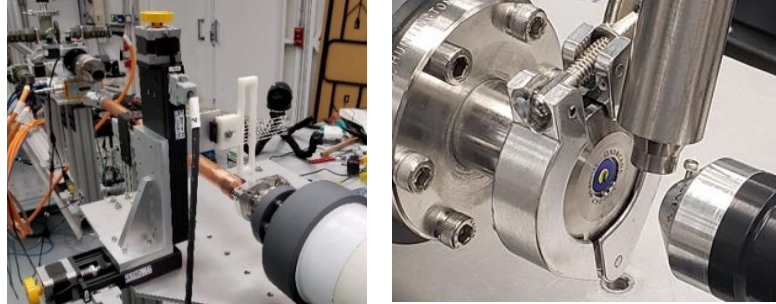
Sample preparation and mounting

Transmission

Freestanding,
Between Scotch tape



Capillaries: 1.0 to 1.5 mm diameter quartz,
borosilicate glass



-193 – +226°C
>1000 °C



Grazing incidence

Thin film sample on Si wafers,
or glass slides



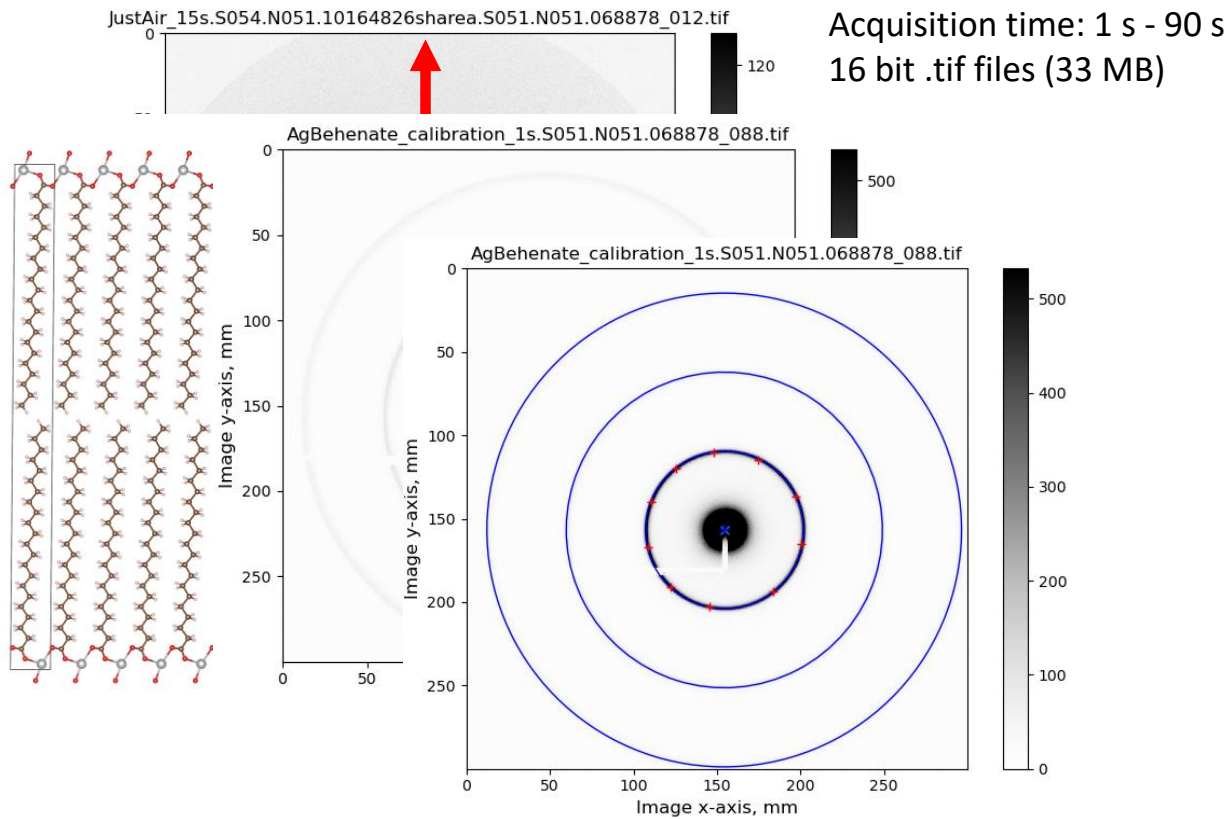
- Calculate the transmission of your sample... $1/e$ is ideal
- Bring blanks
- Bring a sample where you know what to expect
- Please buy your own capillaries

Data collection and reduction



Detector acquisition and calibration

During your beamtime, note the wavelength, the approximate detector distance, and detector details



Ring	q (Å ⁻¹)	d (nm)
1	0.1076	5.839
2	0.2152	2.920
3	0.3228	1.946
4	0.4304	1.460
5	0.5380	1.168

GSAS-II demo, "Calibration of an area detector"

GSAS-II project: <unnamed project>

File Data Calculate Import Export | Calibration Integration Parms | Help

Loaded Data:

- Notebook
- Controls
- Covariance
- Constraints
- Restrains
- Rigid bodies
- IMG JustAir_15s.S054.N051.10164826share.S051.N051.068878_012.tif
 - Comments
 - Image Controls
 - Masks
 - Stress/Strain
- IMG AgBehenate_calibration_1s.S051.N051.068878_088.tif
 - Comments
 - Image Controls
 - Masks
 - Stress/Strain

Image Controls:

Type of image data: SASD - small angle scattering data Color bar Greys Azimuth offset 0.0

Max intensity 532

Min intensity 0

Auto scaler ? Show line scan

Calibration coefficients:

- Beam center X 154.481
- Beam center Y 156.736
- Wavelength* 1.18178
- Distance 2335.831
- Tilt angle* -0.873
- Tilt rotation* 242.48

Integration coefficients:

- Bin style: Constant step bins in log(q) Pink beam source?
- Inner/Outer Q 0.464 4.494
- Start/End azimuth 0.0 360.0
- No. 2-theta/azimuth bins 2500 1
- Do full integration?
- Use for all new images?
- Azimuth at bin center?
- Apply sample absorption?
- Apply polarization? Value (0.001-0.999) 0.99

Dark image multiplier -1.0

Background image multiplier -1.0

Gain map

Calibration controls:

Calibrant Ag behenate Calib lines to skin Min calib denoising 16.0

Min ring l/lb 1.0

Calibration Integration Parms | Help

Copy Controls

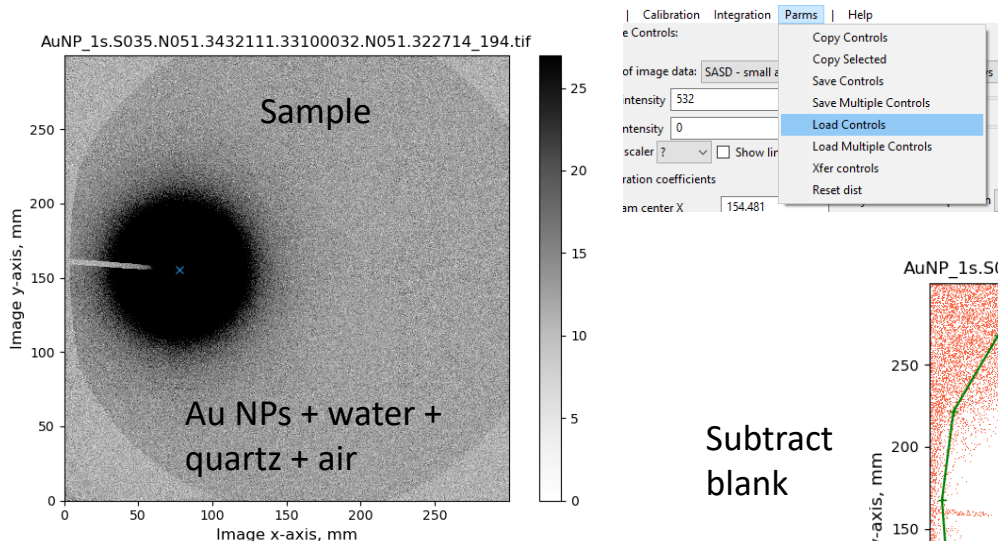
Copy Selected

Save Controls

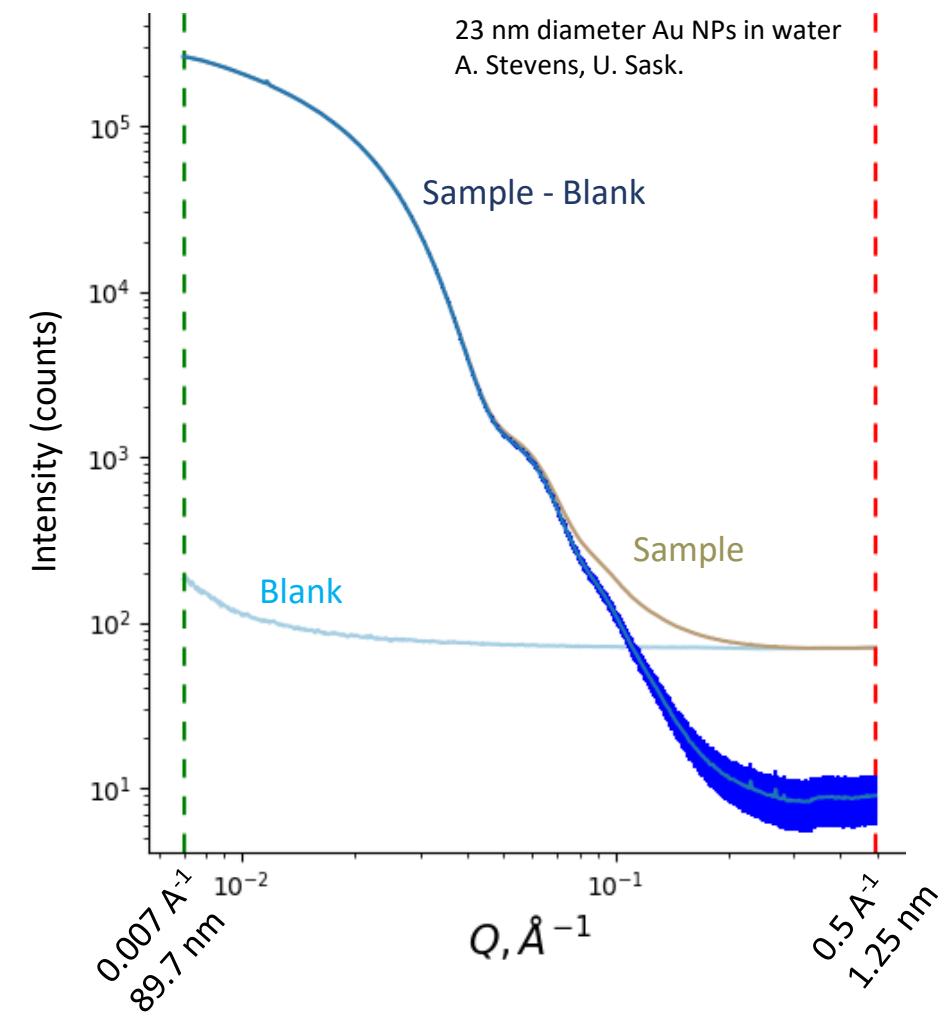
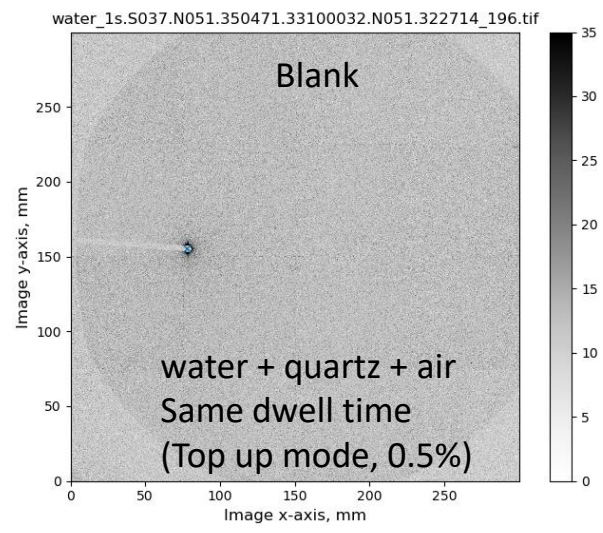
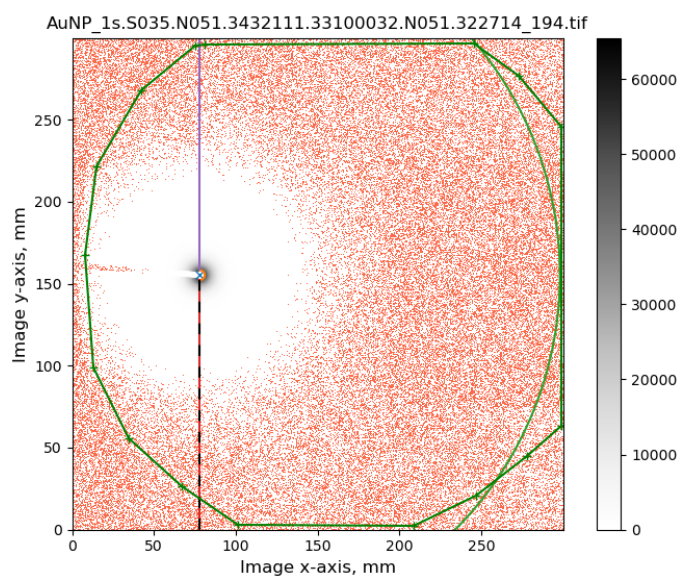
Save Multiple Controls



