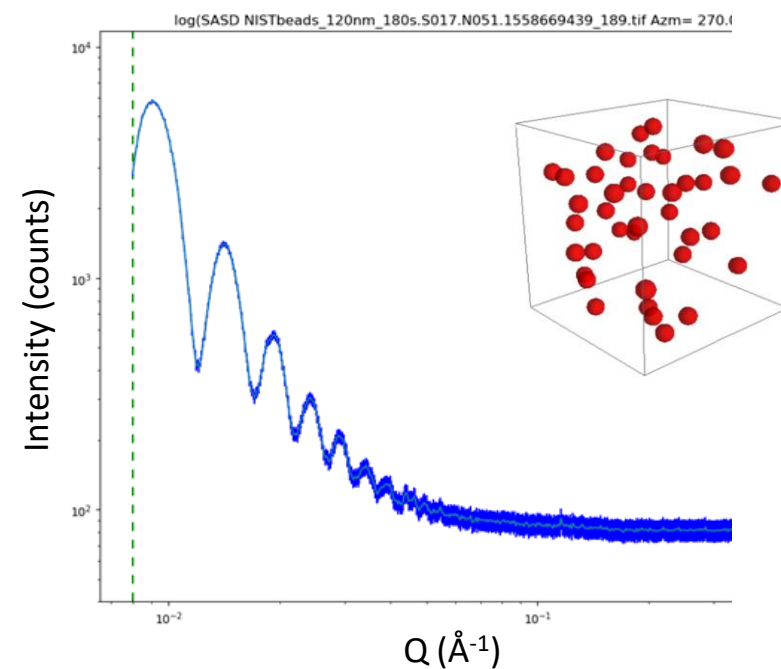
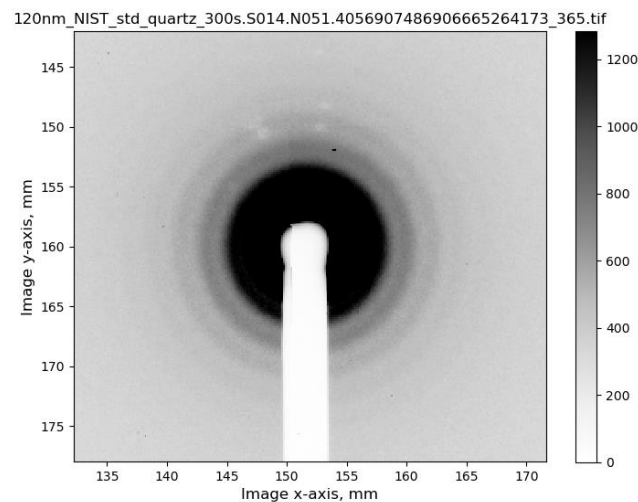
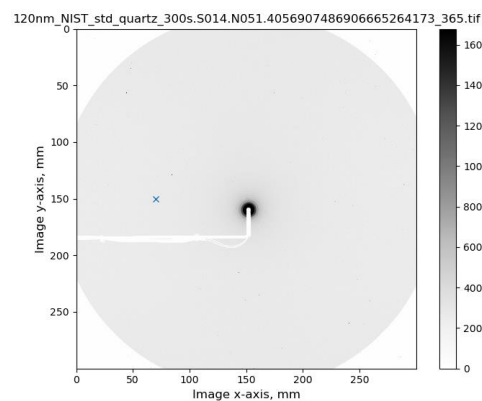
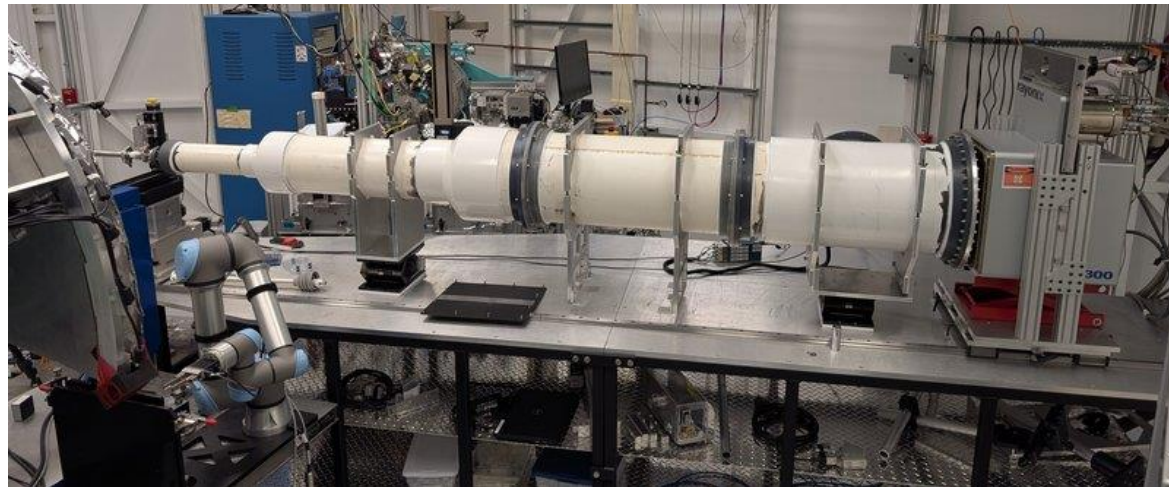


An Introduction to Small Angle X-ray Scattering (SAXS)



Adam Leontowich
CLS XRD school, Thursday, July 16, 2026



Canadian
Light
Source Centre canadien
de rayonnement
synchrotron

The small angle X-ray scattering (SAXS) region

Instrumentation, sample prep, data collection and reduction

Basic data processing with examples from BXDS

Data processing tutorials



Brockhouse X-ray Diffraction and Scattering (BXDS) sector

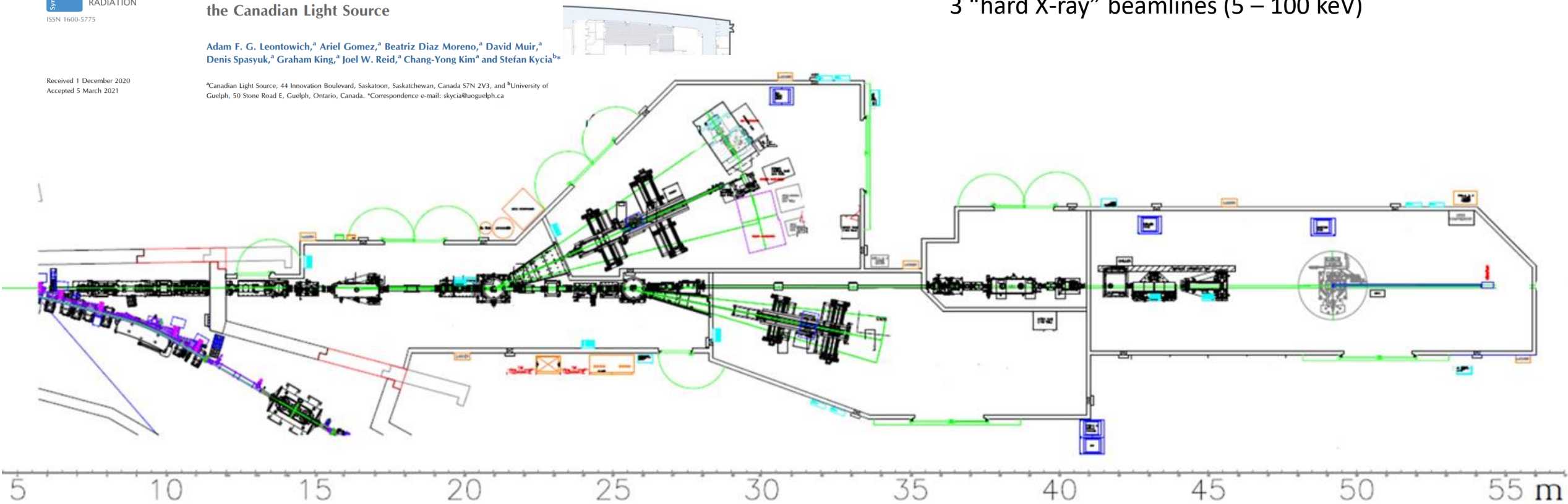
The lower energy diffraction and scattering side-bounce beamline for materials science at the Canadian Light Source

Adam F. G. Leontowich,^a Ariel Gomez,^a Beatriz Diaz Moreno,^a David Muir,^a Denis Spasyuk,^a Graham King,^a Joel W. Reid,^a Chang-Yong Kim^a and Stefan Kycia^{b*}

Received 1 December 2020
Accepted 5 March 2021

^aCanadian Light Source, 44 Innovation Boulevard, Saskatoon, Saskatchewan, Canada S7N 2V3, and ^bUniversity of Guelph, 50 Stone Road E, Guelph, Ontario, Canada. *Correspondence e-mail: skycia@uoguelph.ca

3 “hard X-ray” beamlines (5 – 100 keV)

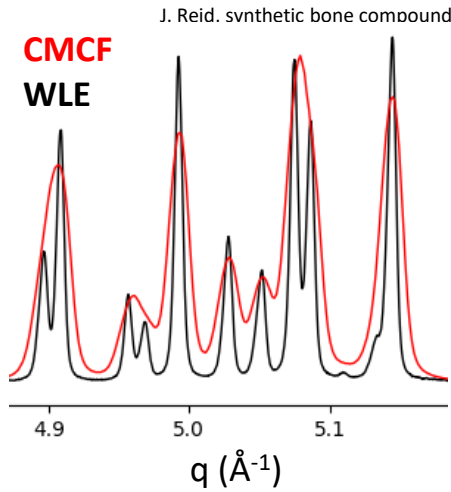


WLE beamline

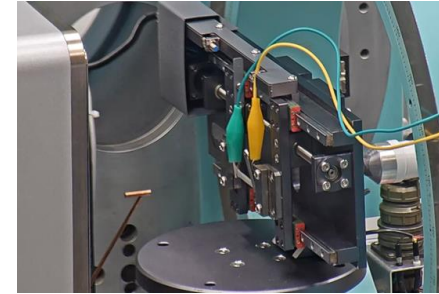
WAXS (XRD with area detector)

Biopolymers for bandages
Youchao Teng, Yimin Wu, U. Waterloo

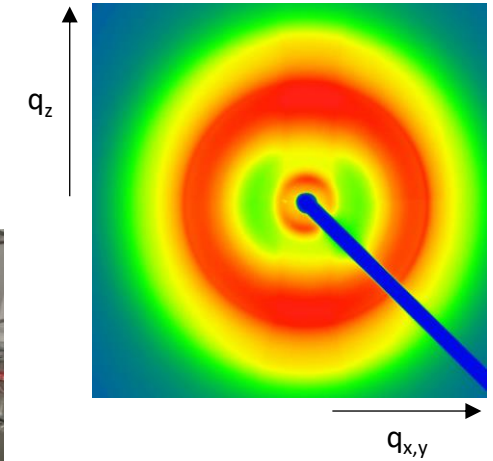
World class high-resolution powder diffraction



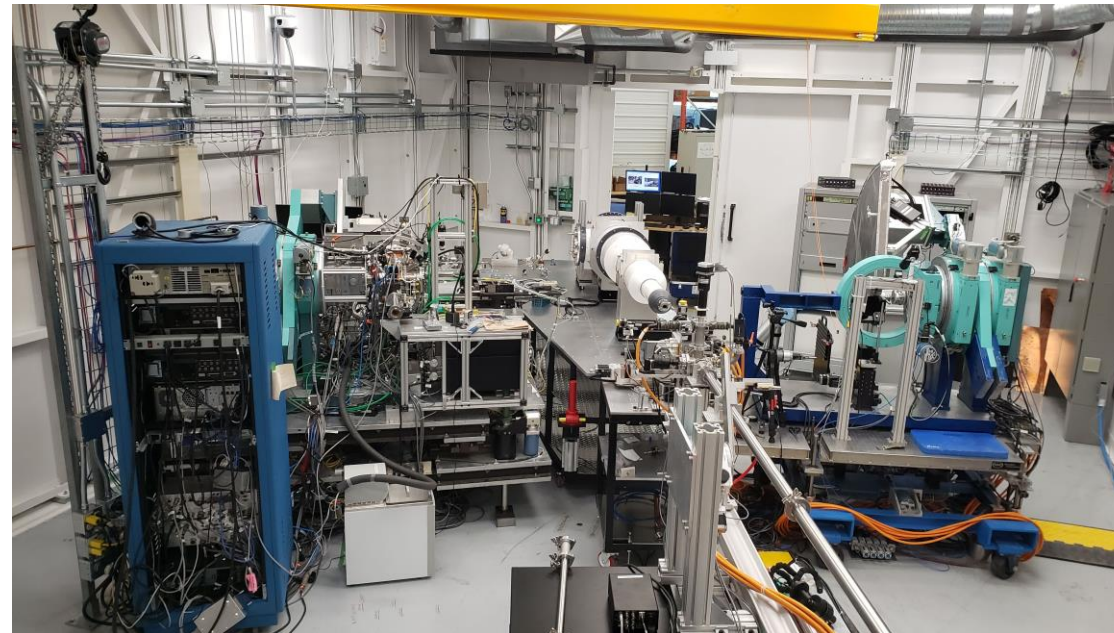
- Complex structures
- Complex mixtures
- Peak shape analysis



Measure WAXS while stretching (0 – 100 N)



- Surfaces
- Orientation
- Speed/in-situ

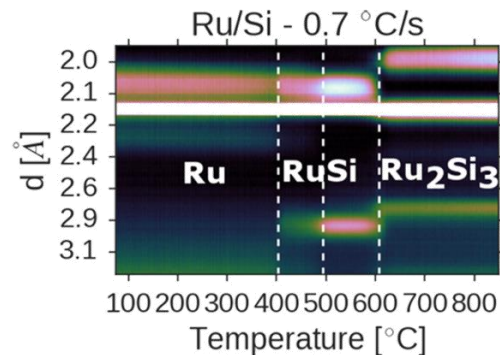


SAXS

SAXS has been running since February 2021

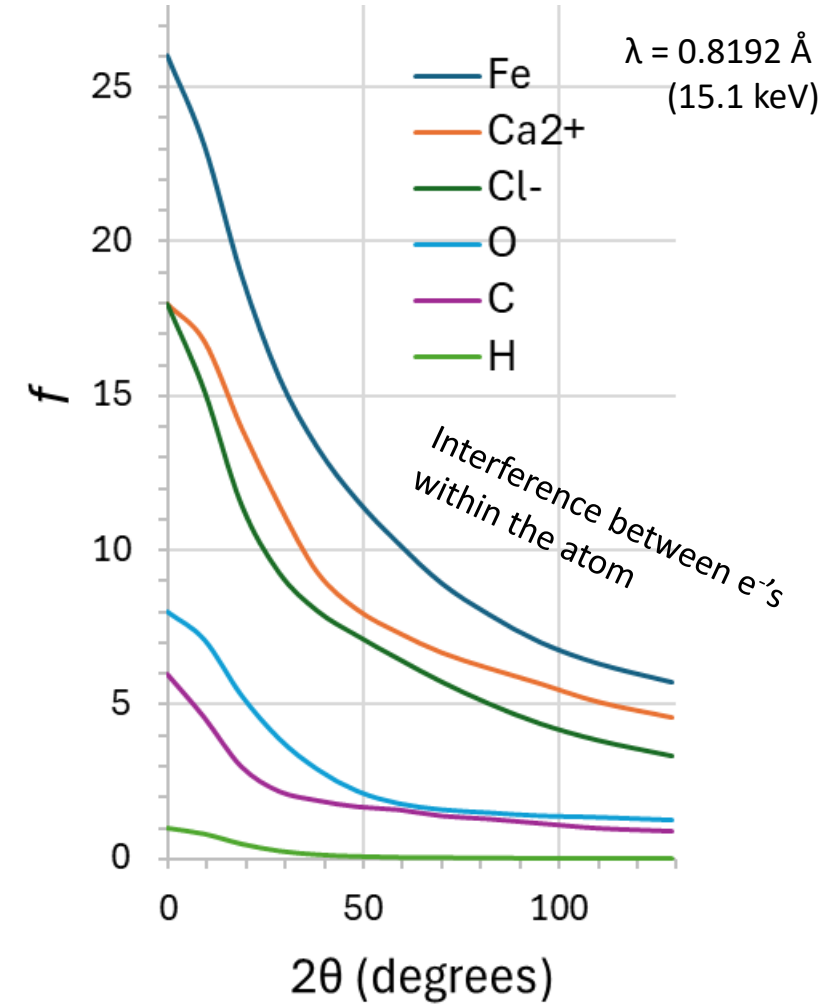
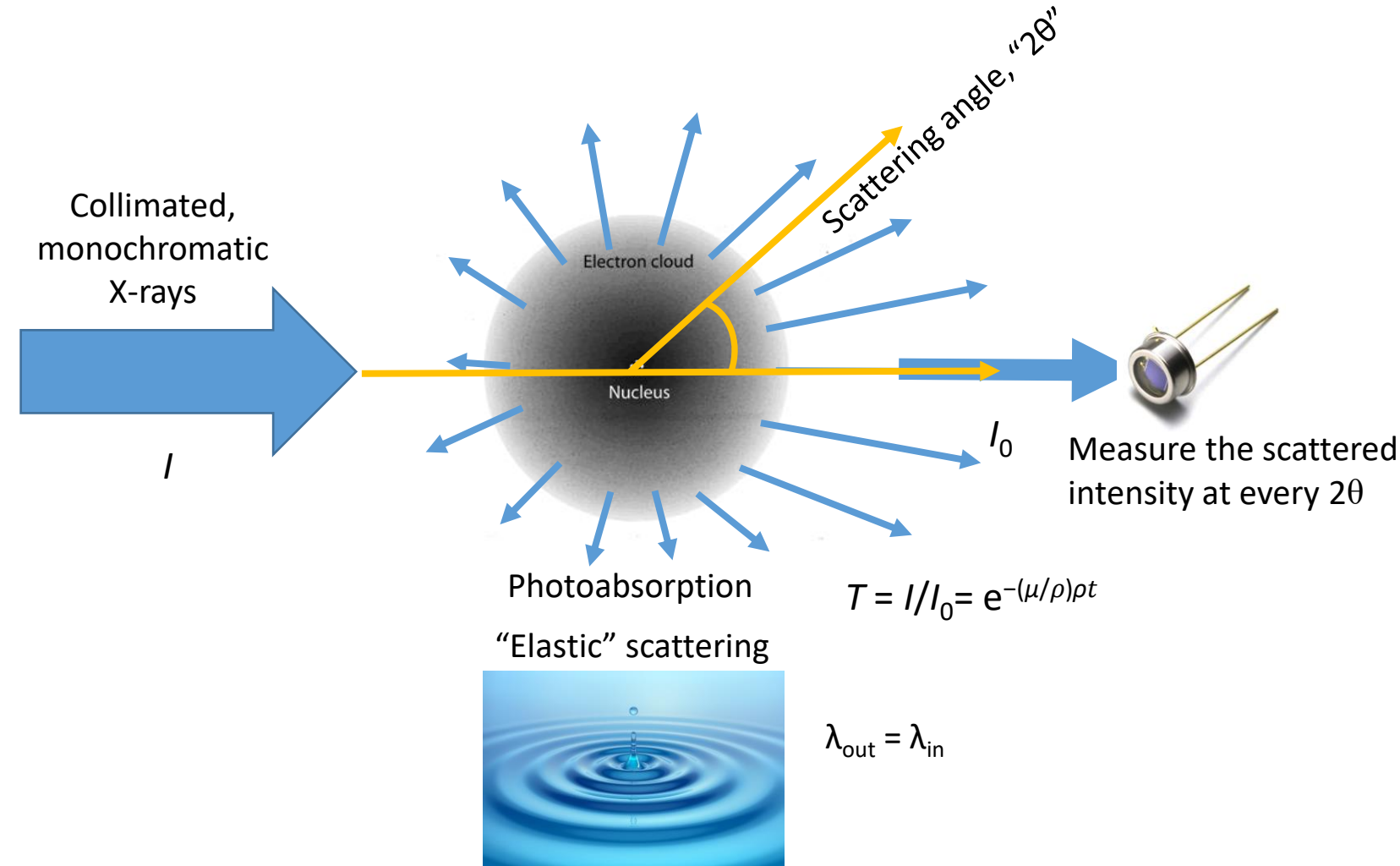
Thin film rapid thermal annealing
~1000 °C, XRD, resistivity, roughness,
ultra-high purity gas