Data correction, and reduction to 1D plot



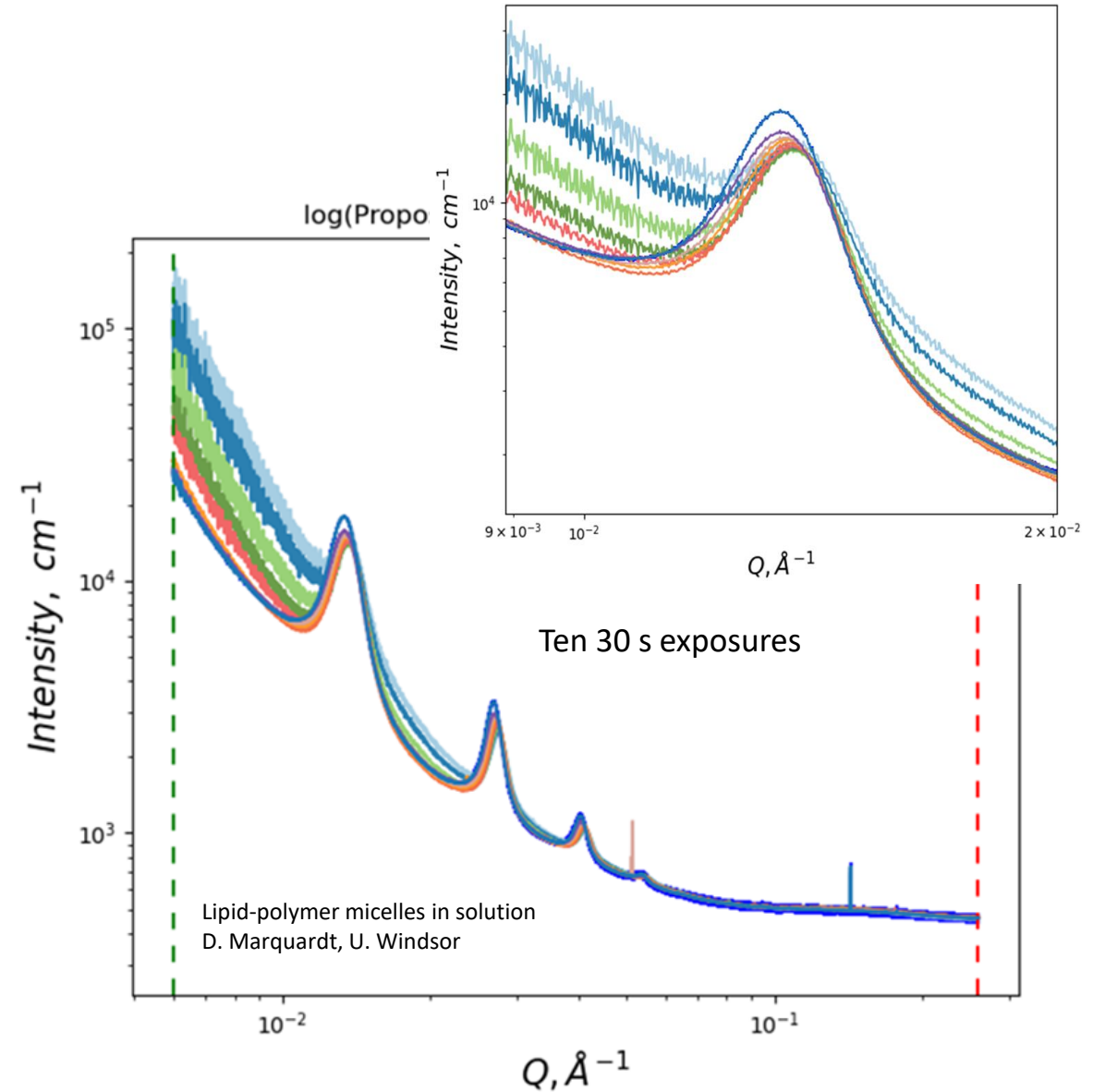
Subtract blank



Proper units of intensity are cm^{-1}
IF data are scaled to absolute intensity

5 tips for a great data collection experience

- 1) It's your experiment.
- 2) Get in contact ~2 weeks in advance.
- 3) Know what q range you want, and include it in your proposal. We can tune the endstation to your problem.
- 4) Know what your data should look like. Find published examples, or bring a known good sample.
- 5) **Chose quality over quantity:** Check for radiation damage at the start, work up data as you go along, many blank measurements, priority list



Data analysis with examples



Origins of the SAXS signal

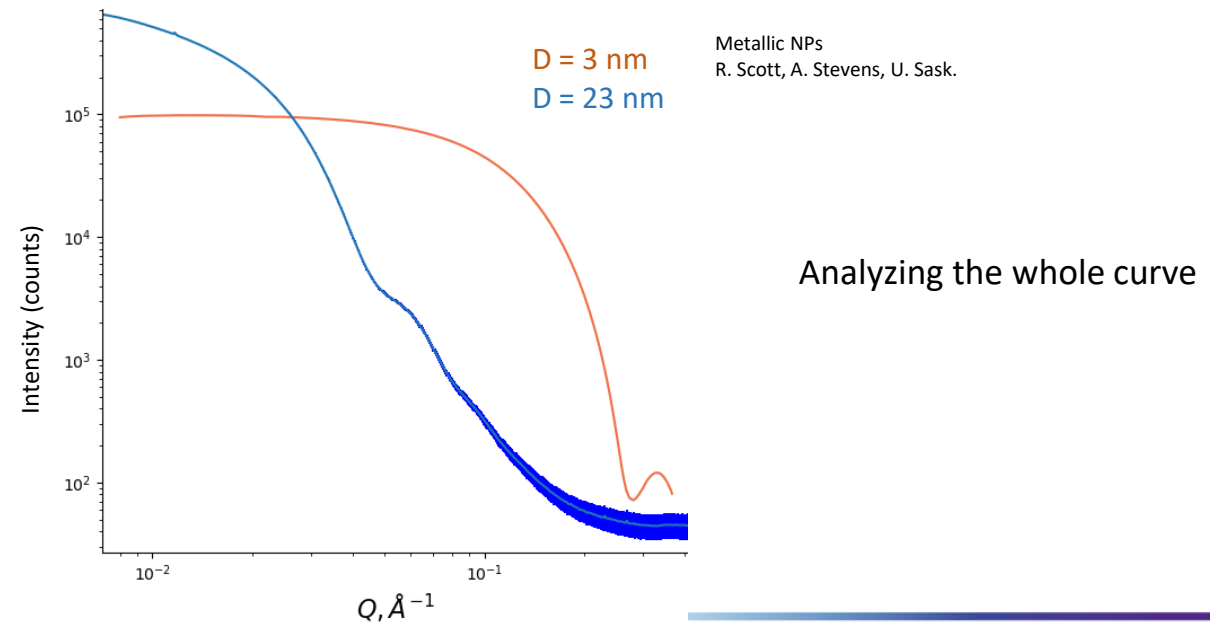
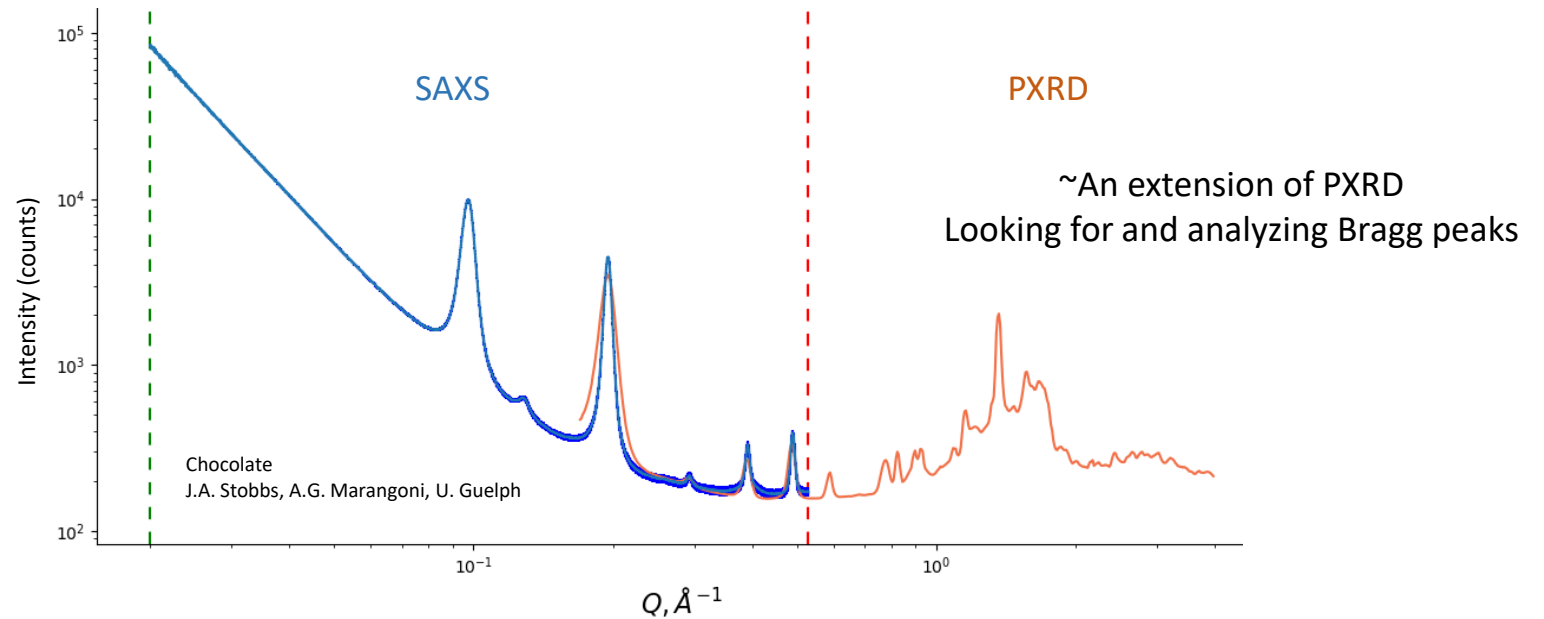
$$I(Q) = S(Q) \times P(Q)$$