- metal films



X-ray elastic scattering

Atomic form factors (or atomic scattering factors)

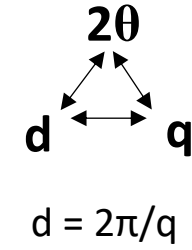


$$f = \frac{\text{amplitude scattered by an atom}}{\text{amplitude scattered by a single electron}}$$

From scattering to diffraction

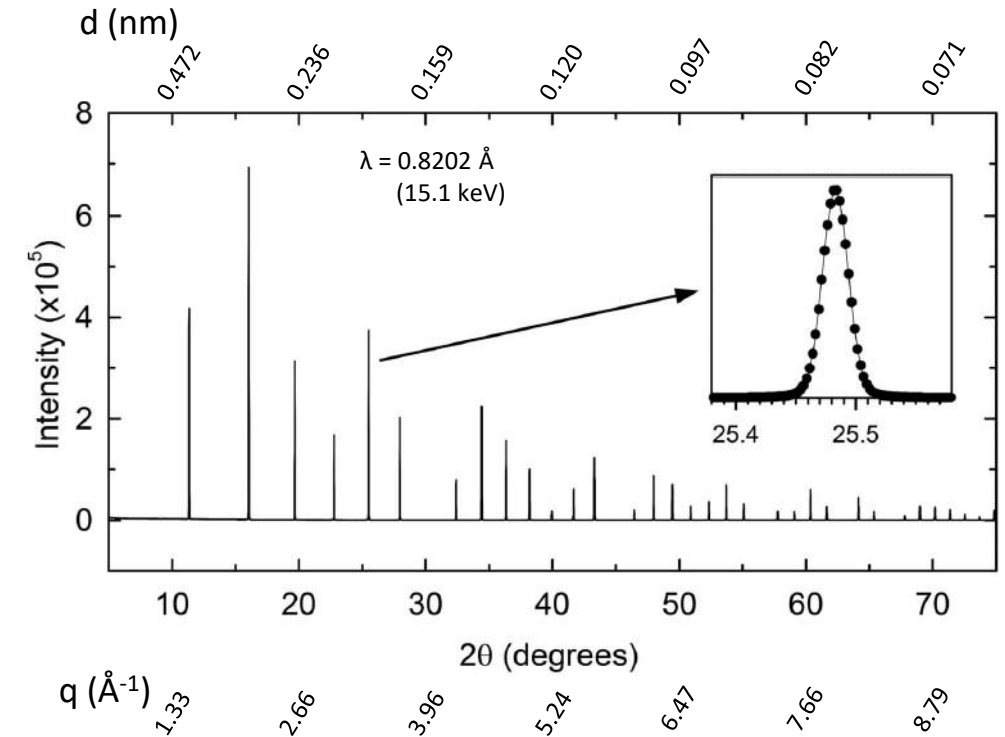
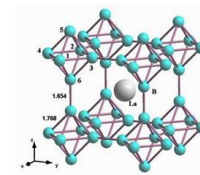
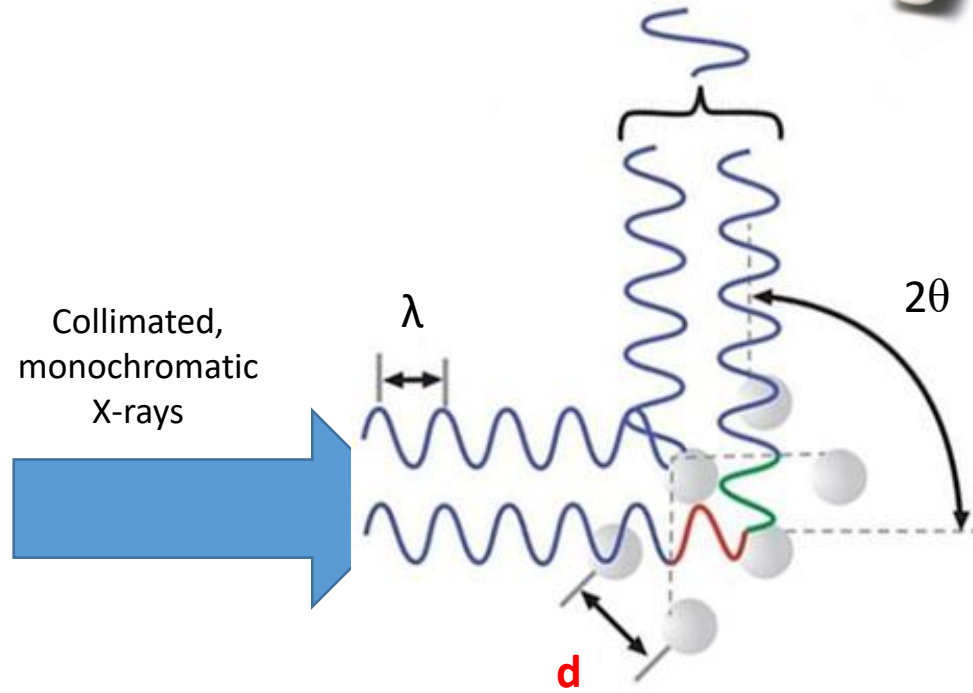
"Bragg" diffraction

$$n\lambda = 2d \sin\theta$$



$$q = (4\pi \sin\theta)/\lambda$$

Constructive interference channels
filtered intensity into specific directions



X-ray scattering and diffraction reveals structural order within materials

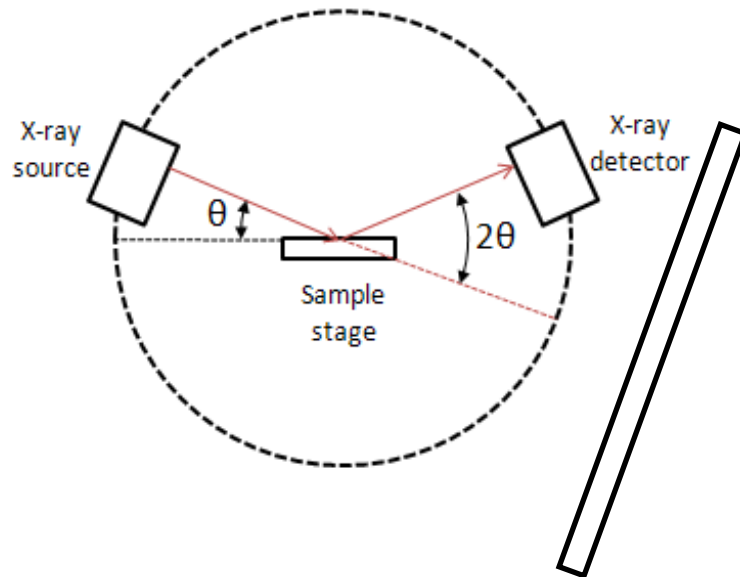


Canadian
Light
Source

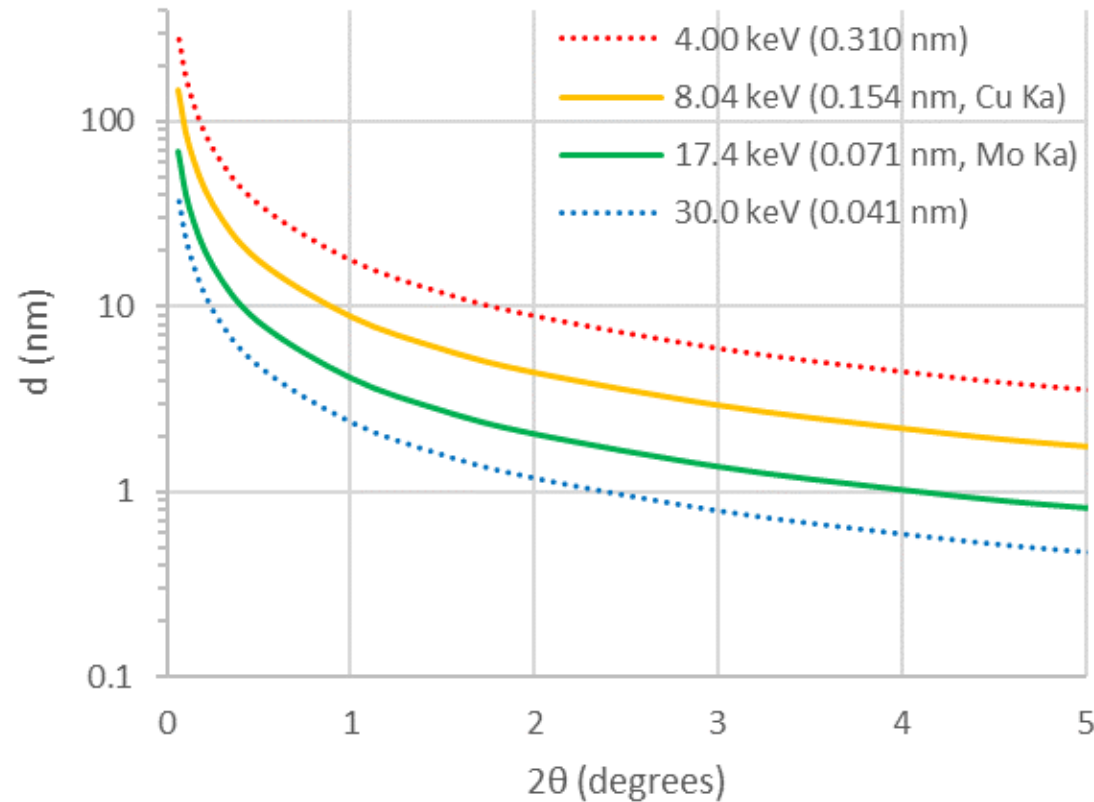
Centre canadien
de rayonnement
synchrotron

What is small angle X-ray scattering all about?

Starting from powder diffraction...



$$\lambda = 2d\sin\theta$$



The nano world

Atomic distances,
spacing between
planes of atoms,
bond lengths

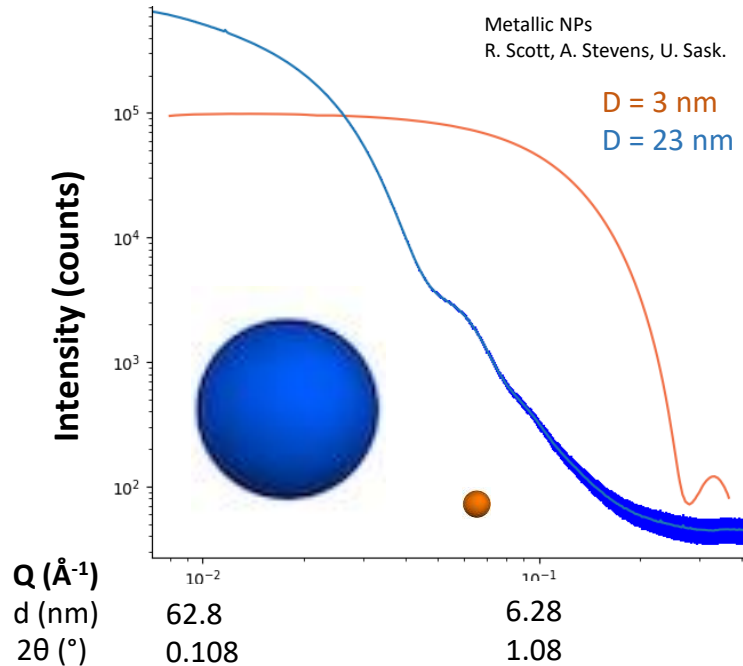
SAXS is generally $2\theta = 0.05 - 5^\circ$

Probe relatively big things (1 - 150 nm) by measuring elastic X-ray scattering at small angles

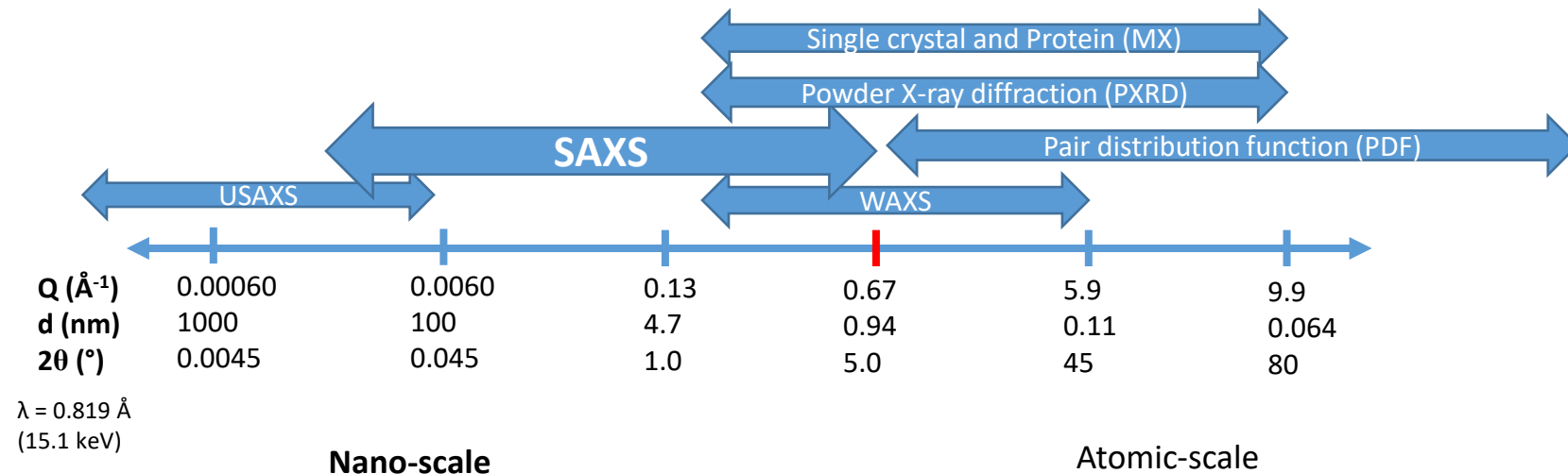
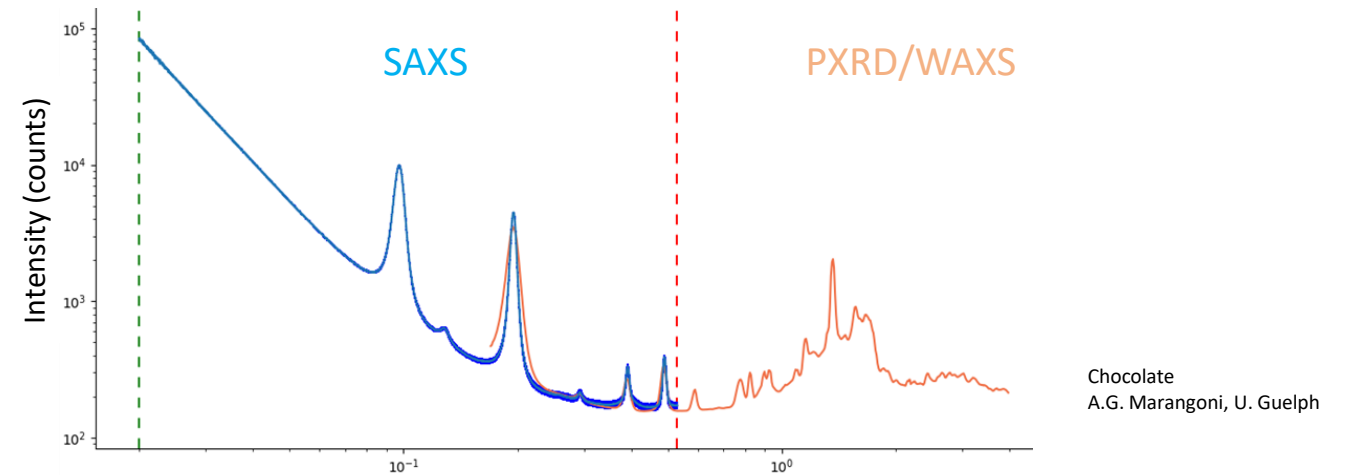


How is SAXS related to other X-ray diffraction techniques?