S(Q) is the **structure** factor

- *Inter*-particle interferences
- High concentrations (>5% vol.)
- **Ordering/packing of particles**

P(Q) is the **form** factor

- *Intra*-particle interferences
- Low concentration (dilute limit)
- **Size/polydispersity/shape of particles**



Phospholipid reverse micelle formation in model cocoa butter: Evidence for in-situ seeding as a route to Form V polymorph in chocolate

J.A. Stobbs, S.M. Ghazani, E. Pensini, A.F.G. Leontowich, B. Barlow, and Alejandro G. Marangoni (in preparation)

Dept. Food Science
University of Guelph

The nanostructure of chocolate is manipulated using time and energy-intensive “tempering”

Guides crystallization of cocoa butter to “polymorph V”

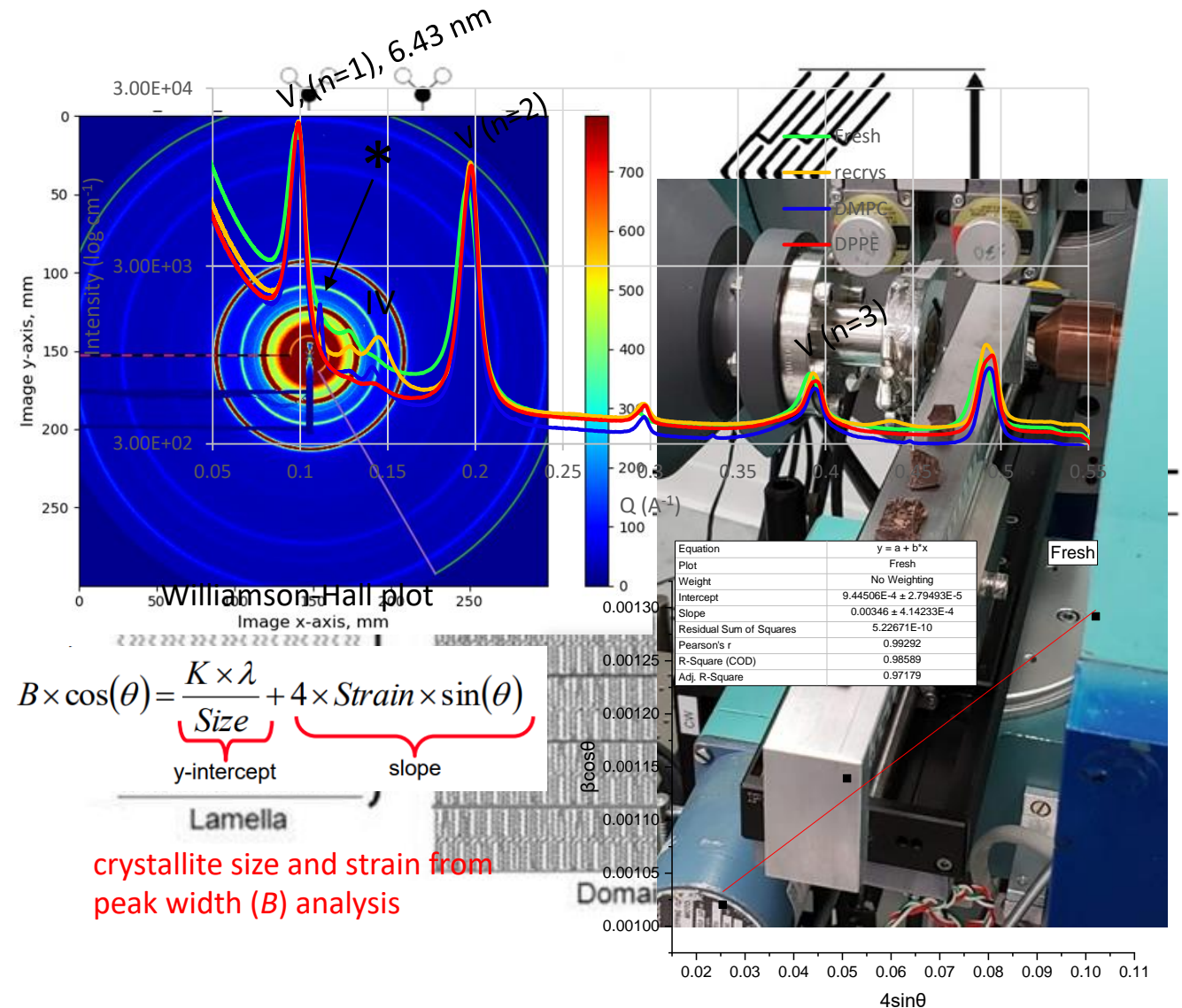


Tempering of cocoa butter and chocolate using minor lipidic components

Jay Chen¹, Saeed M. Ghazani¹, Jarvis A. Stobbs^{1,2} & Alejandro G. Marangoni^{1,2*}

NATURE COMMUNICATIONS | (2021)12:5018

Why does it work?



crystallite size and strain from peak width (B) analysis

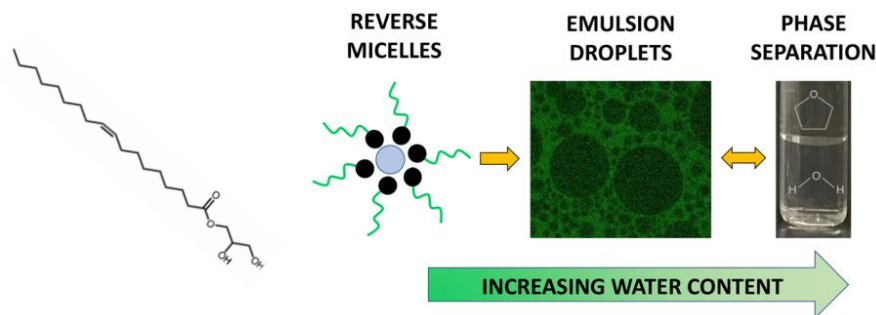
L. Earnden, A.G. Marangoni, T. Laredo, J. Stobbs, Erica Pensini

Scientific Reports **12**, 15832 (2022)

J. Molecular Liquids **367**, 120551 (2022)

Physics of Fluids **34**, 097119 (2022)

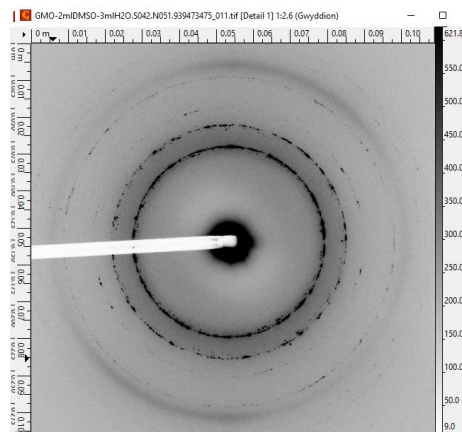
Separating solvents (THF, DMSO, DMF and acetonitrile) and metals from waste water using amphiphiles, emulsifiers and surfactants



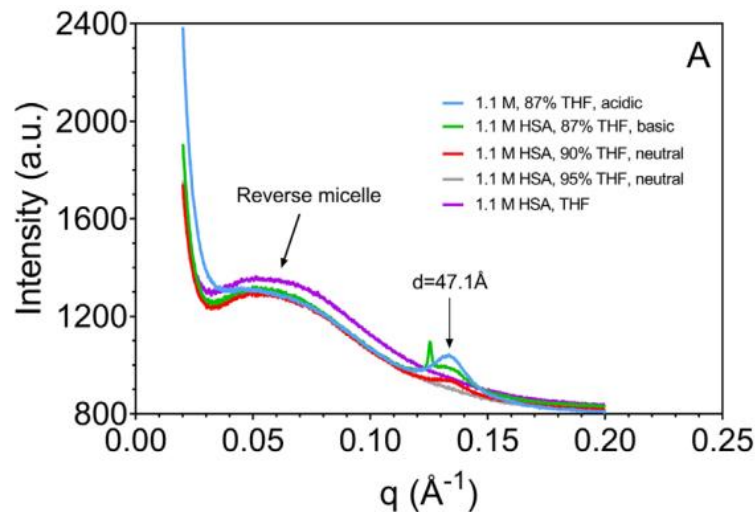
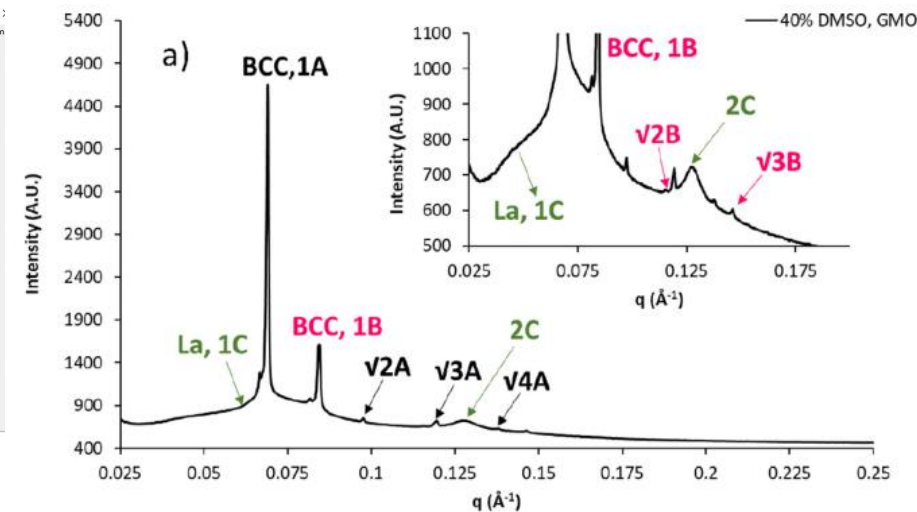
Mechanism of solvent separation?

1.1 M HSA, 87% THF, acidic pH				
		ξ (Å)	d (Å)	d/x
a_2	1.158	19.5	86.0	4.4
c_1	-98.92			
c_2	18.245			

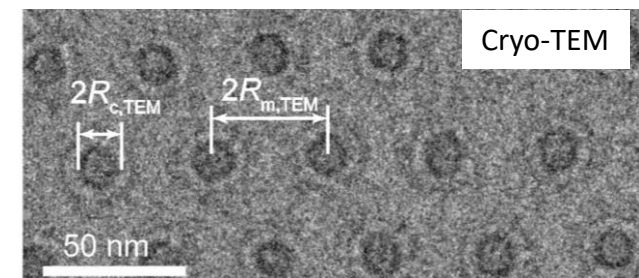
Powder diffraction ... ??



Symmetry	Peak position ratio
BCC (Im3m)	v2, v3, v4 ...



Reverse micelle lattice

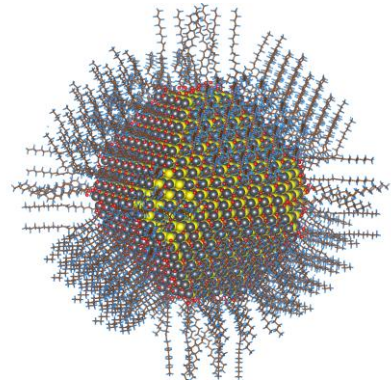


L. Chen et al., *PNAS* **115**, 7218-7223 (2018)



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Energy materials: Quantum dot solar cells



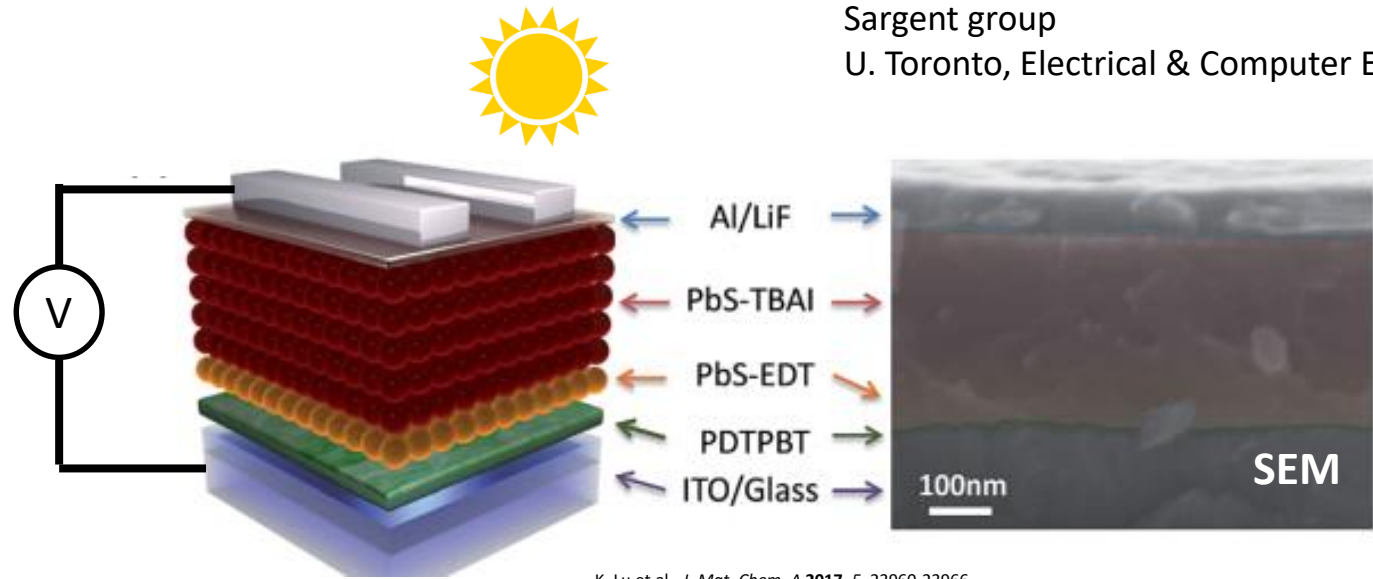
PbS
CsPb(Br/I)₃
InAs

<https://www.olcf.ornl.gov/2015/05/05/demystifying-quantum-dot-conundrums/>

Efficient exciton generation
Size-based, tune-able band gap

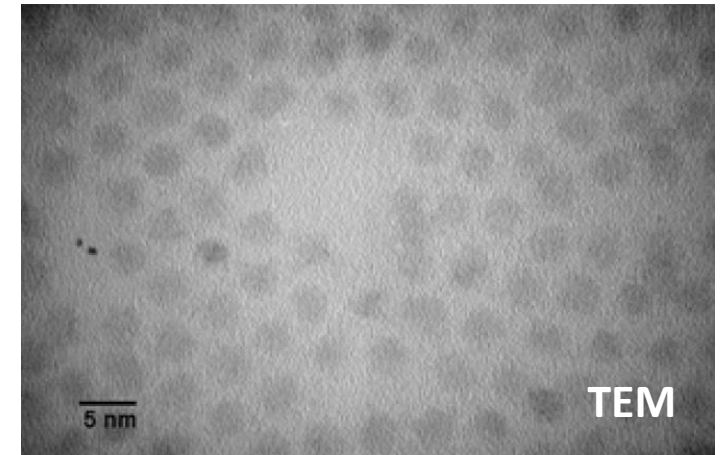
- 18.1% record efficiency in 2022
Performance improved by tuning:
- Particle size distribution or polydispersity
 - How particles pack and the interparticle distance

- Control the size and packing by:
- Ligand exchanges cycles, centrifuge to remove largest QDs
 - Rinsing steps during buildup
 - Spin speeds, solvents, additives

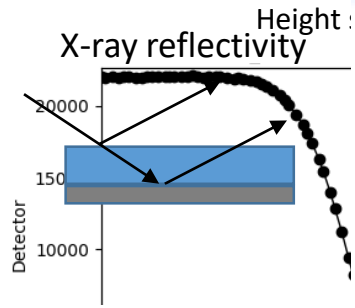


K. Lu et al., *J. Mat. Chem. A* 2017, 5, 23960-23966

QD films deposited layer-by-layer using spin coating
Top electrodes deposited using thermal evaporation

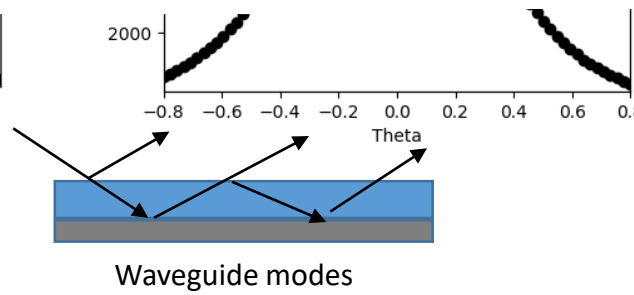
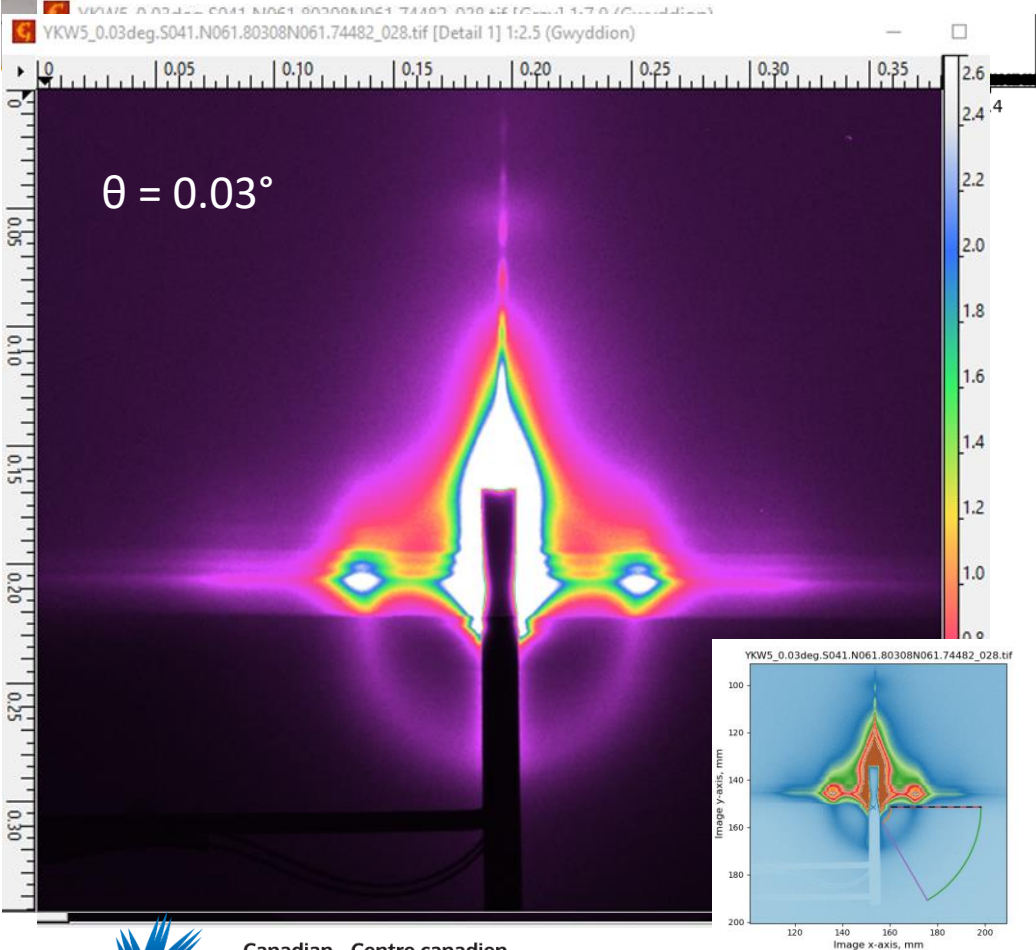
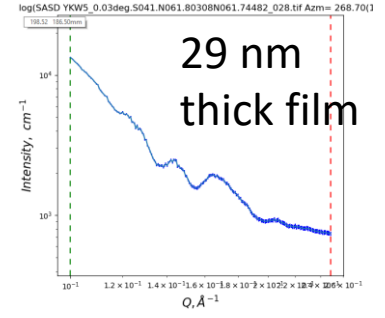
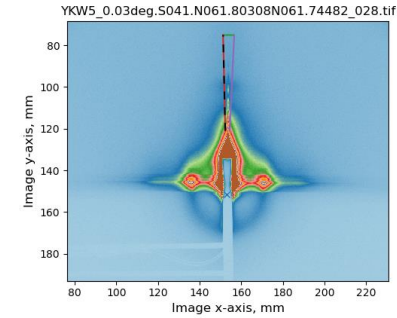
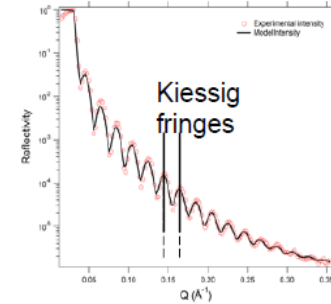


M. Yuan et al., *Adv. Mater.* 2014, 26, 3513-3519



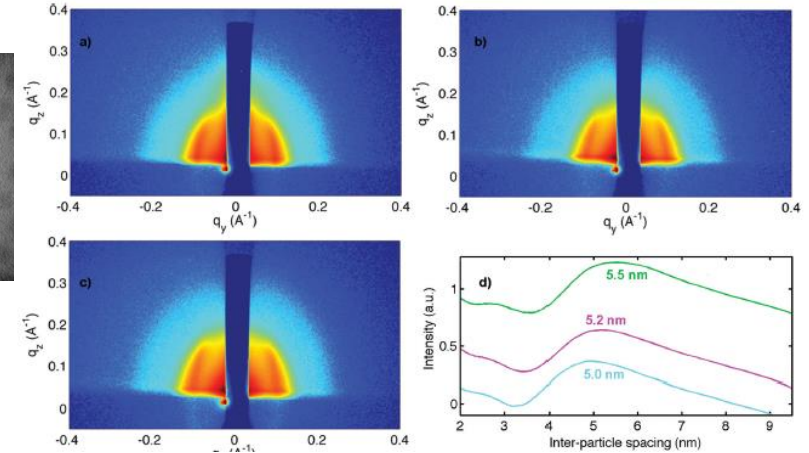
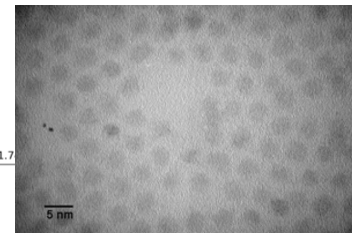
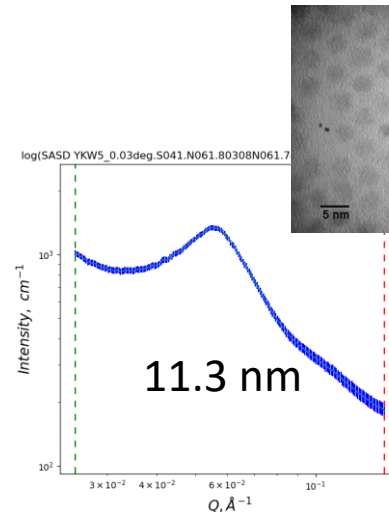
Simple estimation of film thickness