Typical SAXS data presentation



- 1D intensity vs. Q
- log-log plot
- Proper intensity units are cm⁻¹ if data are scaled to absolute intensity



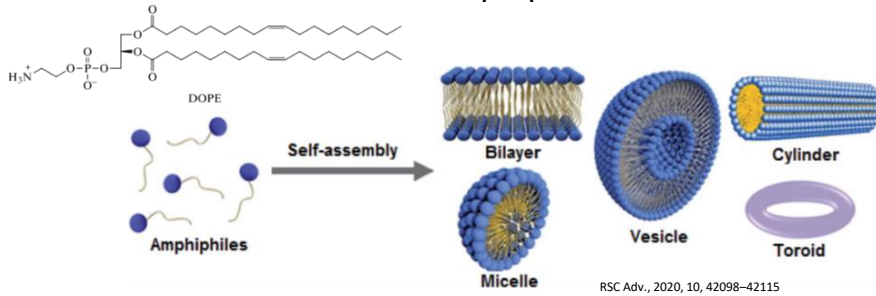
Inter-molecular distances and packing arrangements, particle size and shape

Inter-atomic distances, bond lengths, nearest neighbor atoms, Crystal structures with atomic resolution



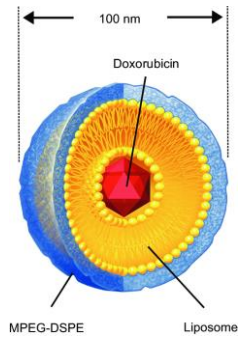
Applications

Amphiphiles



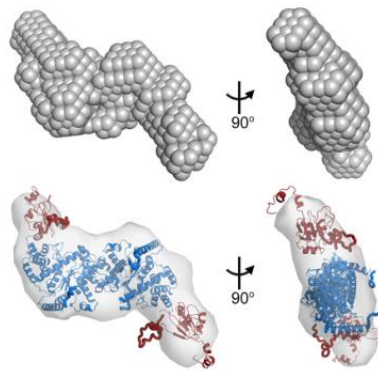
Surfactants,
Micelles, emulsions

Drug delivery,
Pharmaceuticals



Biomacromolecules

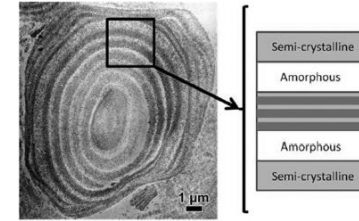
DNA
Alginate
Cellulose
Collagen
Chitosan
Proteins



Food science

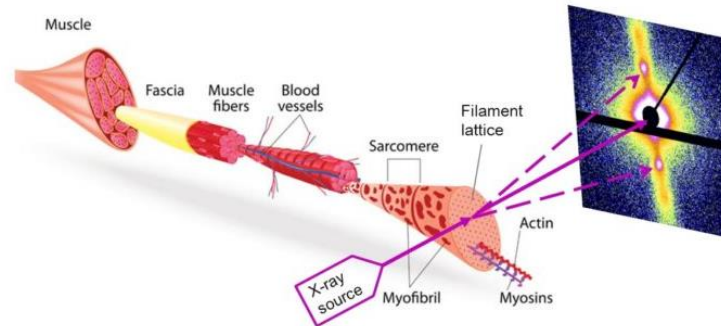


Fats
Starch
Pectin



S. Perez and E. Bertoft, *Starch* **2010**, 62, 389 - 420

Polymers

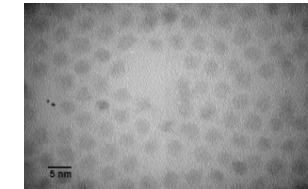
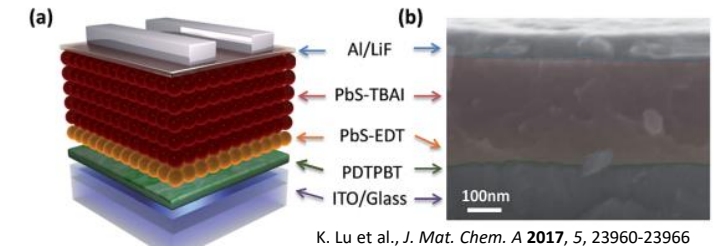


<https://www.techexplorist.com/scientists-investigated-pork-fillet-x-ray-light/11572/>

- 1) Packing, ordering
- 2) Size, size distribution
- 3) Shape

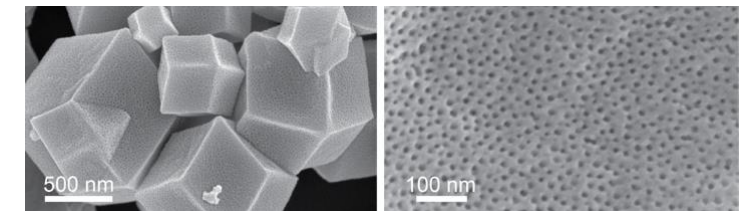


All kinds of nanoparticles



M. Yuan et al., *Adv. Mater.* **2014**, 26, 3513–3519

Nano-porous materials



M. Liu, Z.M Hudson, UBC



Batteries
Biochar

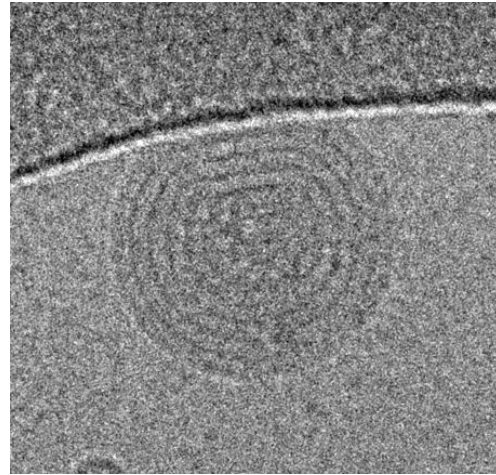
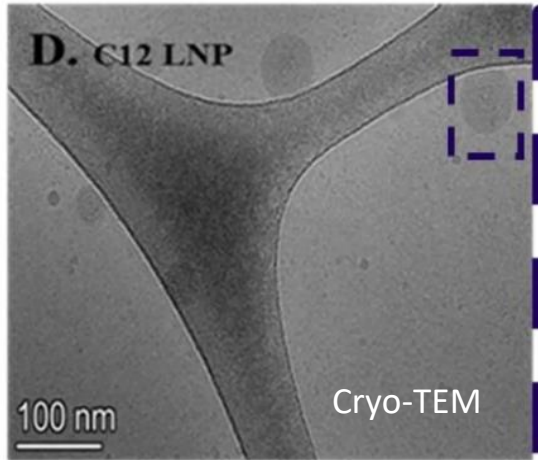
Zeolites

Geology



Canadian
Light
Source
Centre canadien
de rayonnement
synchrotron

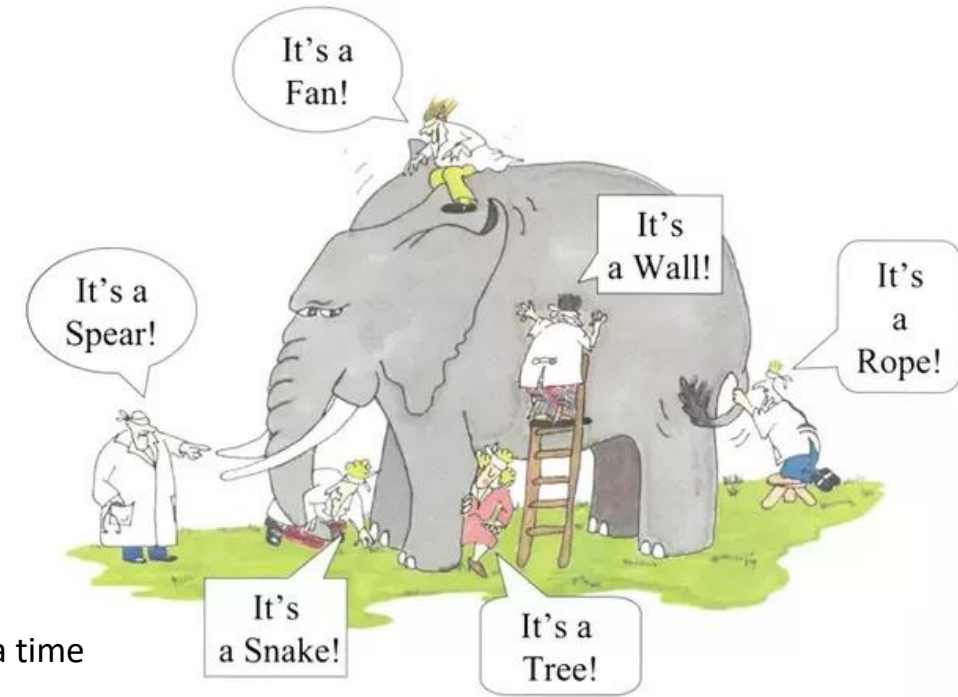
SAXS is highly complementary to nanoscale microscopy



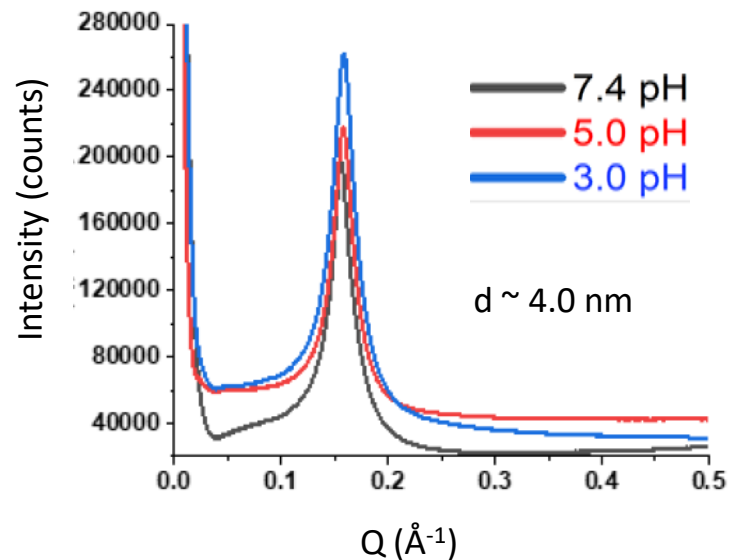
$d \sim 4.0 \text{ nm}$

Naga Kola, Thakur Group, VIDO

- FOV decreases as resolution increases
- Observe a small fraction of the complete sample at a time
- Challenging sample prep at smaller size



"Blind men and an elephant"



- SAXS provides nanoscale information **averaged** over the beam volume ($\sim \text{mm}^3$)
- Often complementary or extra info: surface roughness, folding/unfolding, assembled state in solution

Instrumentation, sample prep, data collection and reduction

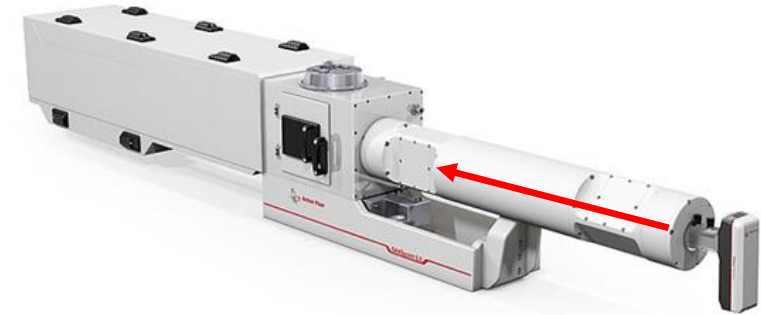


How to measure small angle X-ray scattering?

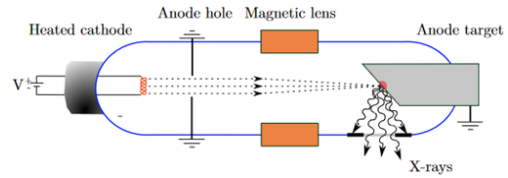
PXRD



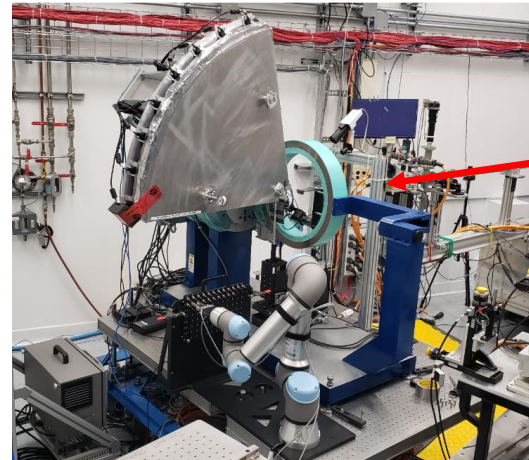
SAXS



Lab source

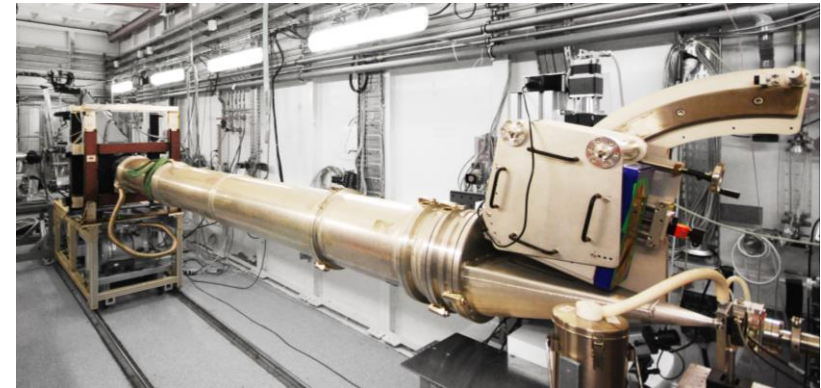


Synchrotron



X-rays

ESRF - BM26

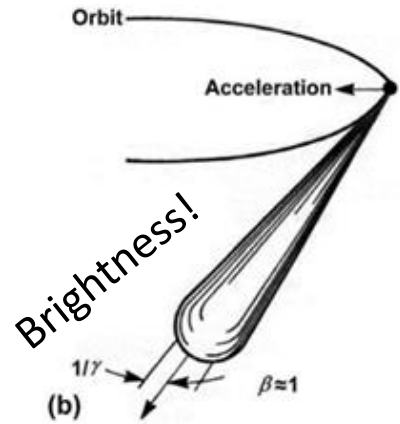
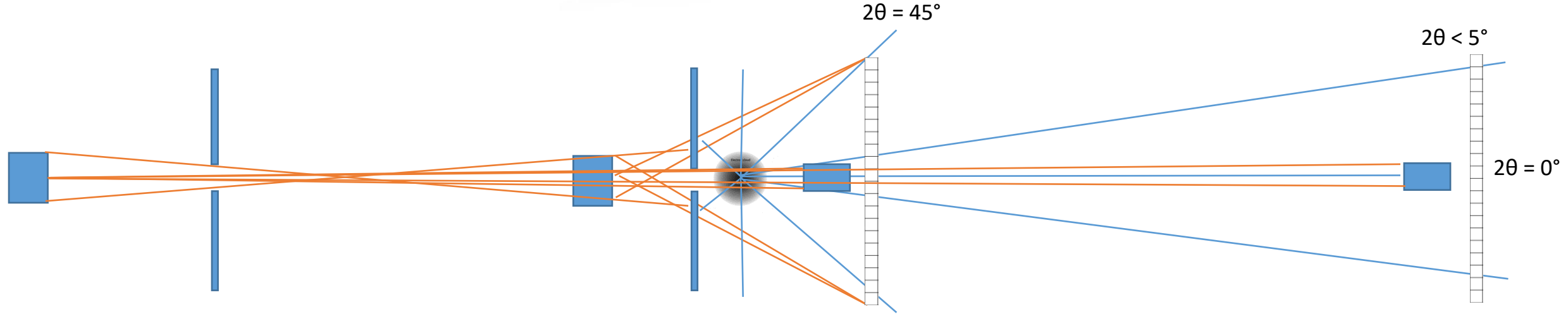


X-rays

- 1) Special instrumentation required for SAXS
- 2) SAXS instruments are long!