Fringes with uniform spacing
 Thickness of the layer : $t = \frac{2\pi}{\Delta q_z}$



- Indicates very flat, high quality film
- Adjusting grazing angle can give depth information

- Reflectivity gives film thickness
- Position gives interparticle distance
- Peak width gives size distribution
- Average over relatively huge area



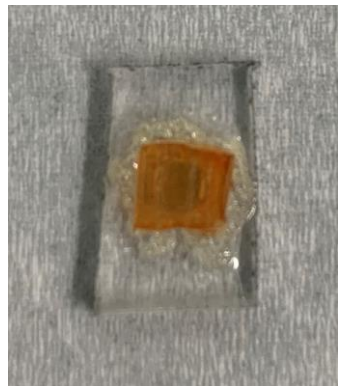
M. Yuan et al., Adv. Mater. 2014, 26, 3513–3519

$$I(Q) = S(Q) \times P(Q)$$

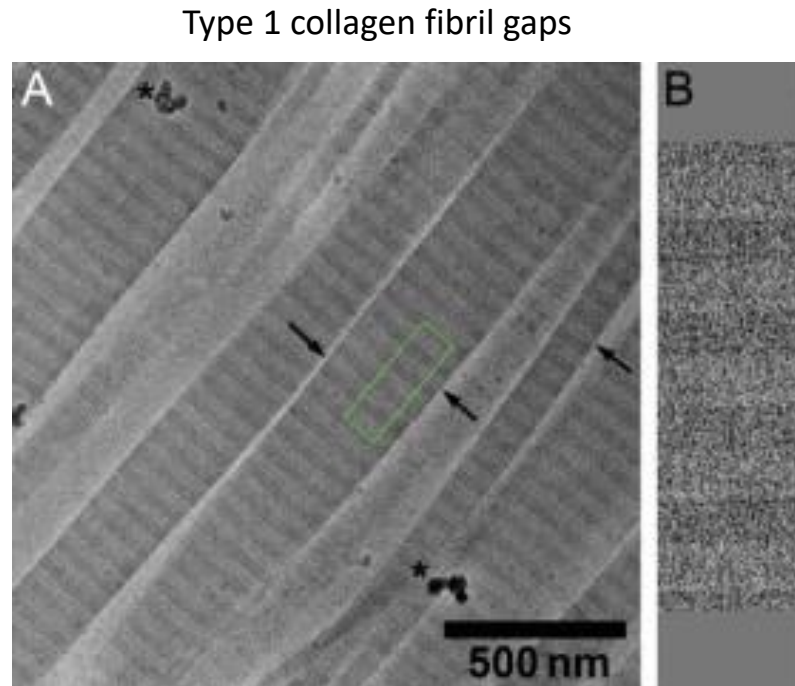
S(Q) is the **structure** factor

- *Inter*-particle interferences
- High concentrations (>5% vol.)
- Ordering/packing of particles

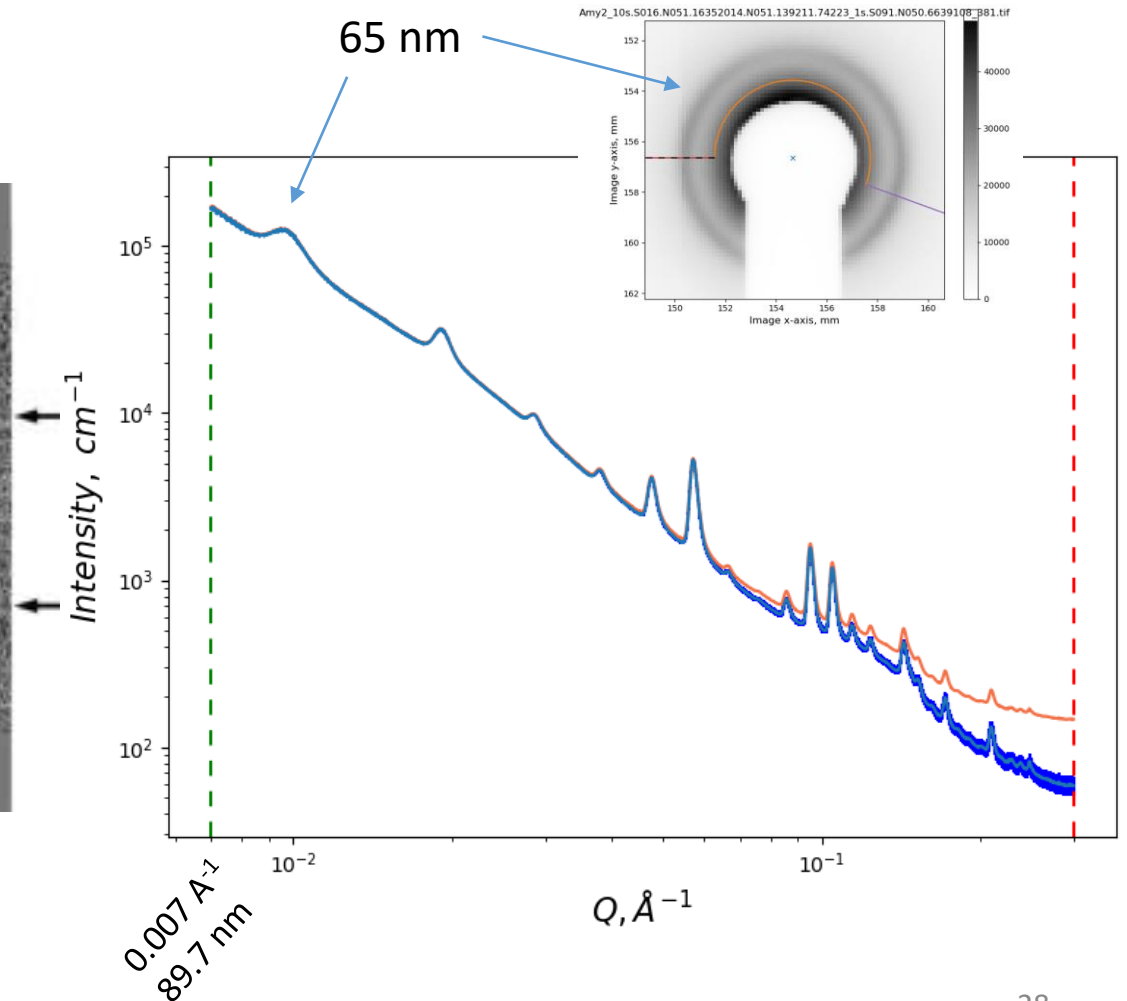
~An extension of PXRD
Looking for and analyzing Bragg peaks



Sea cucumber dermis
M. Harrington, McGill



<https://doi.org/10.1016/B978-0-12-416617-2.00009-6>

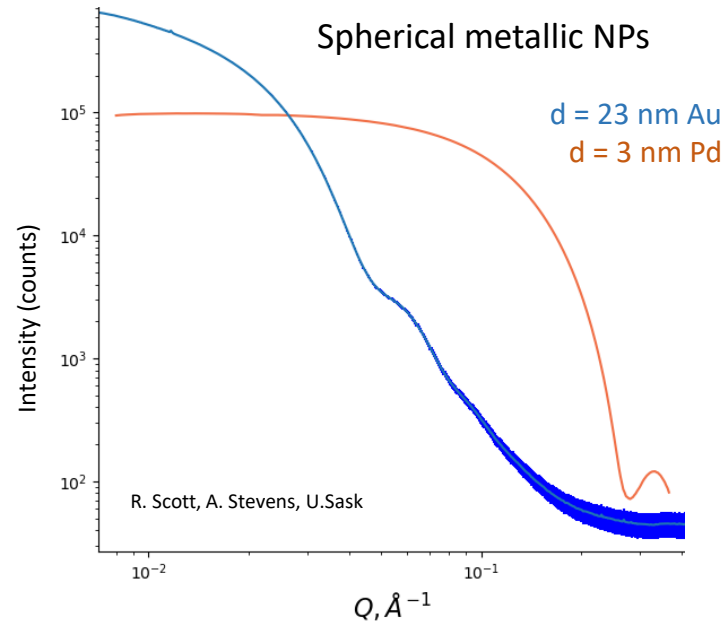


Origins of the SAXS signal

$$I(Q) = S(Q) \times P(Q)$$

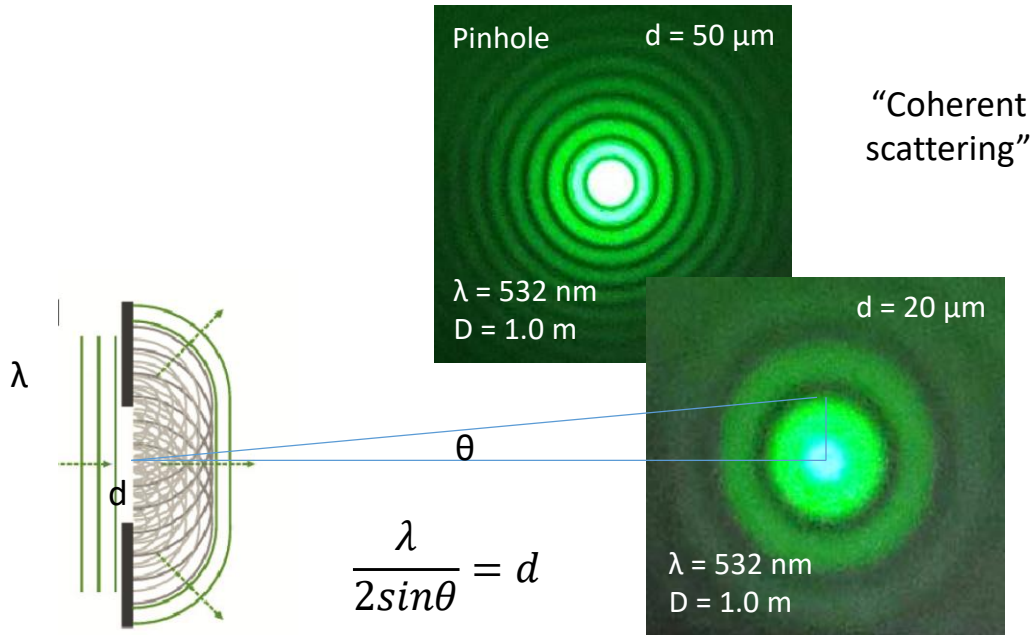
P(Q) is the **form factor**

- *Intra*-particle interferences
- Low concentration (dilute limit)
- **Size, shape, polydispersity of particles**