SAXS instrumentation basics



- More flux (better SNR), weakly scattering objects (organics/biology)
- Better q resolution
- Time resolved, SAXS mapping
- Choice of many wavelengths

- Highly collimated hard X-ray beam (multiple slits)
- Large area detector with beamstop
- Space for >2 m sample to detector distance
- Some ability to change detector distance (SAXS/WAXS)
- Most, or ideally all, components in vacuum



SAXS user facilities in Canada

SFU



SAXSLAB, Ganesha 300XL+

<https://www.4dlabs.ca/our-capabilities/equipment/small-angle-x-ray-scattering-system.html>

UBC
Soon! Ask Anita



CLS

McGill



Anton Paar, SAXSpoint 2.0

<https://www.mcgill.ca/mc2/instrumentation/x-ray-diffraction/saxs>

U. Montreal

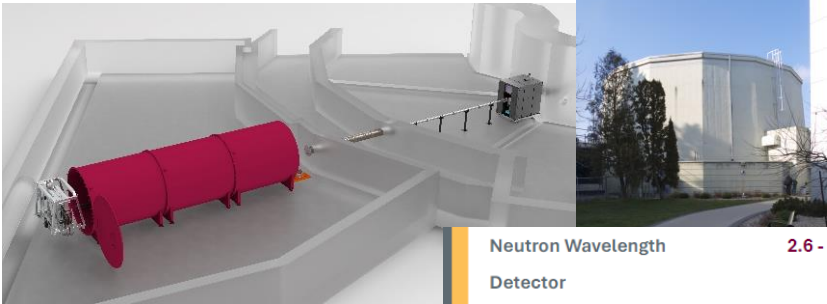


SAXSLAB, Ganesha

http://airen.bcm.umontreal.ca/biostruct/sc heduling_saxs/

U. Ottawa

McMaster



MacSANS – In commissioning

<https://nuclear.mcmaster.ca/neutron-beams/>

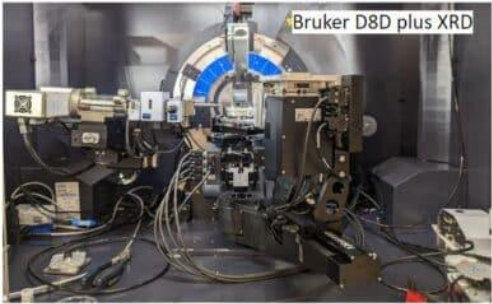
U. Toronto



Anton Paar, SAXSpace

<https://lab.research.sickkids.ca/sbc-facility/saxs/>

U. Alberta



Bruker D8D Plus XRD

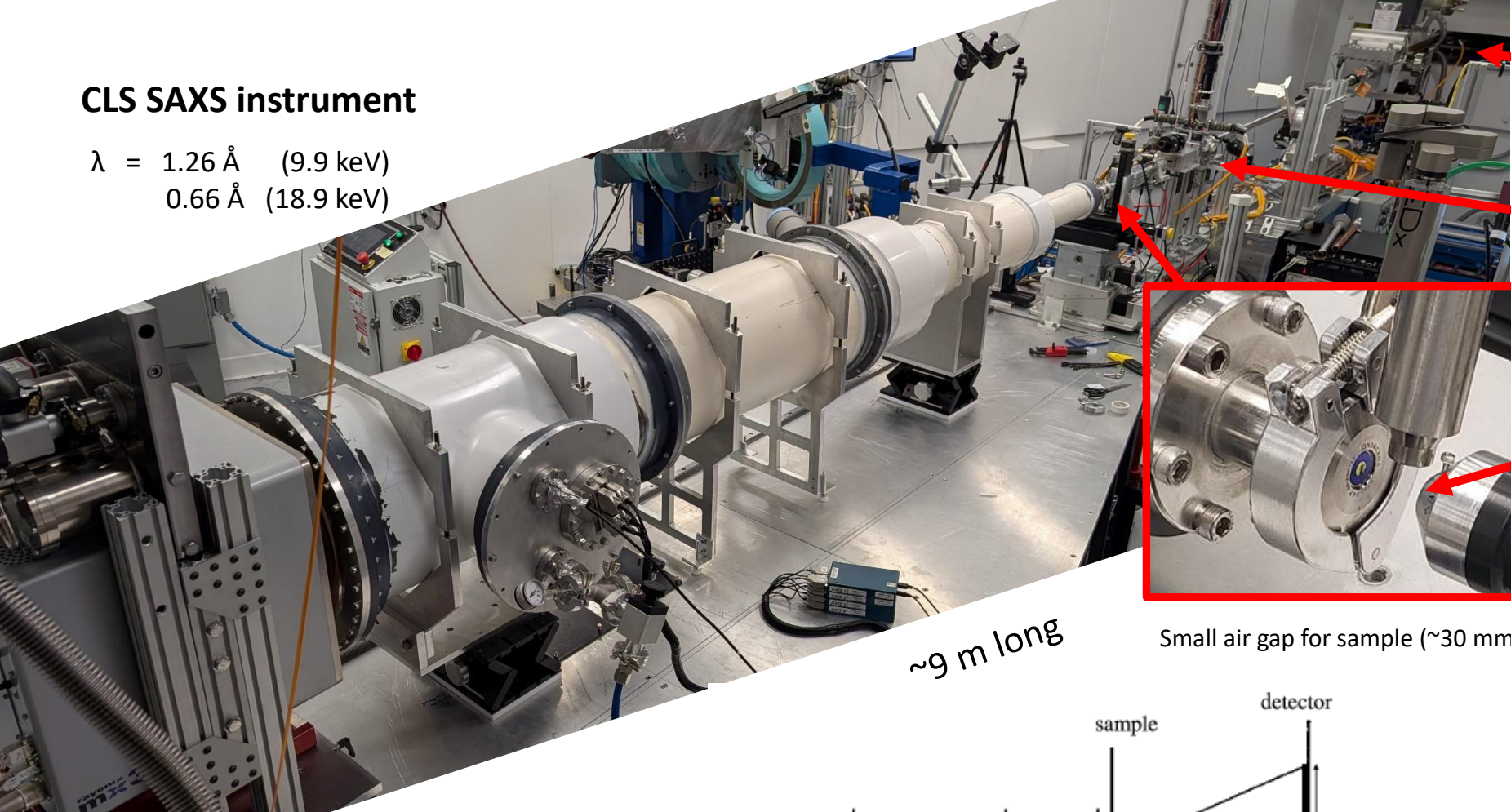
<https://www.nanofab.ualberta.ca/saxs-gi-saxs-waxs-are-available-on-bruker-d8d/>

Neutron Wavelength
Detector
Accessible Q-range
Structure Scale

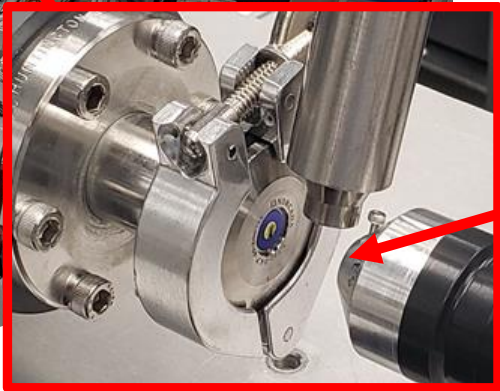
2.6 - 23 Å at $\Delta\lambda/\lambda \approx 17\%$
100 cm² at 0.9 cm
0.005 < Q < 1.25 Å⁻¹
0.6 – 125 nm

CLS SAXS instrument

$\lambda = 1.26 \text{ \AA} \quad (9.9 \text{ keV})$
 $0.66 \text{ \AA} \quad (18.9 \text{ keV})$

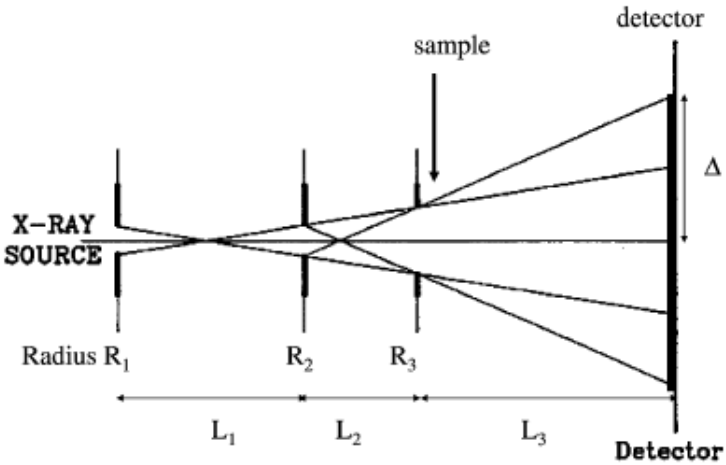


- 1) Beam defining slit
+6.1 m
(1.00 mm x 1.00 mm)
- 2) Anti-scatter slit
+0.9 m
(0.25 mm V x 0.50 mm H)
- 3) Guard slit (0.80 mm)

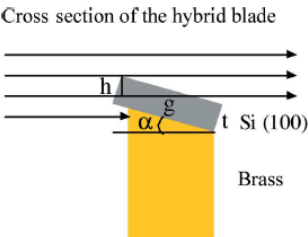


~9 m long

Small air gap for sample (~30 mm)



low parasitic scatter



Y. Li et al., J. Appl. Cryst. (2008). 41, 1134–1139
N.M. Kirby et al., J. Appl. Cryst. 46, 1670–1680 (2013)

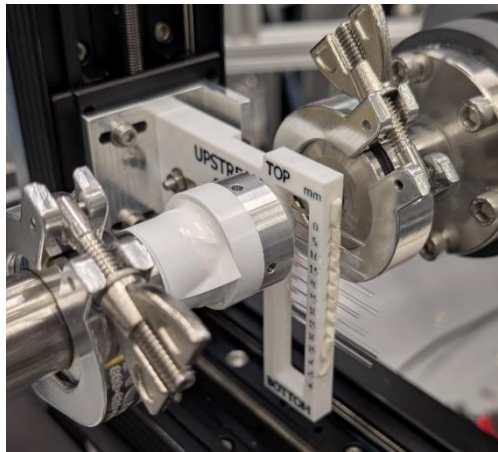
Figure 4
Schematic layout of a SAXS pinhole camera.

J. Appl. Cryst. (2004). 37, 369–380

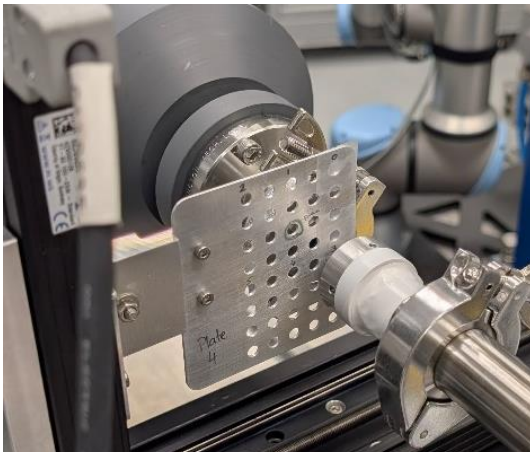
Sample preparation and mounting

Transmission

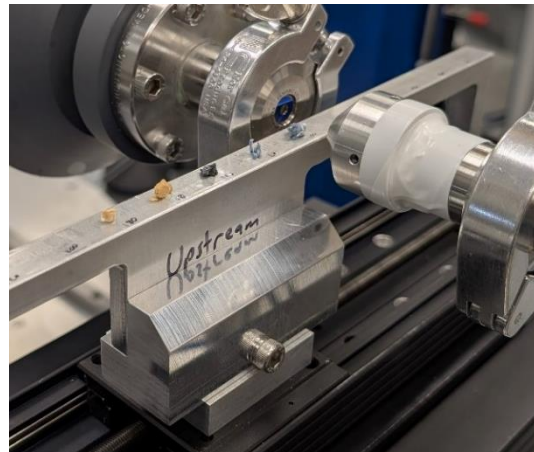
capillaries



between tape

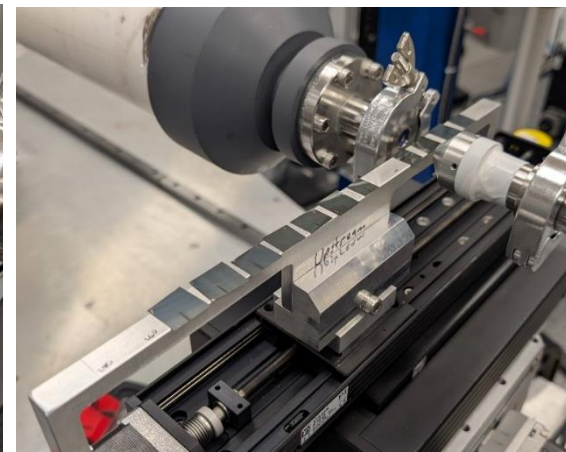


freestanding



Grazing incidence

Thin *flat* films on Si wafers, or glass slides



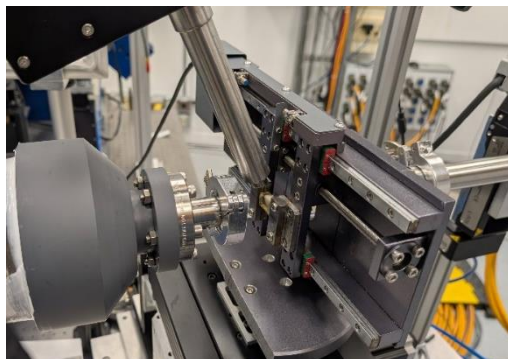
$\theta=0.01^\circ$

Automated measurement of ~10 samples in 30 to 60 minutes

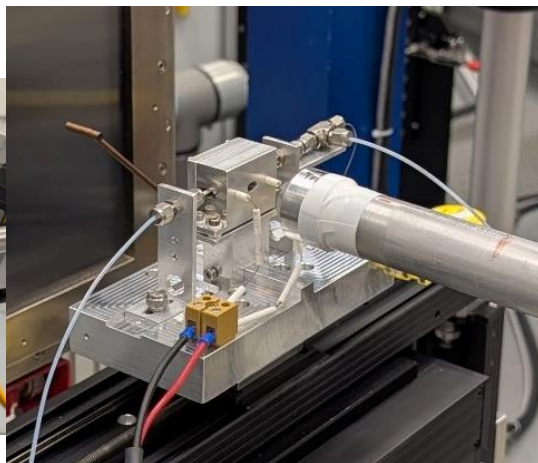
In-situ sample environments

Oxford cryostream (-193 – +226°C)

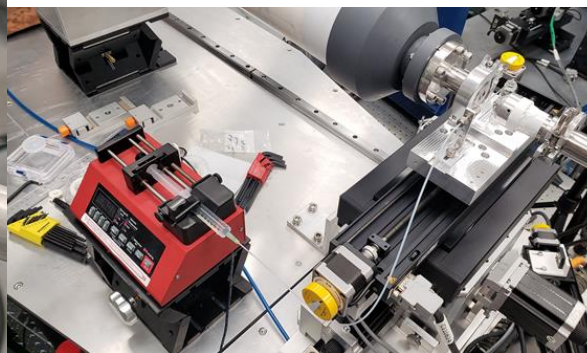
Stretching



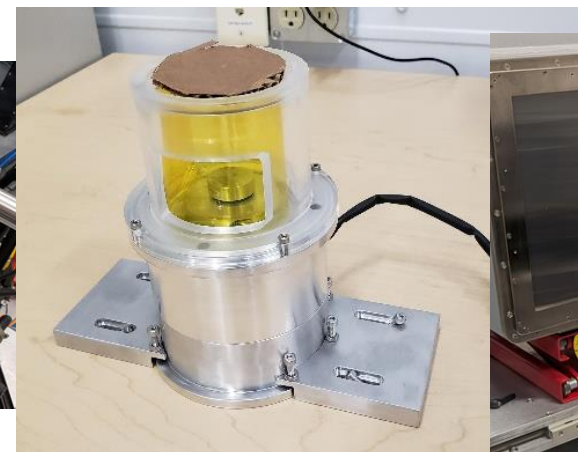
1200°C furnace with gas flow



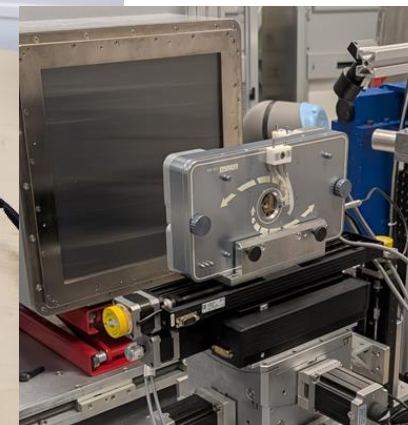
Liquid flow cell and syringe pump



In-situ spin coater with heating

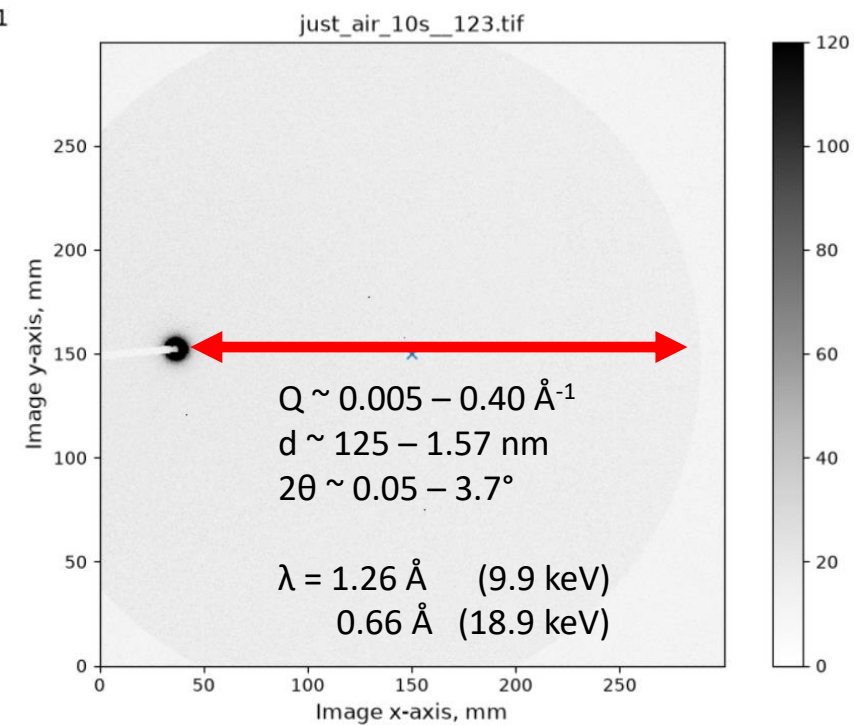
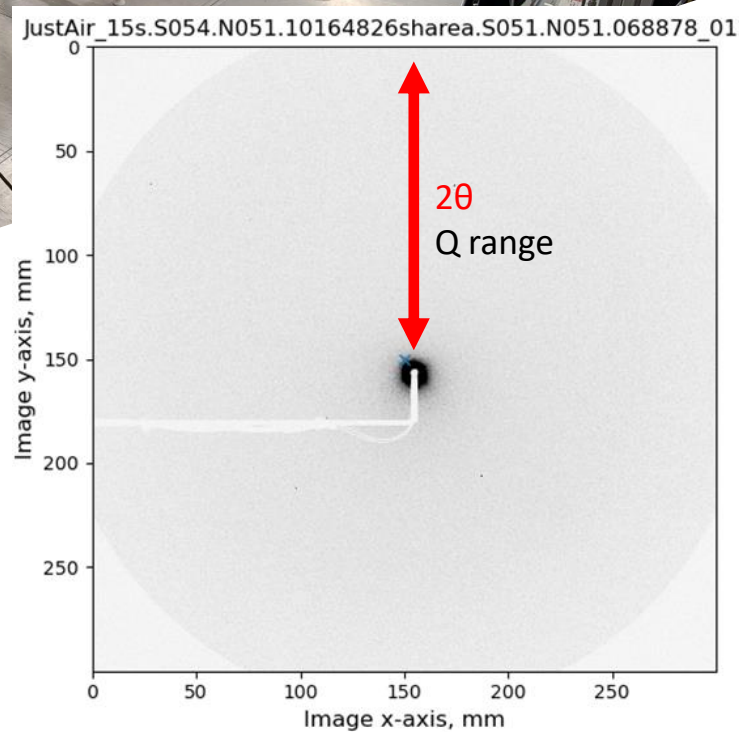
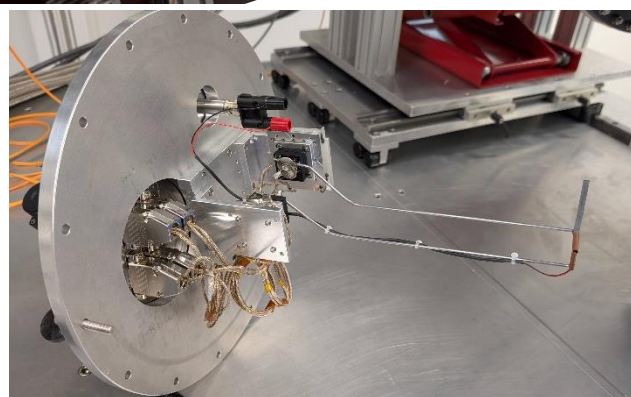
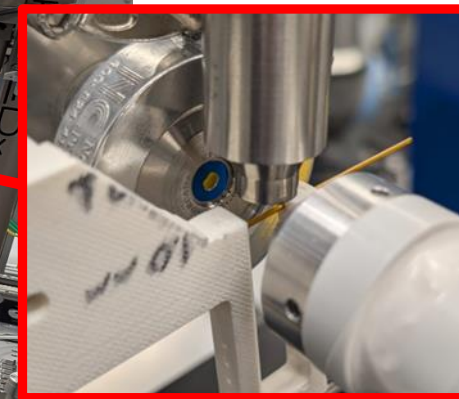
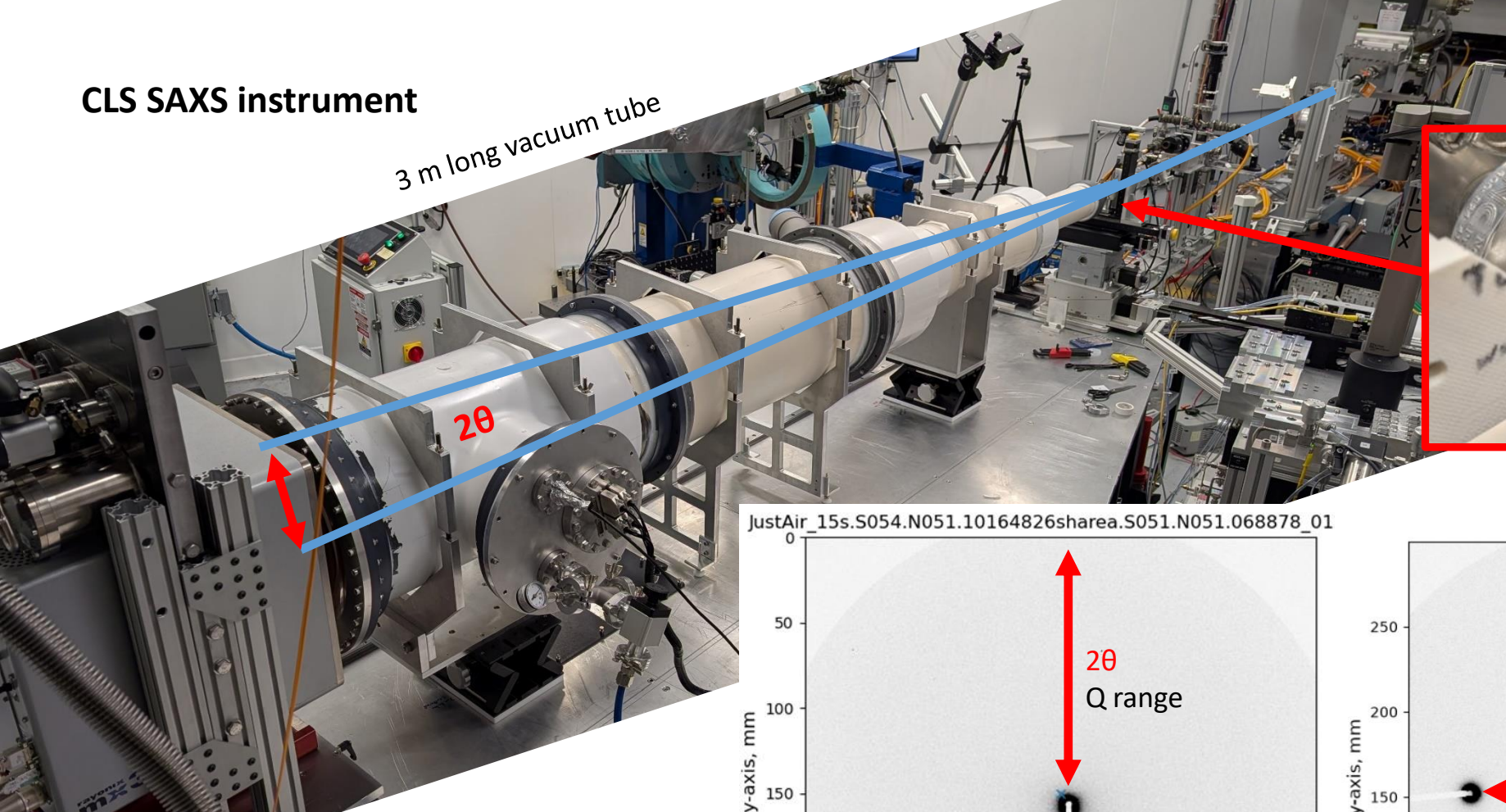


New shear stage



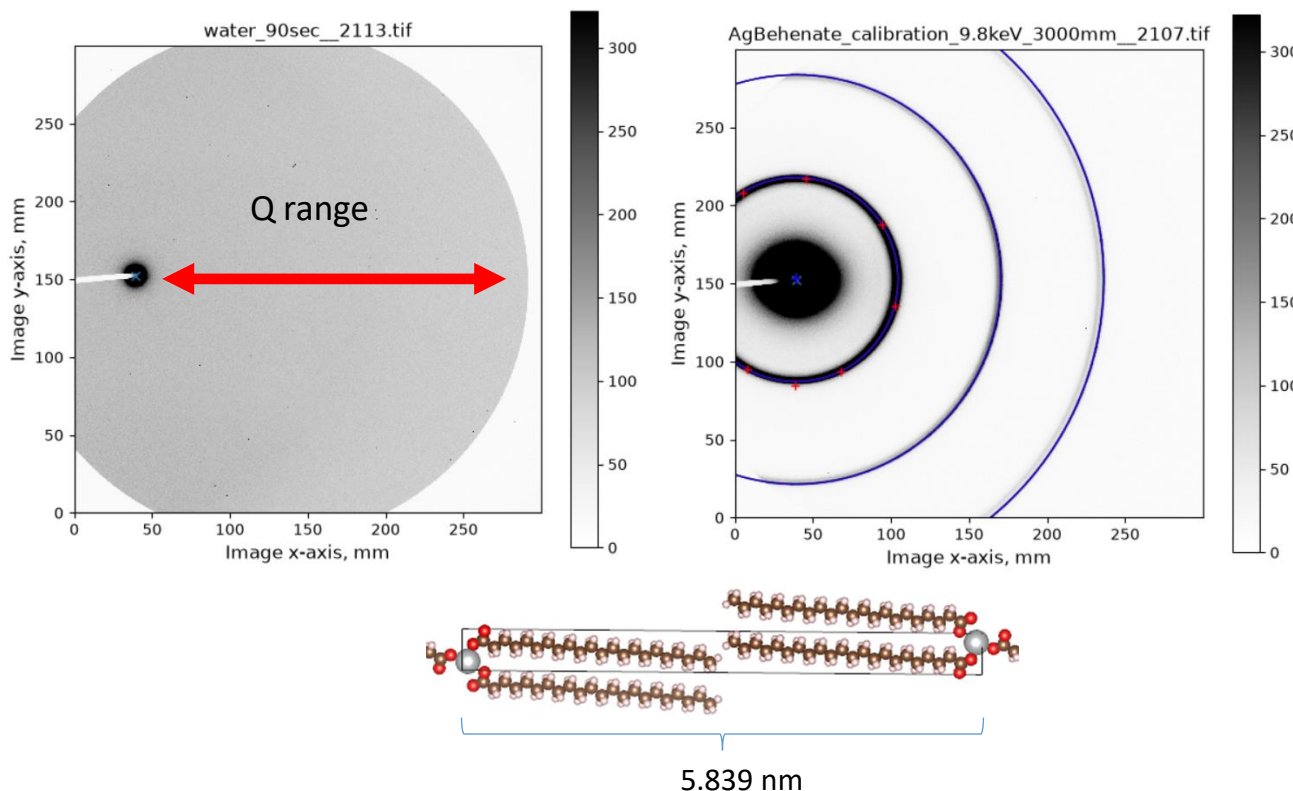
CLS SAXS instrument

3 m long vacuum tube



Detector acquisition and calibration

GSAS-II



Ring	q ($\text{\AA}^{-1}$)	d (nm)
001	0.1076	5.839
002	0.2152	2.920
003	0.3228	1.946
004	0.4304	1.460
005	0.5380	1.168

Need the wavelength, detector details, and approximate distance

Image Controls:

Type of image data: SASD - small angle scattering data Color bar Greys Azimuth offset 0.0

Max intensity 322

Min intensity 0

Auto scaler ? Show line scan

Calibration coefficients

☒ Beam center X 39.084

☒ Beam center Y 152.569

☐ Wavelength* 1.25602

☒ Distance 3045.074

☒ Tilt angle* 1.301

☒ Tilt rotation* -79.81

Integration coefficients

Bin style: Constant step bins in log(q) ☐ Pink beam source?

Inner/Outer Q 0.005 0.4

Start/End azimuth 270.0 450.0

No. 2-theta/azimuth bins 1000 1

☐ Show integration limits? ☐ Do full integration?

☐ Use for all new images? ☐ Azimuth at bin center?

☐ Apply sample absorption?

☐ Apply polarization? Value (0.001-0.999) 0.99

Dark image multiplier -1.0 Flat Bkg: 0

Background image multiplier -1.0

Gain map

Calibration controls:

Calibrant Ag behenate Calib lines to skip 0 Min calib d-spacing 16.0

Calibration Integration **Parms** Help

Image Controls:

Type of image data: SASD

Max intensity 322

Copy Controls

Copy Selected

Save Controls

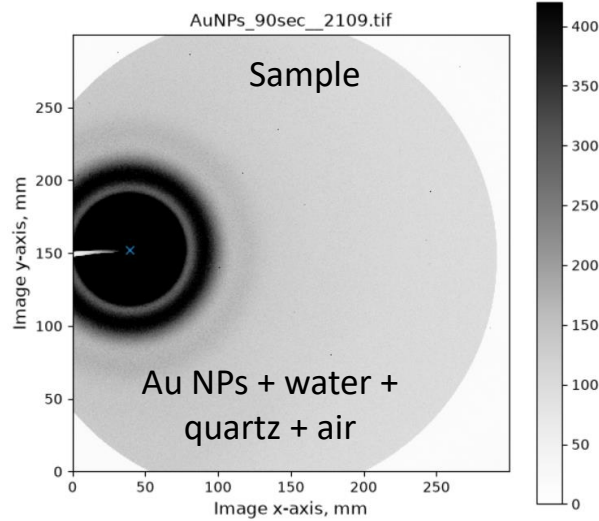
Save Multiple Controls

Load Controls

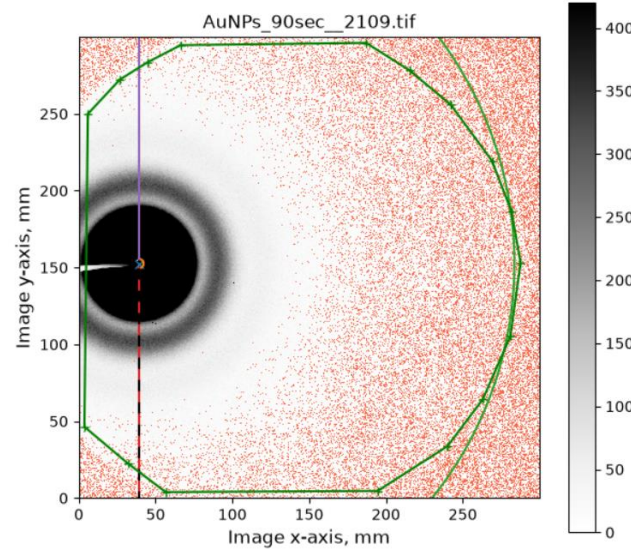


Canadian Light Source
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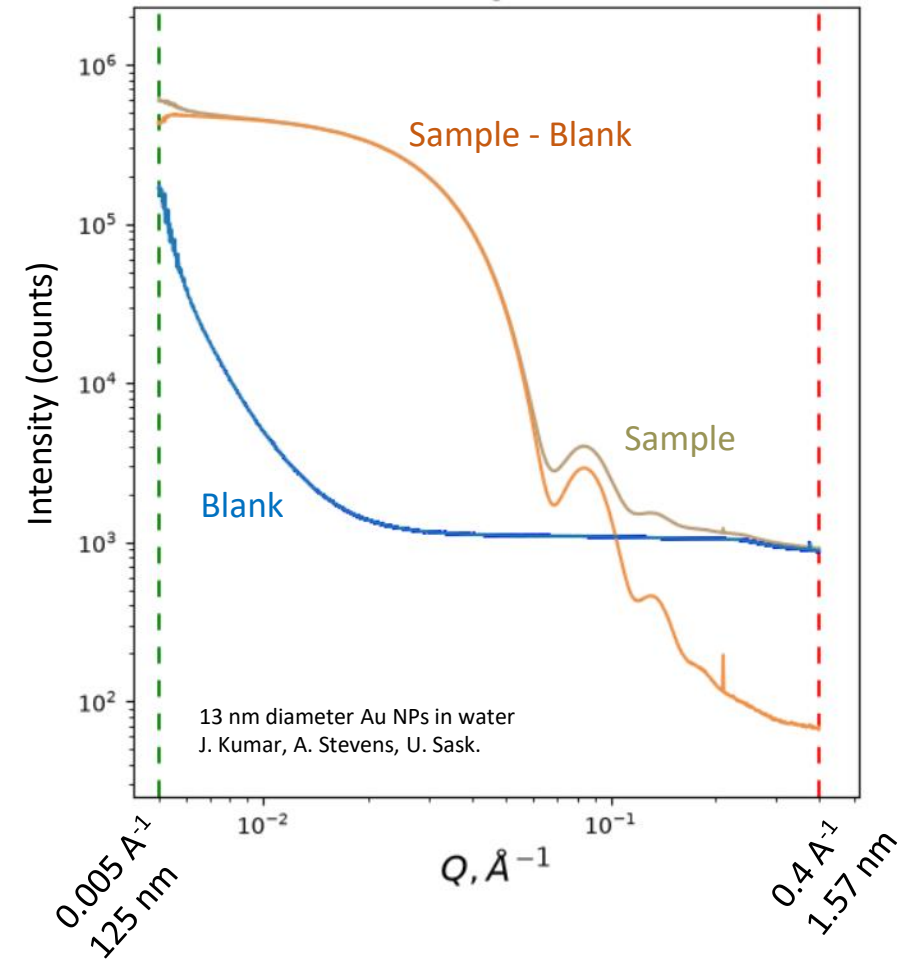
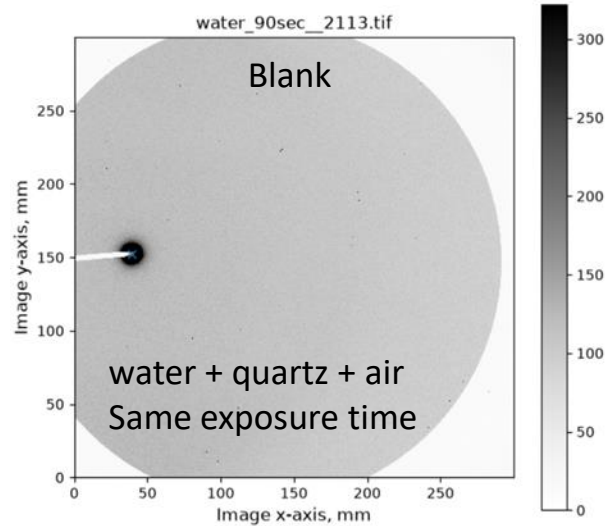
Data correction, and reduction to 1D plot