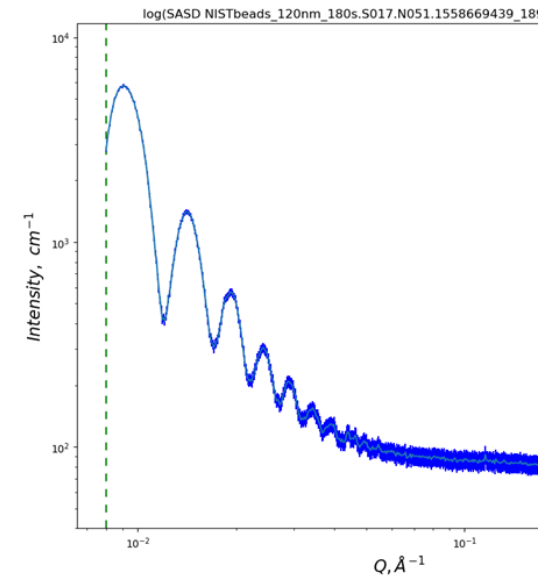
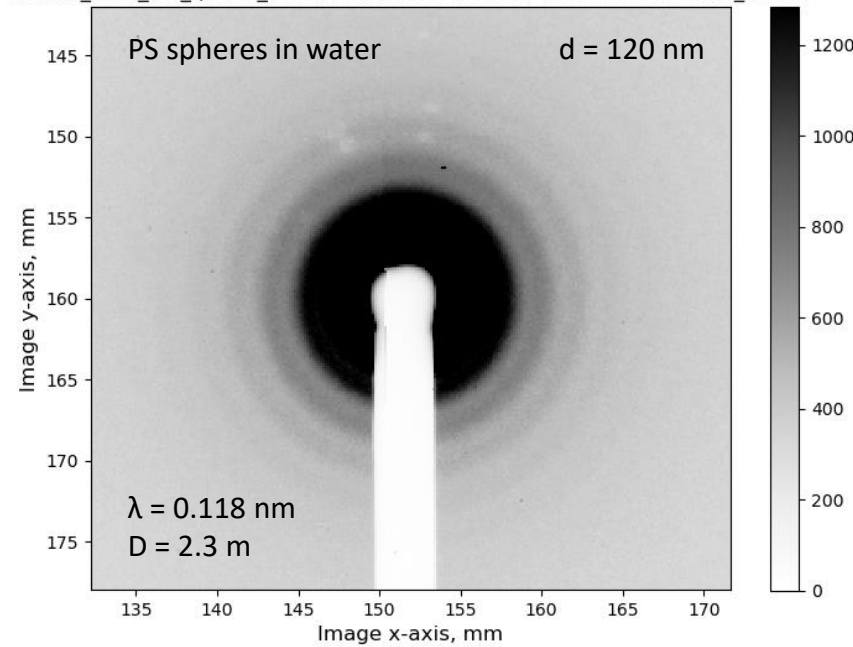
Sample prep is critical !!!

1. Dilute (~0.5 – 10 mg/mL)
no interparticle interactions
2. Pure
3. Monodisperse



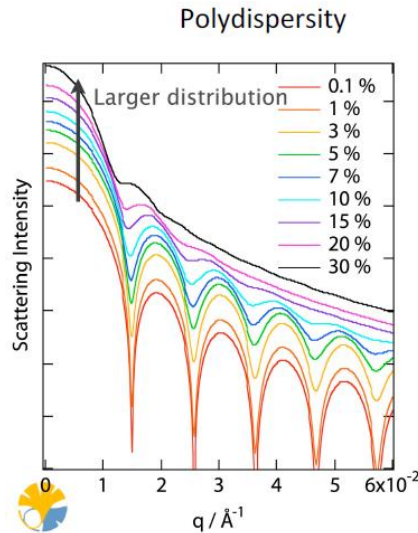
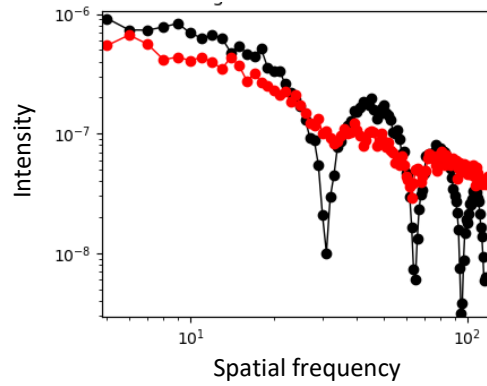
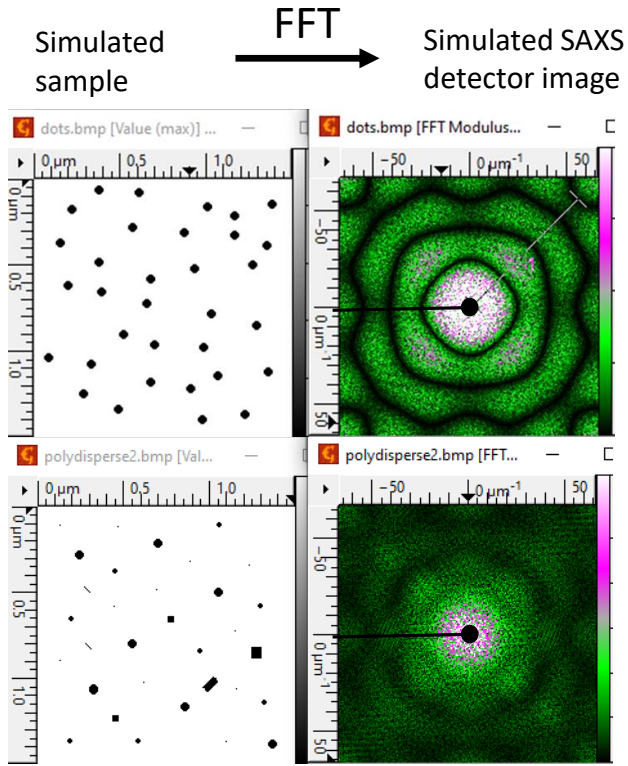
“Coherent scattering”

120nm_NIST_std_quartz_300s.S014.N051.4056907486906665264173_365.tif



But there are limits!

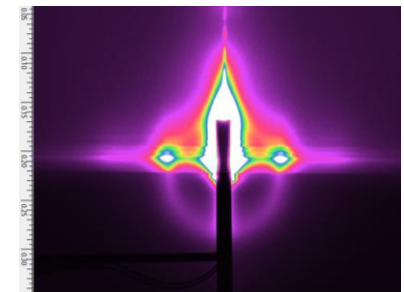
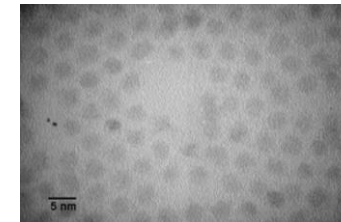
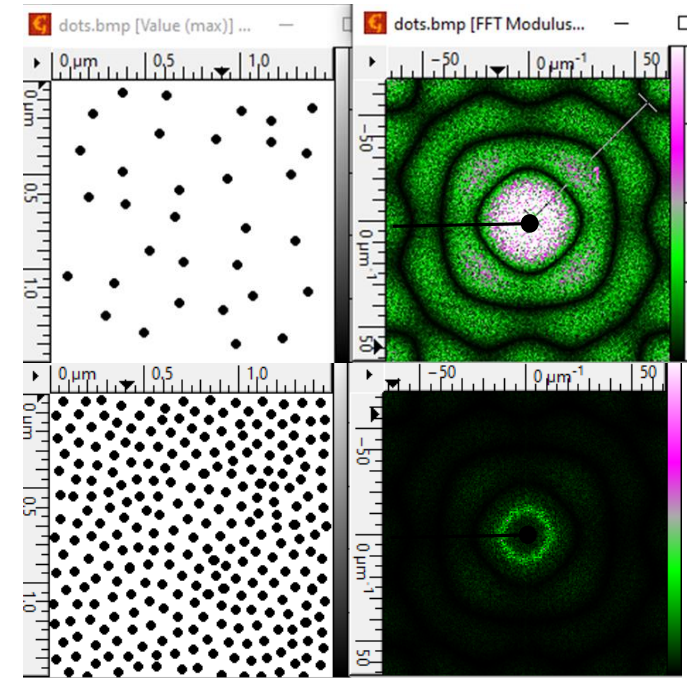
Solutions must be monodisperse, and pure



Form factor oscillations smear out when size polydispersity is $\geq 15\%$

Solutions must be dilute: No interparticle effects

$\sim 1 - 10$ mg/mL... Bring many concentrations

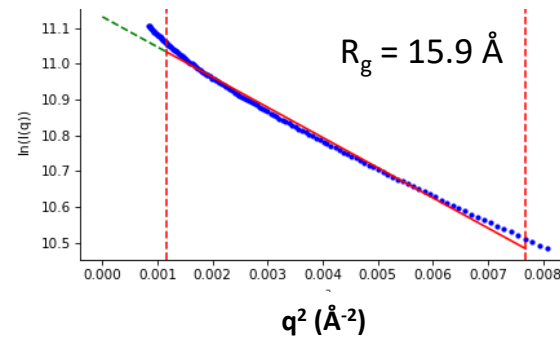
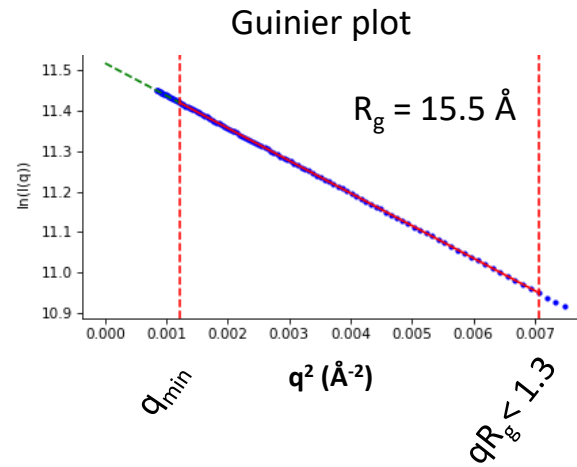
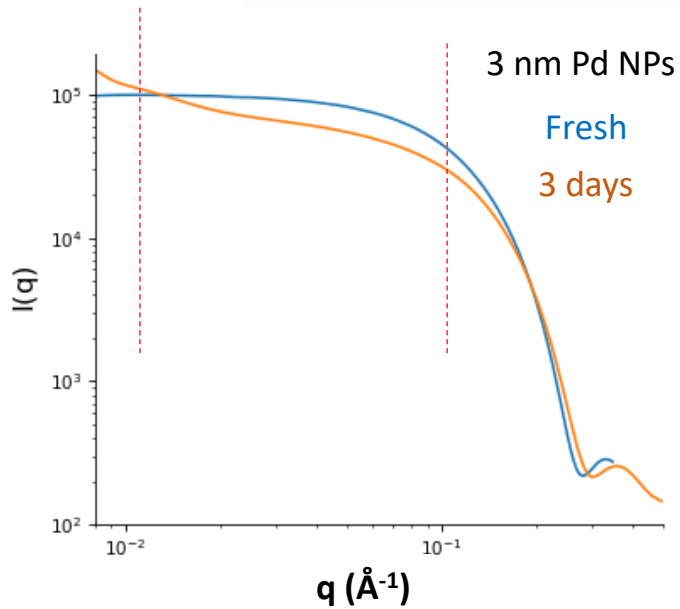


M. Yuan et al., Adv. Mater. 2014, 26, 3513–3519

Form factor data analysis: Catalytic nanoparticles

Guinier's approximation: $I(q) \approx I(0)e^{-q^2 R_g^2/3}$

R_g : Radius of gyration
 $I(0)$: Intensity at $q=0$



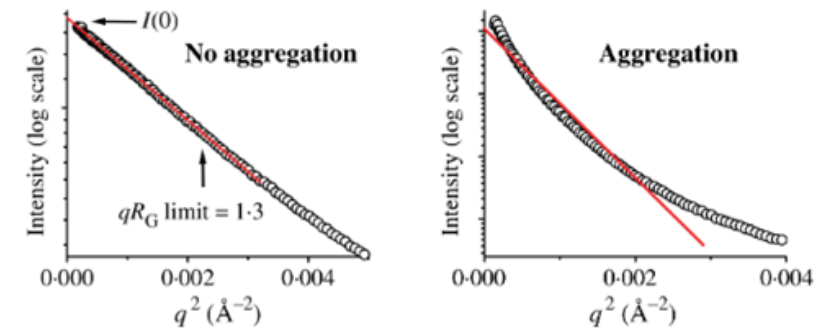
R_g is the square root of the average squared distance of each scatterer from the particle center