Acquisition time: 1 s - 90 s



Subtract the blank



Part 1: SAXS data calibration and reduction using GSAS-II

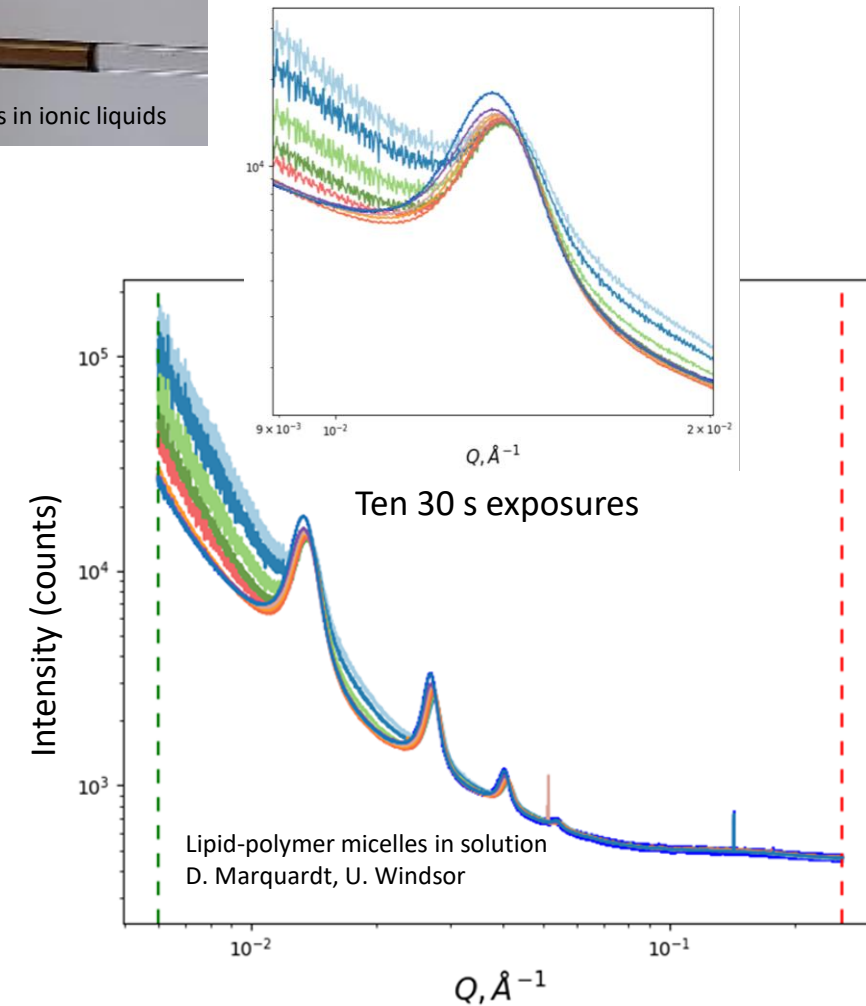
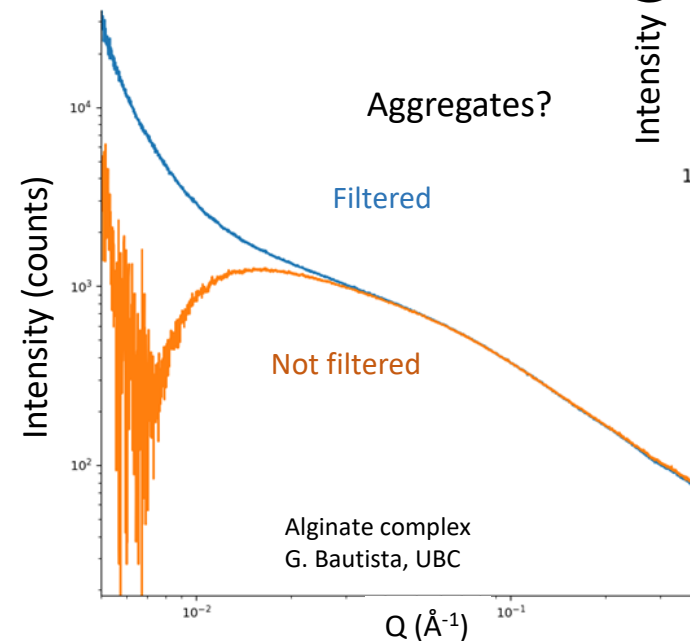
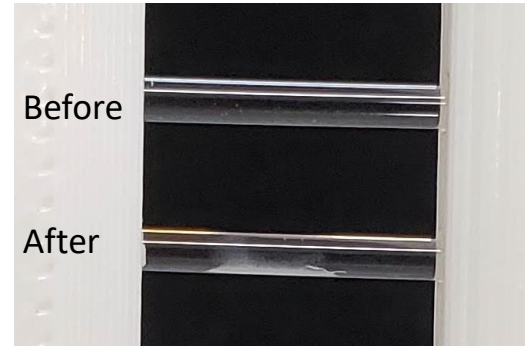
https://www.youtube.com/watch?v=BqnX_f0Hcdw

Part 2: SAXS data manipulation and analysis using BioXTAS RAW and SASView

<https://www.youtube.com/watch?v=-Yab0XOpsug>

Some tips for a great data collection experience

- 1) If the sample is in solution, bring blanks/solvent/buffer, and bring a range of concentrations (1 - 10 mg/mL)
- 2) Have a good idea of what you're looking for. SAXS patterns can be ambiguous and there are numerous interpretations and models. Bring some examples.
- 3) Quality over quantity: Work up data as you go along
- 4) Buy your own capillaries



Data analysis with examples



Origins of the SAXS signal

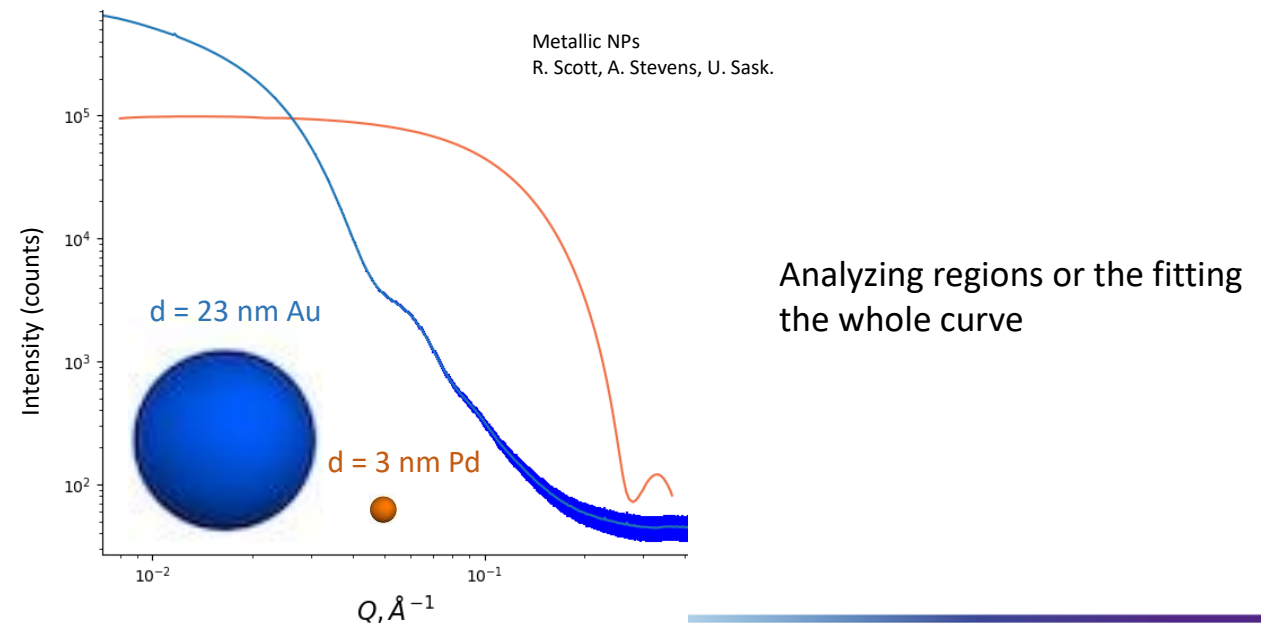
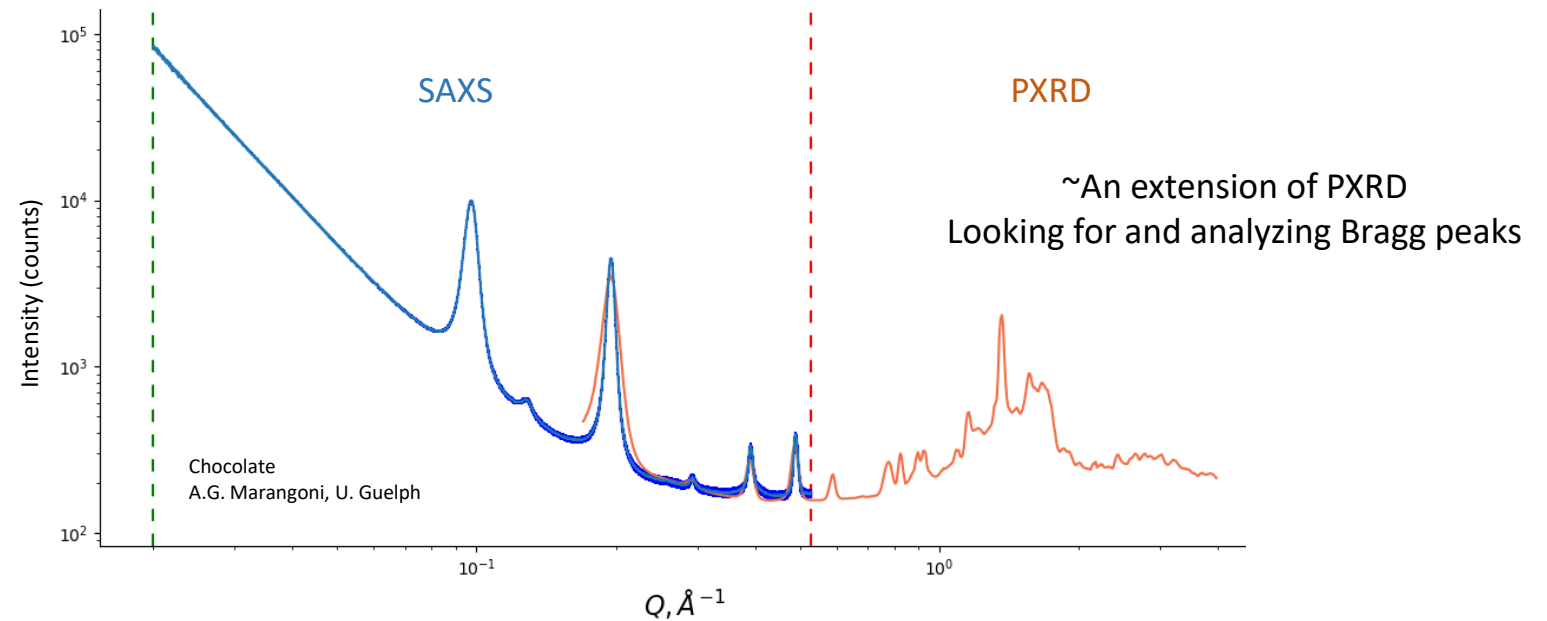
$$I(Q) = S(Q) \times P(Q)$$

S(Q) is the **structure** factor

- *Inter*-particle interferences
- High concentrations (>5% vol.)
- Ordering/packing of particles

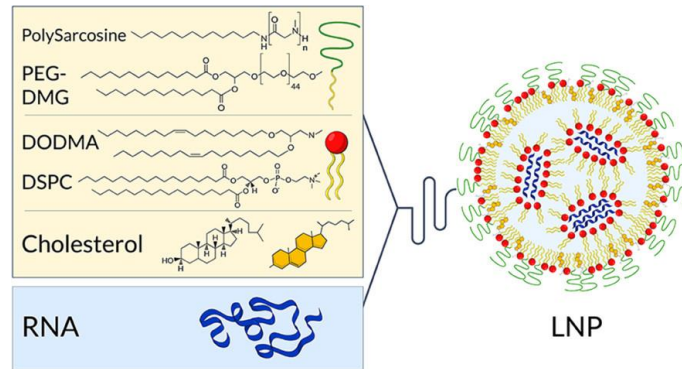
P(Q) is the **form** factor

- *Intra*-particle interferences
- Low concentration (dilute limit)
- Size/polydispersity/shape of particles



Pharmacy: Liposomes for RNA delivery

Pure mRNA degraded by enzymes in tissues
Hide it inside a liposome NP to get it into cells



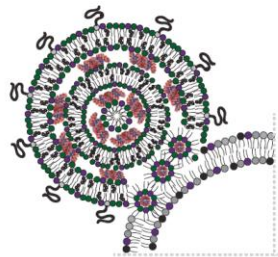
ACS Appl. Nano Mater. 2020, 3, 11, 10634–10645

Pfizer, Moderna COVID-19 mRNA vaccines

“transfection”

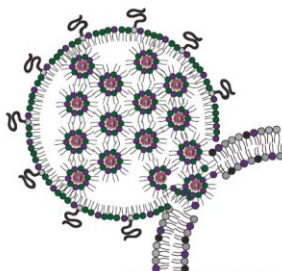
1.

2.



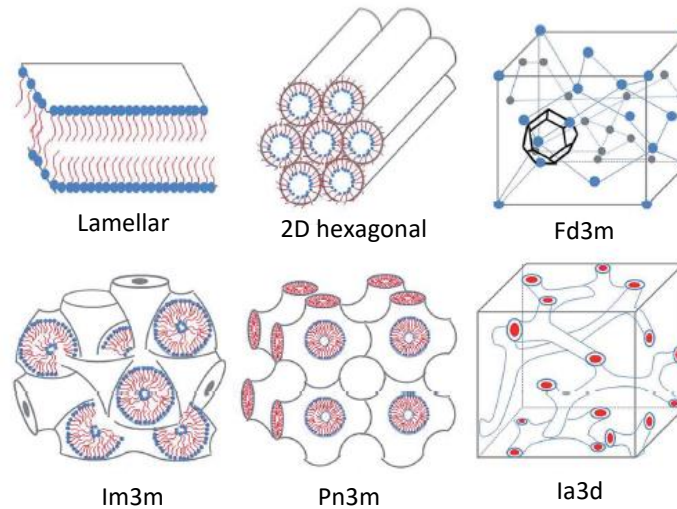
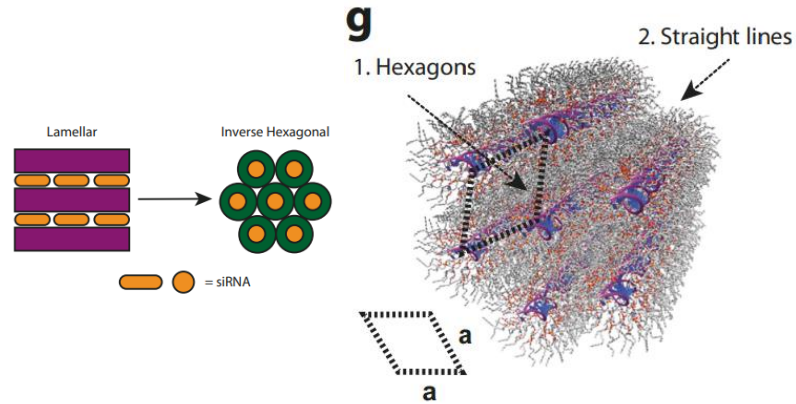
In situ L_α to H_{II} transition

Nature Comm. 15, 1303 (2024)



Pre-programmed H_{II}
remains intact

Lipid phase affects transfection efficiency

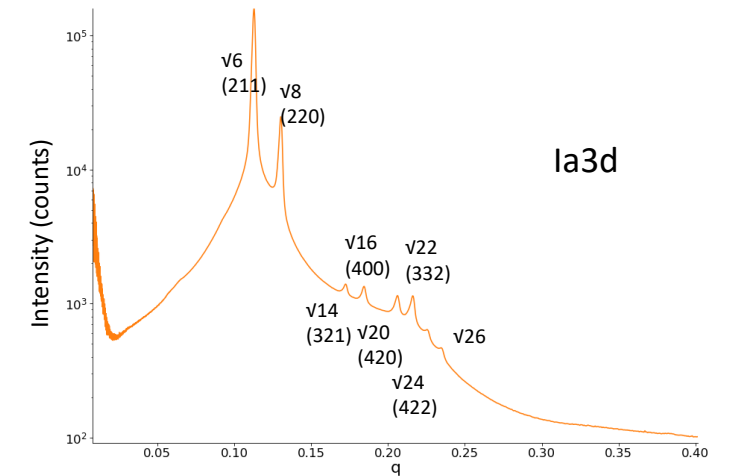
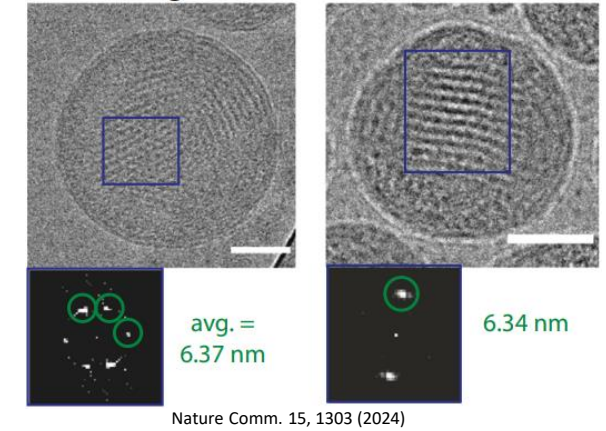


Y. Huang, S. Gui, RSC Adv., 2018, 8, 6978–6987

Cryo-TEM

Hexagons

Lines

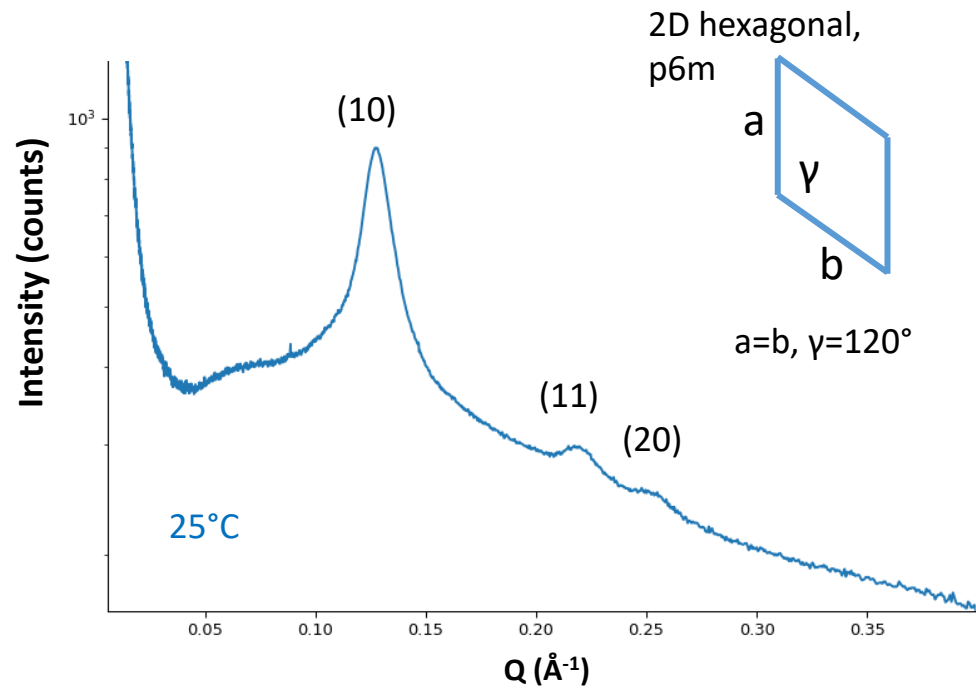


Types of LLC phase	SAXS
Lamellar phase	1 : 2 : 3 : 4
<i>Im3m</i>	$\sqrt{2} : \sqrt{4} : \sqrt{6} : \sqrt{8}$
<i>Pn3m</i>	$\sqrt{2} : \sqrt{3} : \sqrt{4} : \sqrt{6} : \sqrt{8} : \sqrt{9}$
<i>Ia3d</i>	$\sqrt{6} : \sqrt{8} : \sqrt{14} : \sqrt{16} : \sqrt{20} : \sqrt{22}$
<i>Fd3m</i>	$\sqrt{3} : \sqrt{8} : \sqrt{11} : \sqrt{16} : \sqrt{19}$
Reverse hexagonal	1 : $\sqrt{3}$: 2 : $\sqrt{7}$

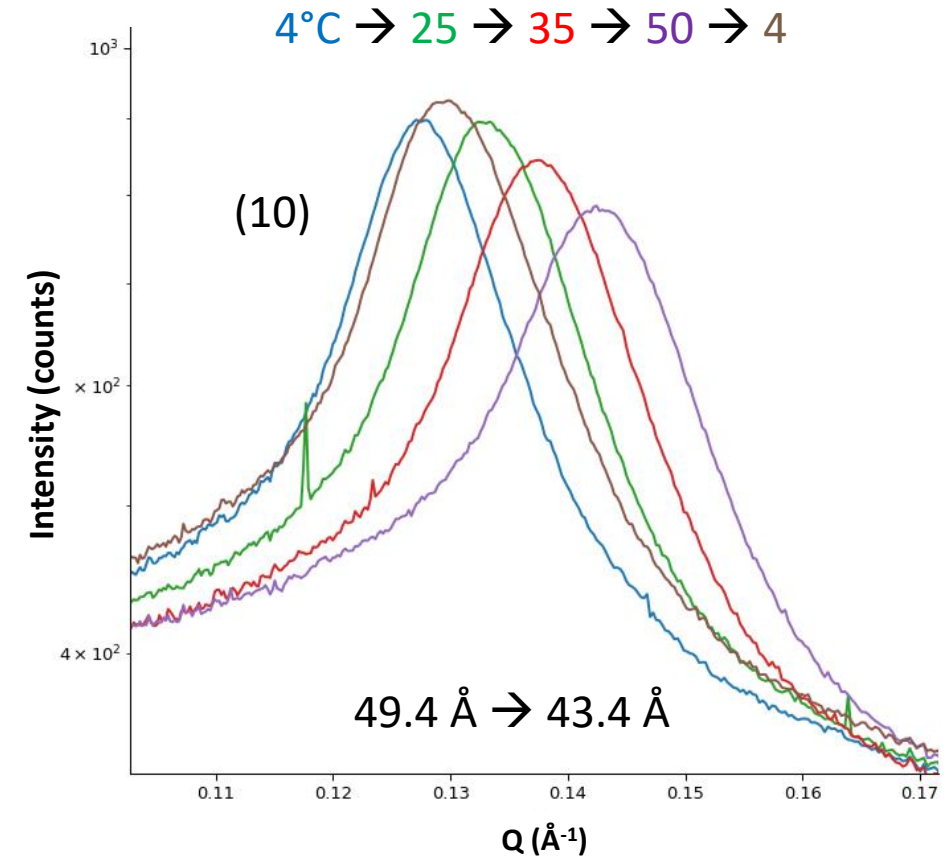
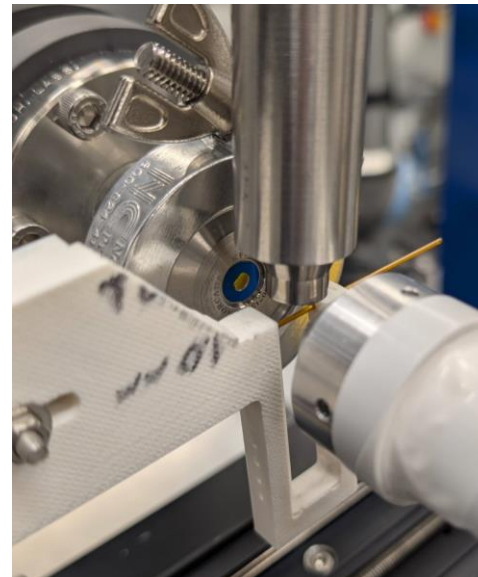
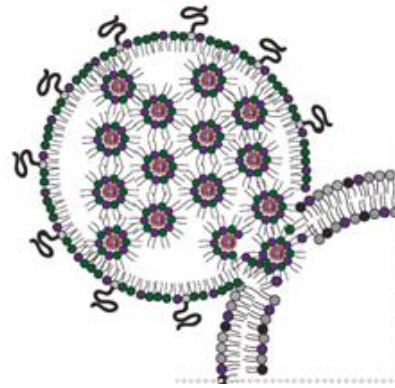
23

Pharmacy: Liposomes for RNA delivery

Naga Kola, Thakur Group, VIDO



What happens with temperature?



What is happening here?
Is radiation damage at play?

ARTICLE

Received 7 Aug 2014 | Accepted 25 Nov 2014 | Published 7 Jan 2015

DOI: 10.1038/ncomms5948

OPEN

Volume and porosity thermal regulation in lipid mesophases by coupling mobile ligands to soft membranes

Lucia Parolini¹, Bortolo M. Moggetti², Jurij Kotar¹, Erika Eiser³, Pietro Cicuta¹ & Lorenzo Di Michele¹

Nat. Comm. 6, 5948 (2015)

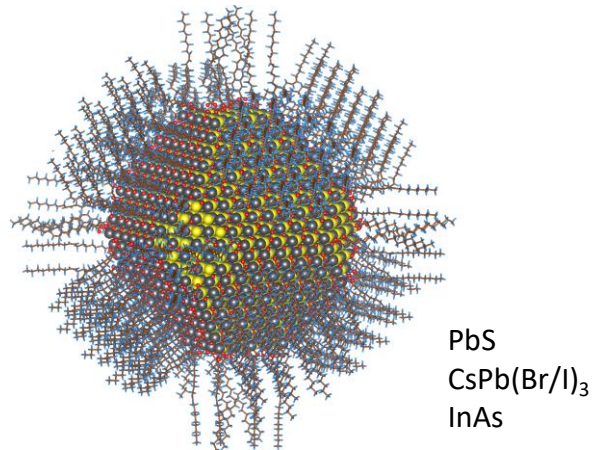


Canadian
Light
Source

Centre canadien
de rayonnement
synchrotron

Energy materials: Quantum dot solar cells

Sargent group
Elec. & Computer Engineering
U. Toronto/Northwestern U.

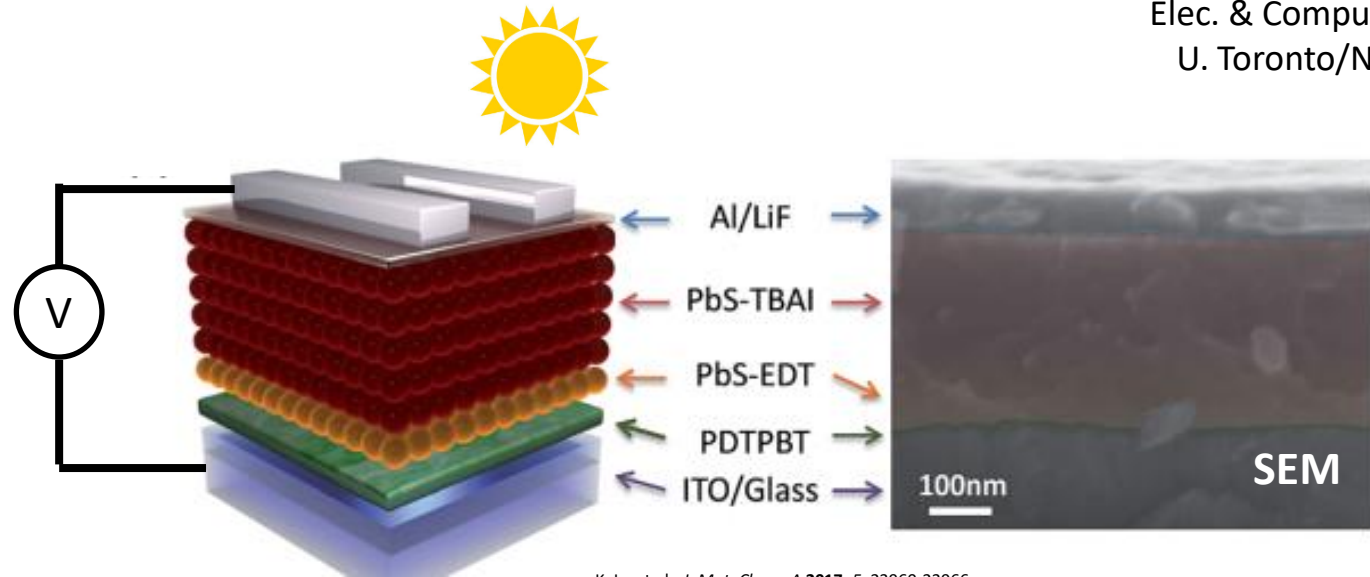


<https://www.olcf.ornl.gov/2015/05/05/demystifying-quantum-dot-conundrums/>

Efficient exciton generation
Size-based, tune-able band gap

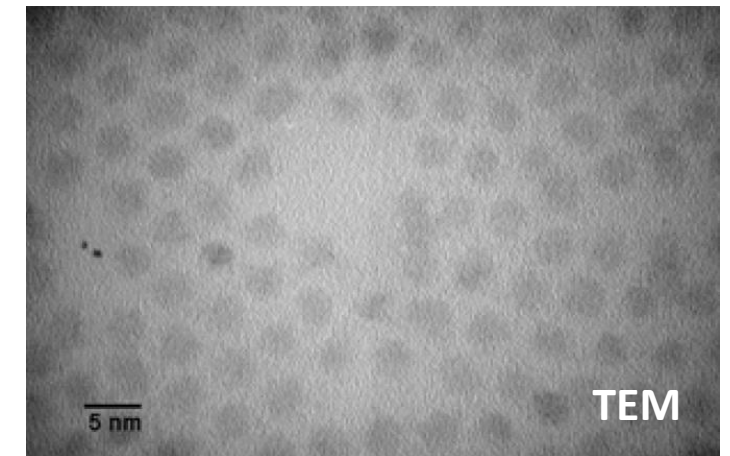
- 18.1% record efficiency in 2022
Performance improved by tuning:
- Particle size distribution or polydispersity
 - How particles pack and the interparticle distance

- Control the size and packing by:
- Ligand exchanges cycles, centrifuge to remove largest QDs
 - Rinsing steps during buildup
 - Spin speeds, solvents, additives



K. Lu et al., *J. Mat. Chem. A* **2017**, 5, 23960-23966

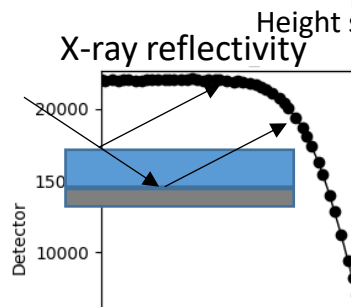
QD films deposited layer-by-layer using spin coating
Top electrodes deposited using thermal evaporation



M. Yuan et al., *Adv. Mater.* **2014**, 26, 3513-3519



Canadian
Light
Source
Centre canadien
de rayonnement
synchrotron

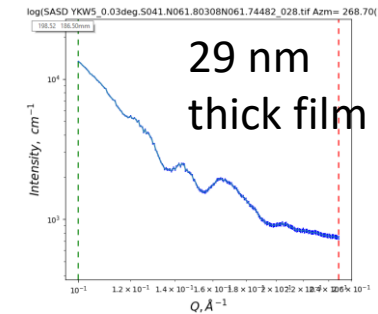
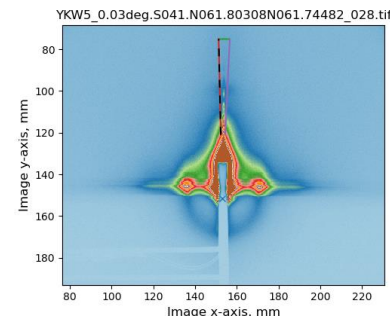
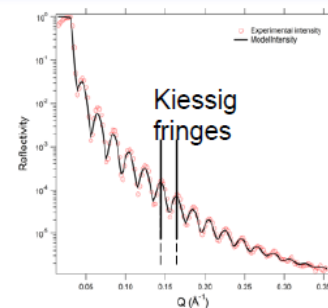


Simple estimation of film thickness

Fringes with uniform spacing

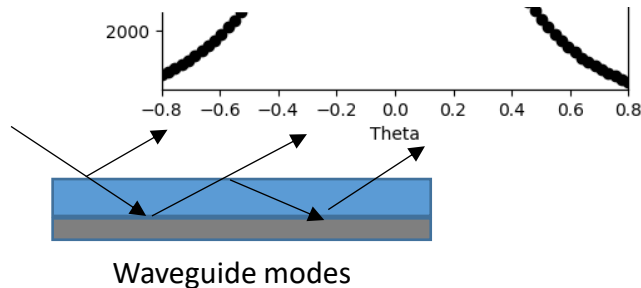
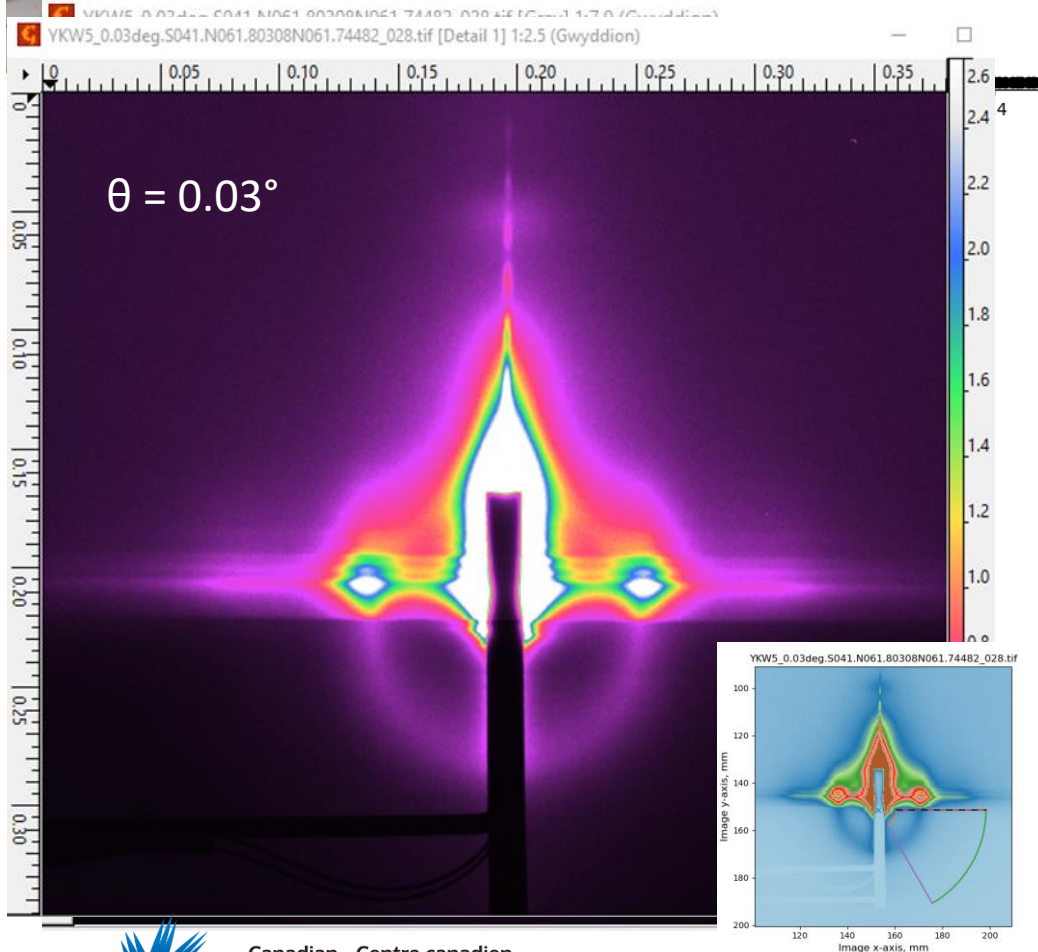
Thickness of the layer :

$$t = \frac{2\pi}{\Delta q_z}$$

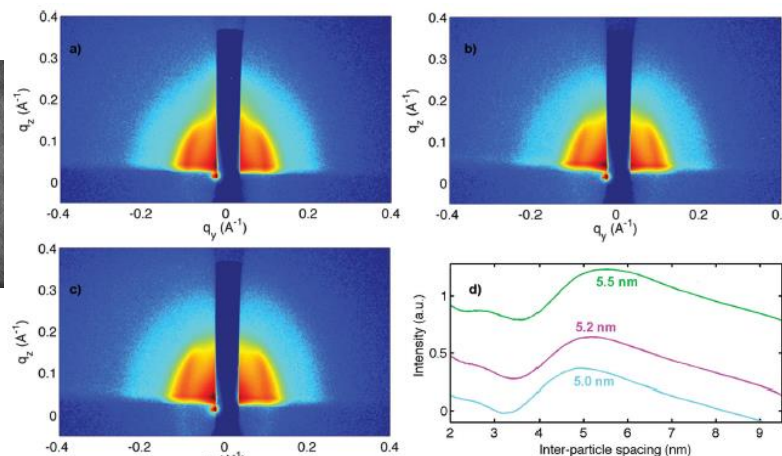
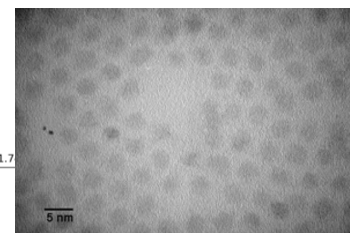
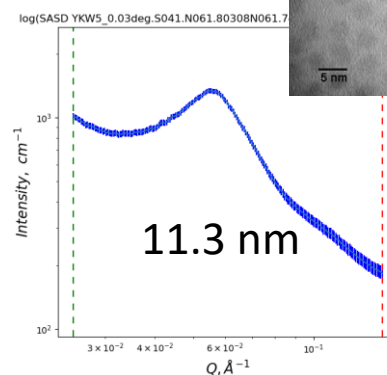