RMS distance of the objects parts from its center of mass

➤ Particle radius

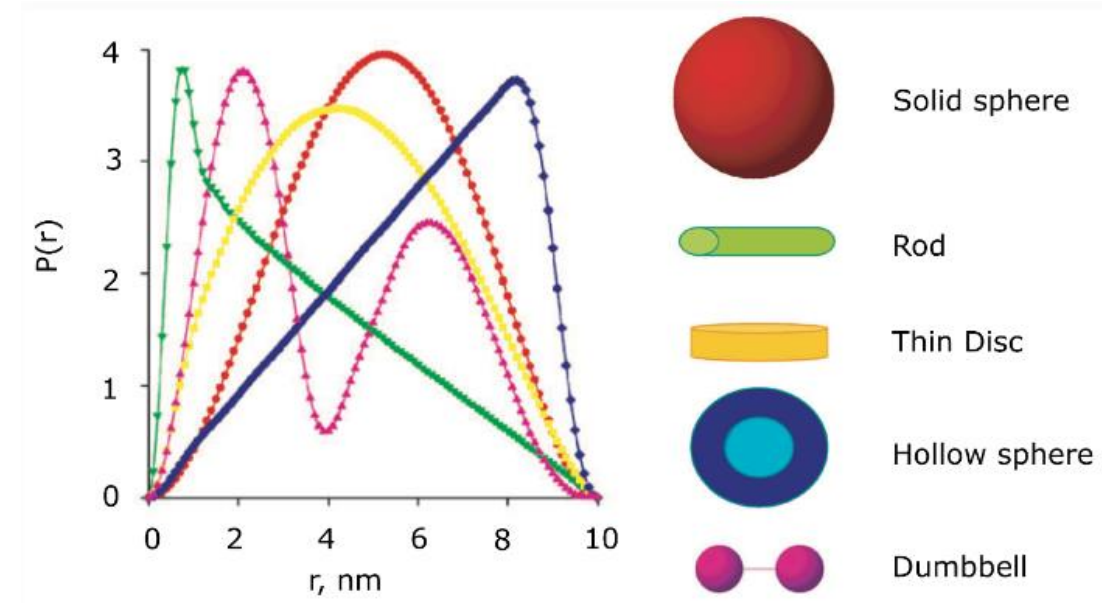
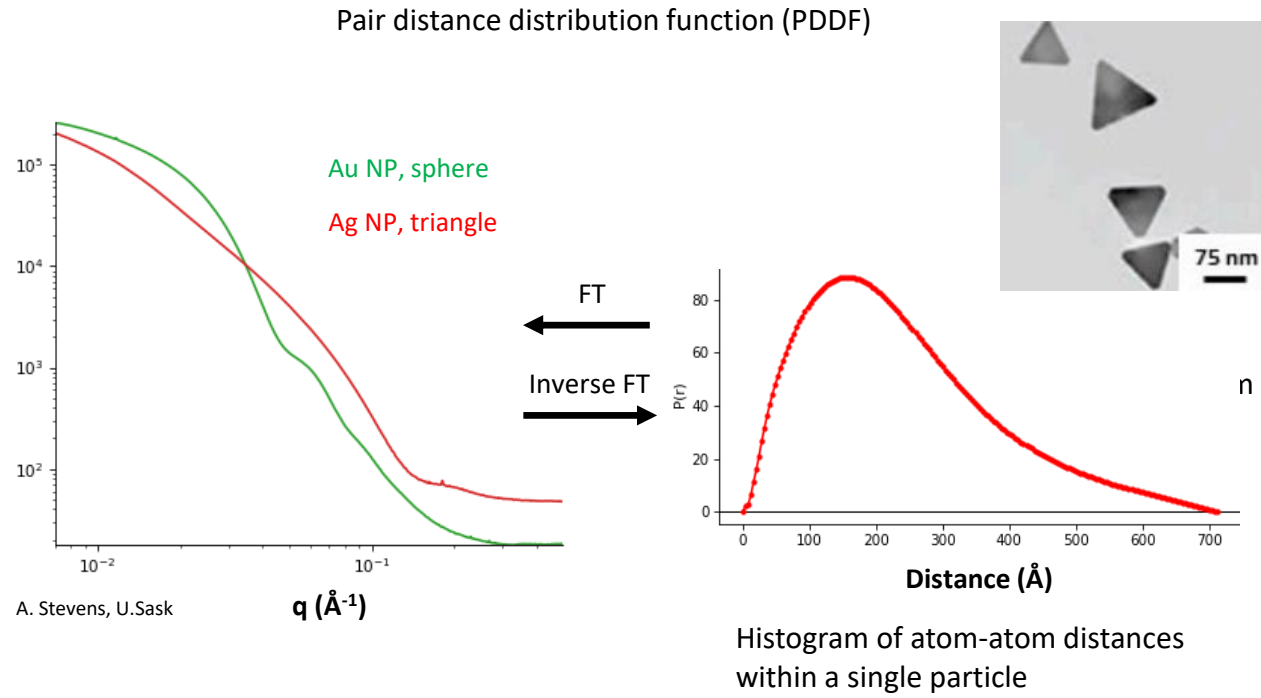
Require no assumption about the shape or internal structure of the particle

➤ Sample quality: non-linearity can indicate aggregation, repulsion, radiation damage



D. Putnam et al. Quart. Rev. Biophys. 40, 191-285 (2007)

Form factor data analysis: Catalytic nanoparticles



D. Svergun, M.H.J. Koch, Rep. Prog. Phys. 66 1735 (2003)

D_{max} - Maximum dimension of the particle

Shape of the $P(r)$ function is related to the shape of the particle, with low spatial resolution

Optimized Chitosan-Based Nanoemulsion Improves Luteolin Release

C. Diedrich, I.C. Zittlau, N.M. Khalil, A.F.G. Leontowich, R.A. de Freitas, Ildiko Badea, R.M. Mainardes

Pharmaceutics 15, 1592 (2023)

College of Pharmacy and Nutrition
U. Saskatchewan

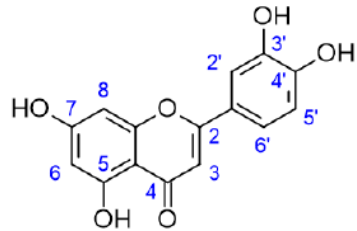
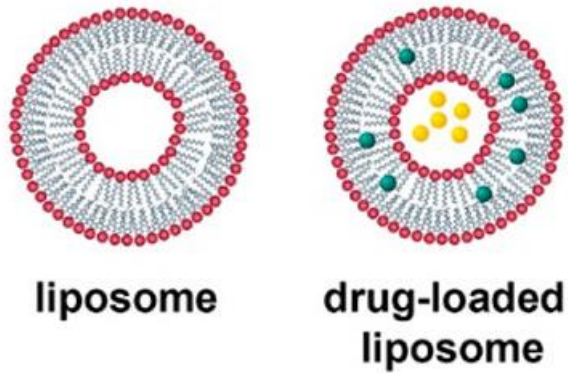


Figure 1. Luteolin structure.

Anti-oxidant, -inflammatory, -tumor, -viral
Poor absorption after oral administration, limited water solubility

Nano-encapsulation might improve the solubility of luteolin?

Diameter: 20 – 200 nm
Bilayer: ~5 nm



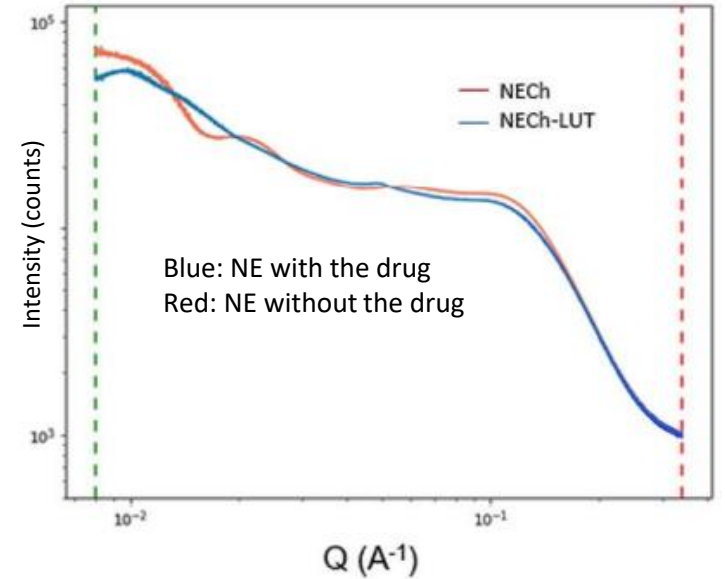
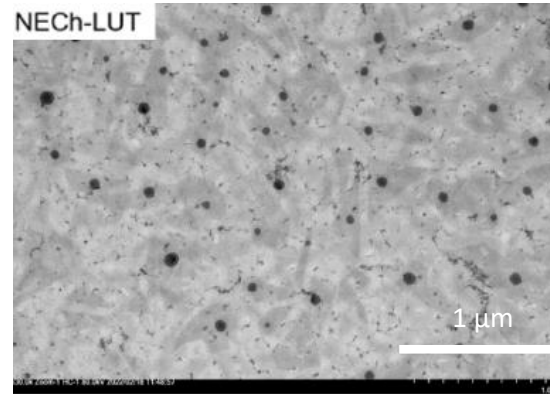
liposome

drug-loaded liposome

Delivery vehicle of the COVID-19 mRNA vaccines by Moderna and Pfizer/BioNTech

Oil phase: Luteolin, oleic acid, with ethylene glycol and Tween 20 as surfactants
Aqueous: Chitosan solubilized in 0.25% acetic acid.

Drop aqueous into oil while sonicating



The most reasonable fit was obtained modeling two spherical droplets

$d = 27.0 \pm 2.6$ nm
 $d = 1.4 \pm 0.26$ nm

Liposomes
Tween 20 micelles

Where is luteolin?

1. Dilute (~0.5 – 10 mg/mL), no interparticle interactions
2. Pure
3. Monodisperse

Sample prep is critical !!!



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Proteins in solution

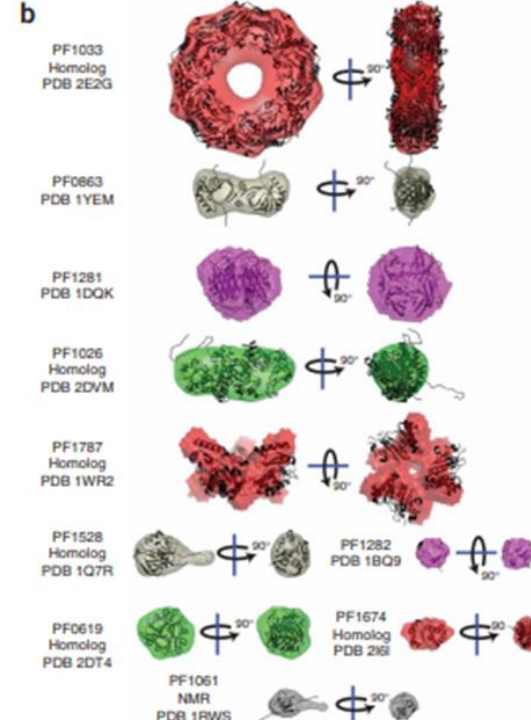
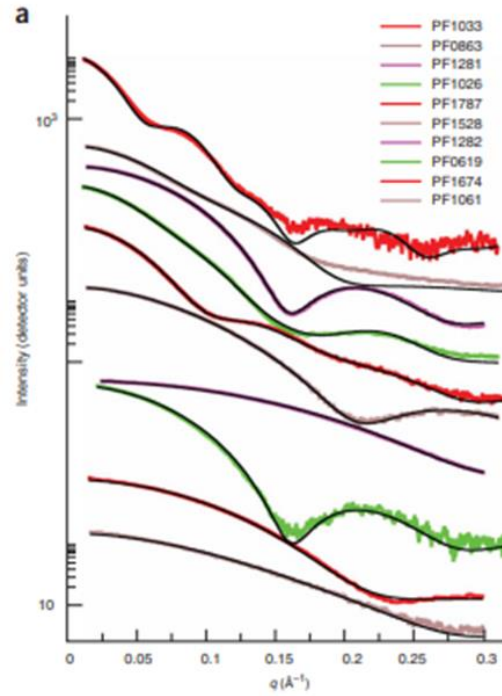
Aid structure determination

- Size and low resolution shape
- Molecular weight (>10%) to verify the oligomeric state

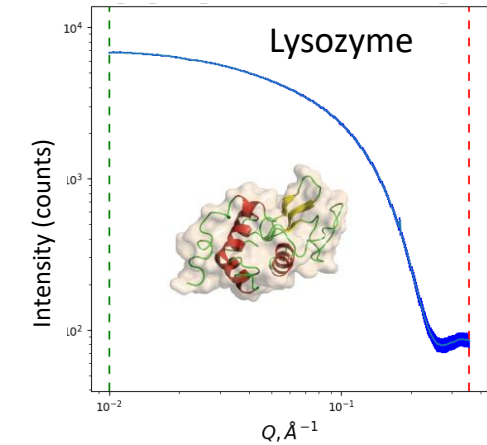
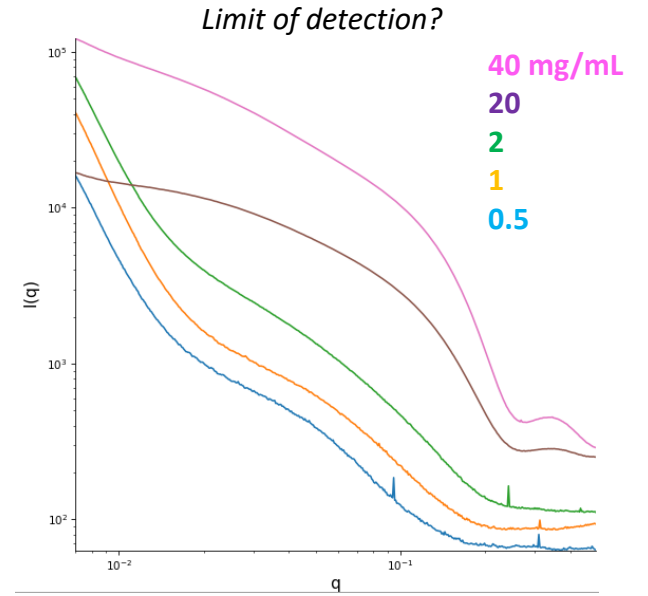
Studies of crystallization (nucleation vs. precipitation)

Challenging !!

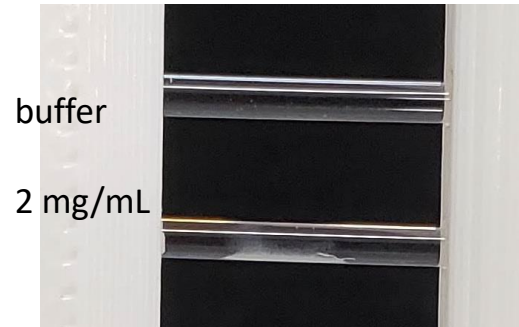
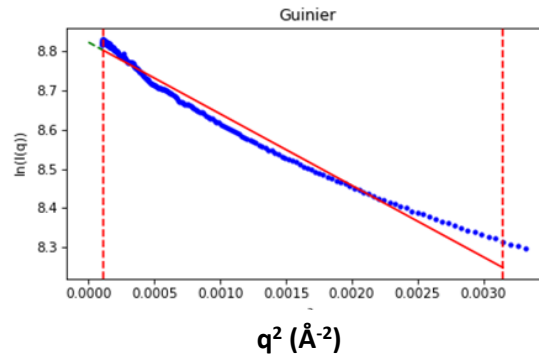
- Limited amounts of material
- Weakly scattering
- Radiation damage



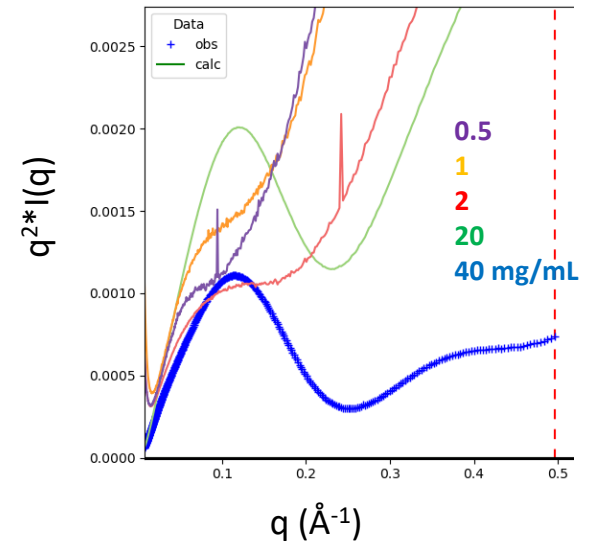
G.L. Hura et al., *Nature Methods* 6(8), 606 (2009)



April 2023



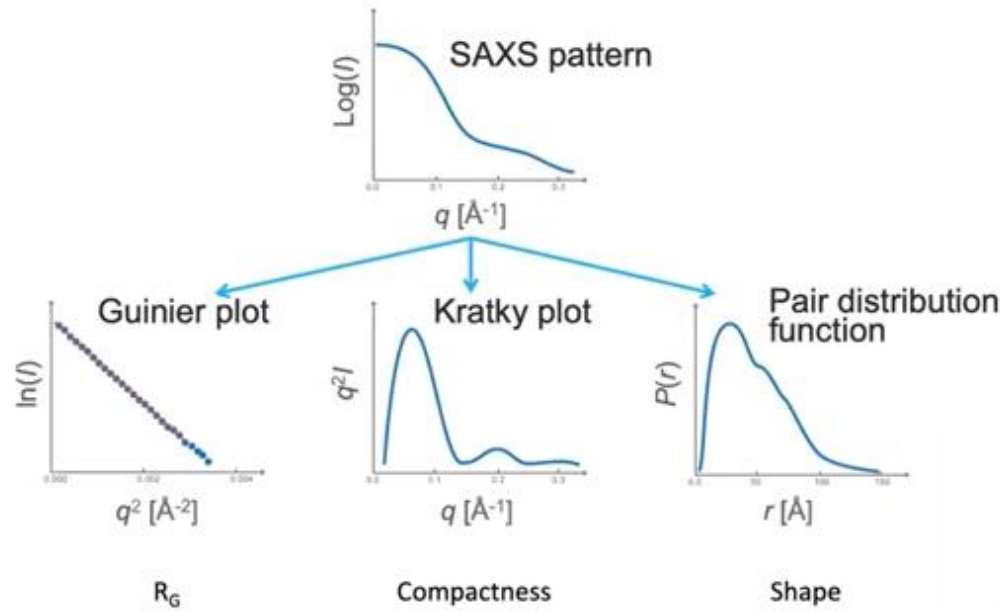
Kratky plots (degree of unfolding)



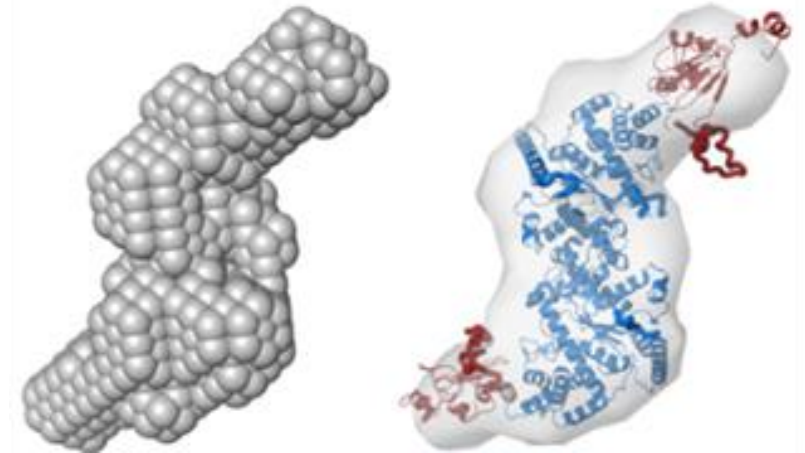
$$I(Q) = S(Q) \times P(Q)$$

P(Q) is the **form** factor

- *Intra*-particle interferences
- Low concentration (dilute limit)
- Size/polydispersity/shape of particles



These provide boundary conditions for modelling programs



ATSAS package
DAMMIF
Irena

SAXS combined with crystallography and computation 263

Table 5. SAXS parameters for data validation and interpretation

Parameters	Assessment
Experimental <i>q</i> -range	Range must be suitable through the entire spatial resolution required for determined models
Guinier plot R_G	Non-linear behavior indicates aggregation or inappropriate <i>q</i> -range Consistency of extracted R_G with multiple methods (Table 1) increases confidence in not only R_G but also assigned D_{max}
$I(0)$	Should correlate with molecular weight and concentration
D_{max}	Proper description of the range of D_{max} for well behaved $P(r)$ functions
$P(r)$	High frequency oscillations or discontinuities in $P(r)$ may indicate problematic Fourier transform process

C.D. Putnam et al. Quart. Rev. Biophys. 40, 191-285 (2007)



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Summary

The CLS has a SAXS endstation available and our user community is growing!

Probe relatively big things (1 - 150 nm) with X-rays by measuring elastic scattering at small angles

Can learn about 1) packing, 2) particle size, and 3) particle shape, but generally not all 3 at the same time

Limited structural information

Complementary information, that should be supported with supplementary techniques such as microscopy



Further reading

Basic SAXS concepts

H. Schnablegger, Y. Singh, “The SAXS Guide: Getting Acquainted With the Principles”, Anton Parr, Austria. (2017)

Making a good measurement

B.R. Pauw, “Everything SAXS: small-angle scattering pattern collection and correction” J. Phys.: Condens. Matter 25 (2013) 383201

Basic data work up

J.B. Hopkins, R.E. Gillilan, S. Skou
https://bioxtas-raw.readthedocs.io/en/latest/saxs_tutorial.html

BioSAXS, data workup, introduction to structure modelling

C.D. Putnam, M. Hammel, G.L. Hura, J.A. Tainer, “X-ray solution scattering (SAXS) combined with crystallography and computation: defining accurate macromolecular structures, conformations and assemblies in solution” Quarterly Reviews of Biophysics 40, 3 (2007), pp. 191–285.



The Brockhouse Sector Team



Stefan Kycia
Professor, U. Guelph
Beamteam Leader



Al Rahemtulla



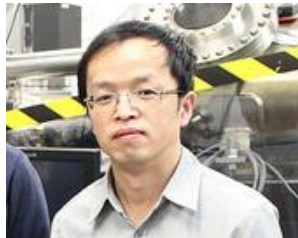
Chang-Yong Kim



Narayan Appathurai



Beatriz Diaz Moreno
Beamline Responsible
Beatriz.Moreno@lightsource.ca



Feizhou He
Mat. Chem. Sci. Manager



Joel Reid



Graham King



Adam Leontowich
Adam.Leontowich@lightsource.ca

<https://brockhouse.lightsource.ca>

Call for General User Proposals closes August 23, 2023 at noon (Sask. time)