- Reflectivity gives film thickness

- Position gives interparticle distance
- Peak width gives size distribution
- Average over relatively huge area



- Indicates very flat, high quality film
- Adjusting grazing angle can give depth information



M. Yuan et al., Adv. Mater. 2014, 26, 3513–3519



Canadian
Light
Source

Centre canadien
de rayonnement
synchrotron

Origins of the SAXS signal

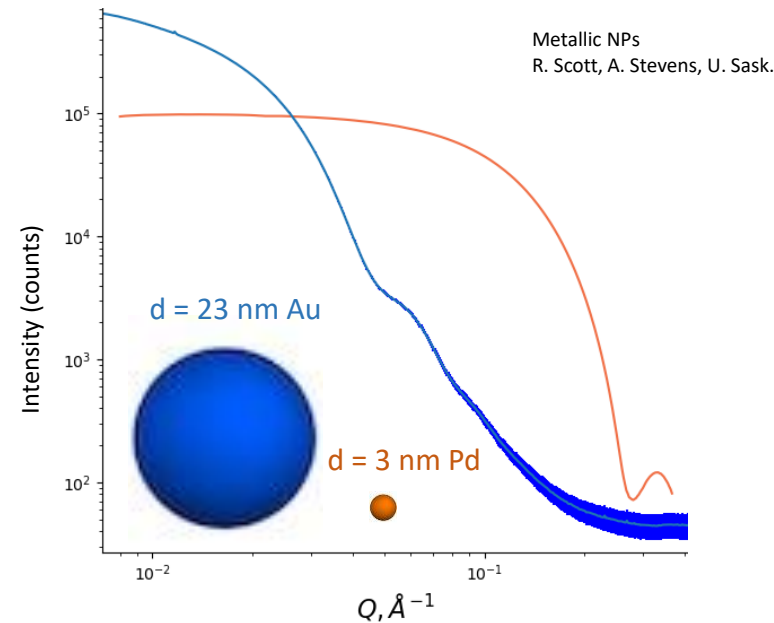
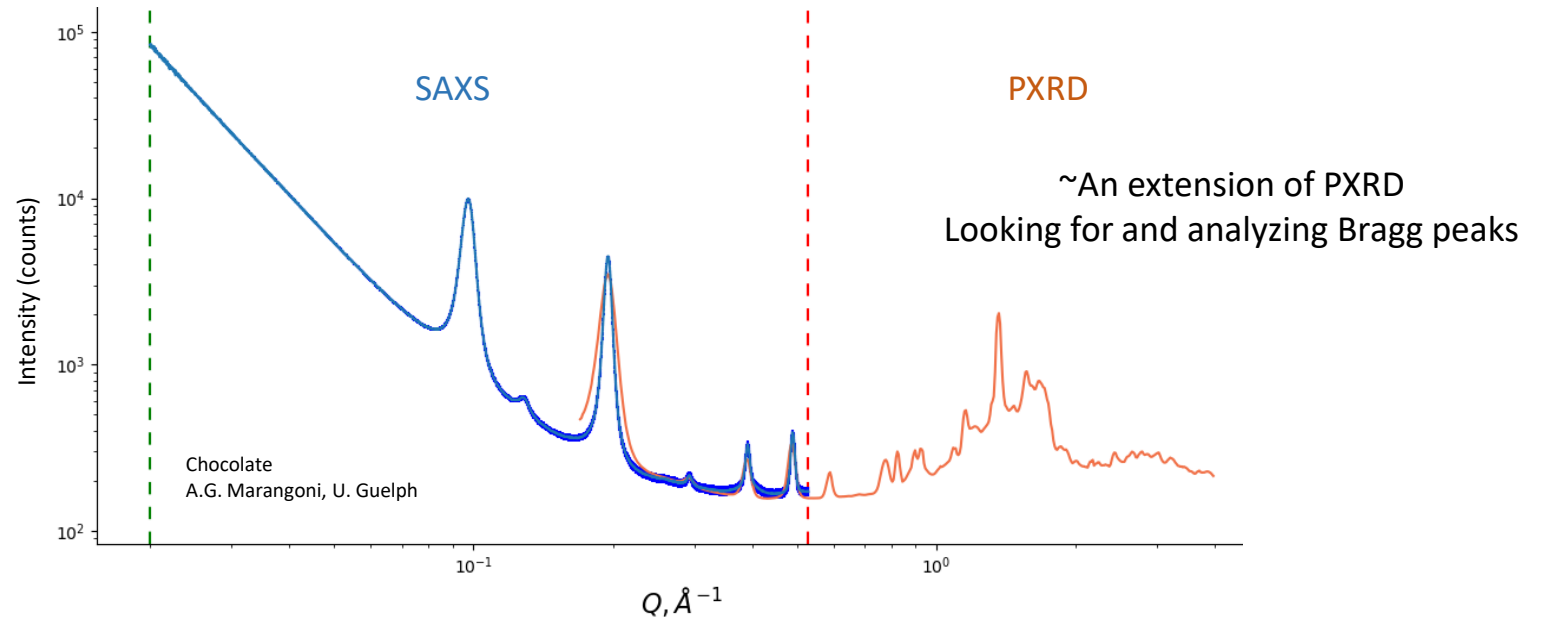
$$I(Q) = S(Q) \times P(Q)$$

S(Q) is the **structure** factor

- *Inter*-particle interferences
- High concentrations (>5% vol.)
- Ordering/packing of particles

P(Q) is the **form** factor

- *Intra*-particle interferences
- Low concentration (dilute limit)
- Size/polydispersity/shape of particles



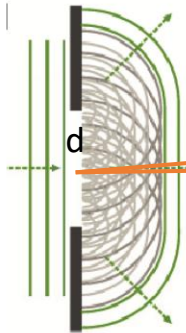
Analyzing large regions or
the whole curve

Form factor, $P(Q)$

- Intra-particle interferences
- Low concentration (dilute limit)
- Size, shape, polydispersity of particles

Monochromatic beam

$\lambda = 532 \text{ nm}$

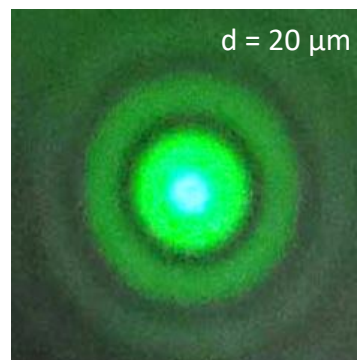
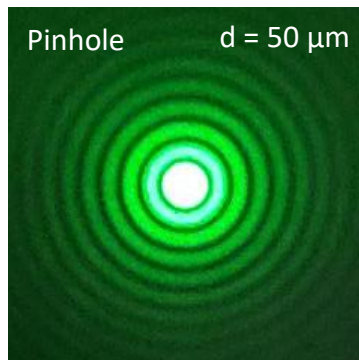
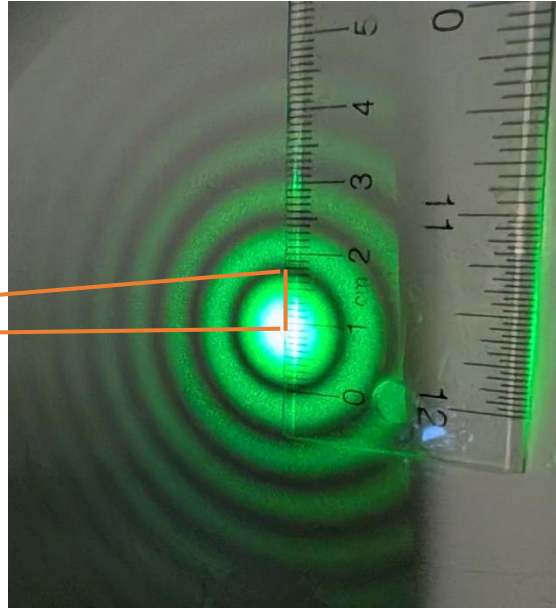


1st minima observed at

$$\frac{\lambda}{\sin\theta} = d$$

$\lambda = 532 \text{ nm}$
 $D = 1.0 \text{ m}$

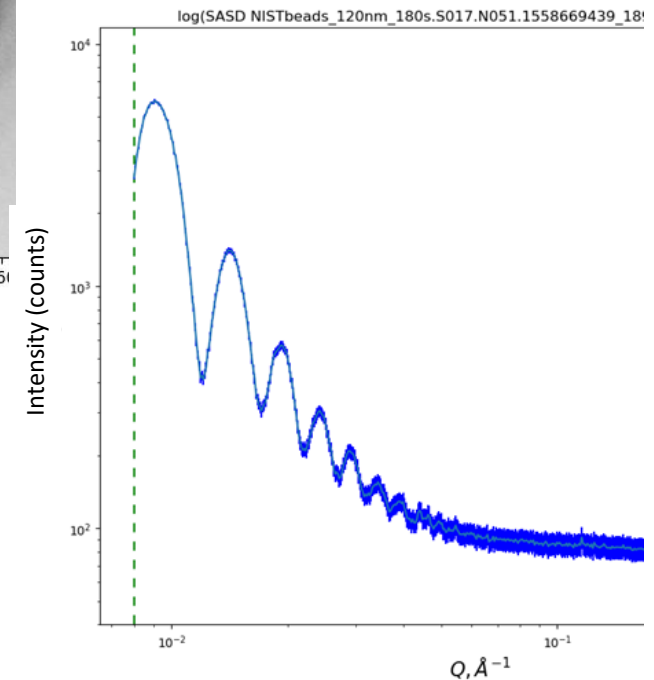
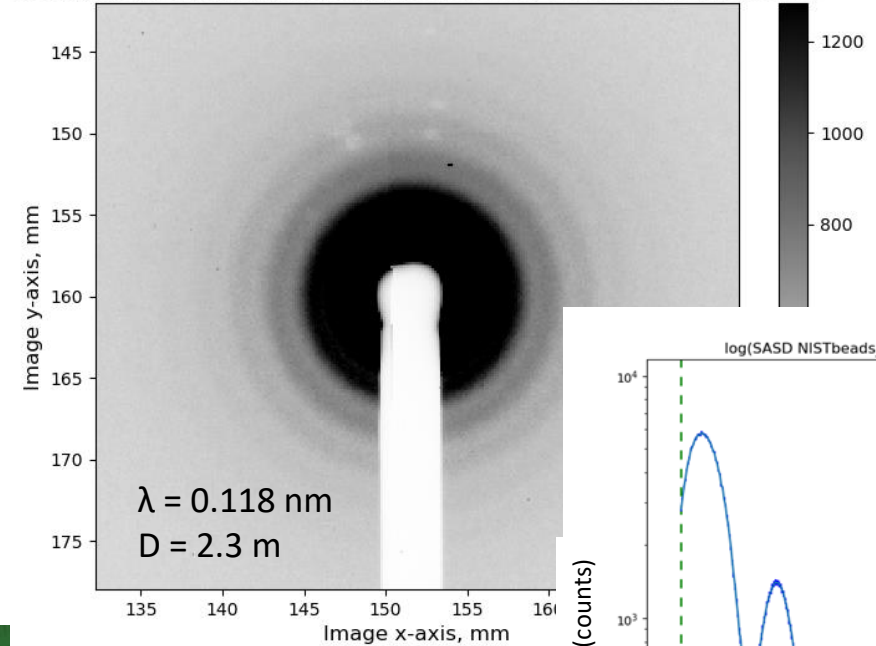
Airy pattern



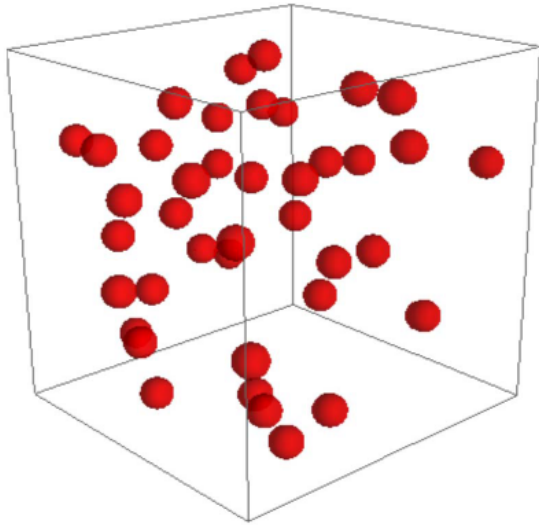
120 nm diameter PS spheres in water

Pure, monodisperse, and dilute
(~1 – 10 mg/mL) - no interparticle interactions

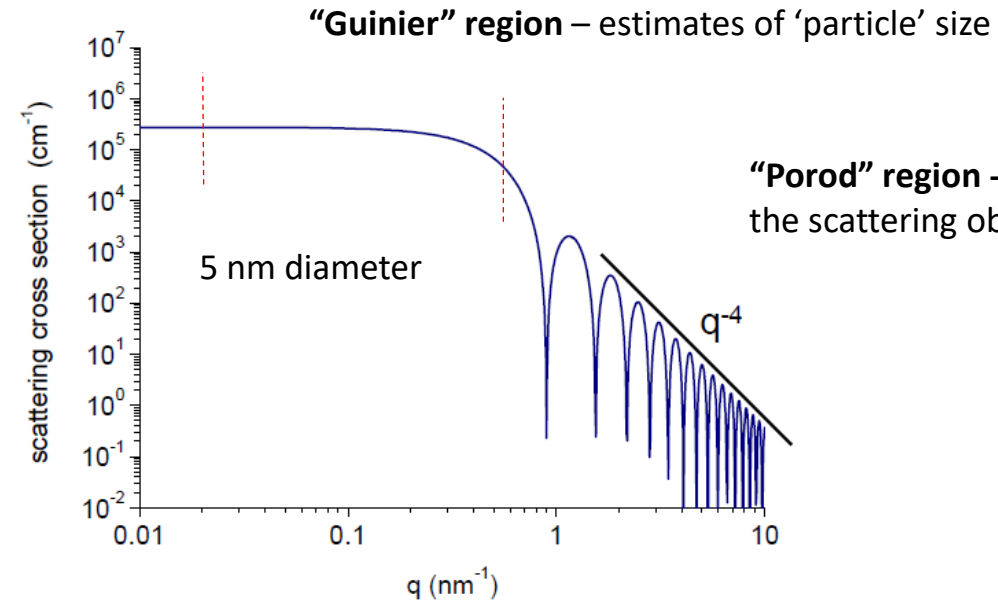
120nm_NIST_std_quartz_300s.S014.N051.4056907486906665264173_365.tif



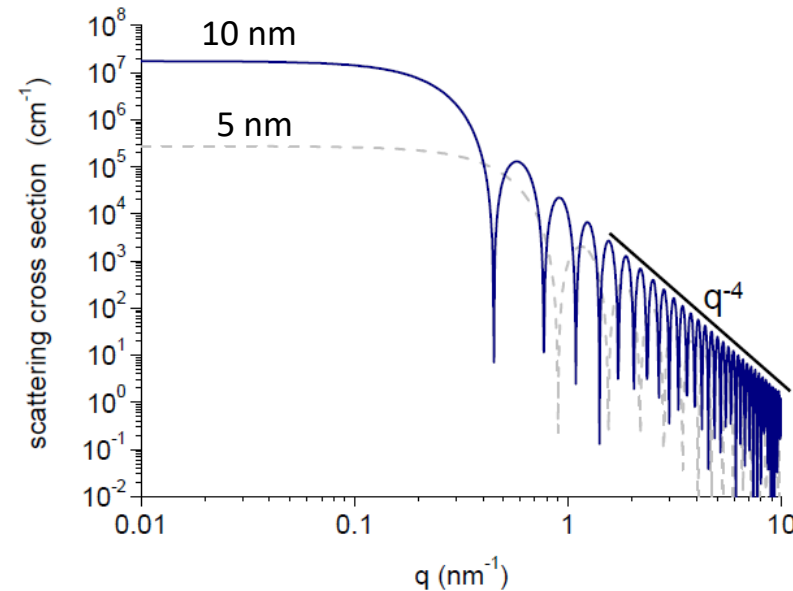
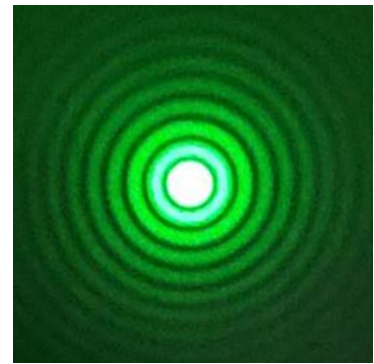
Monodisperse, dilute spheres in solution



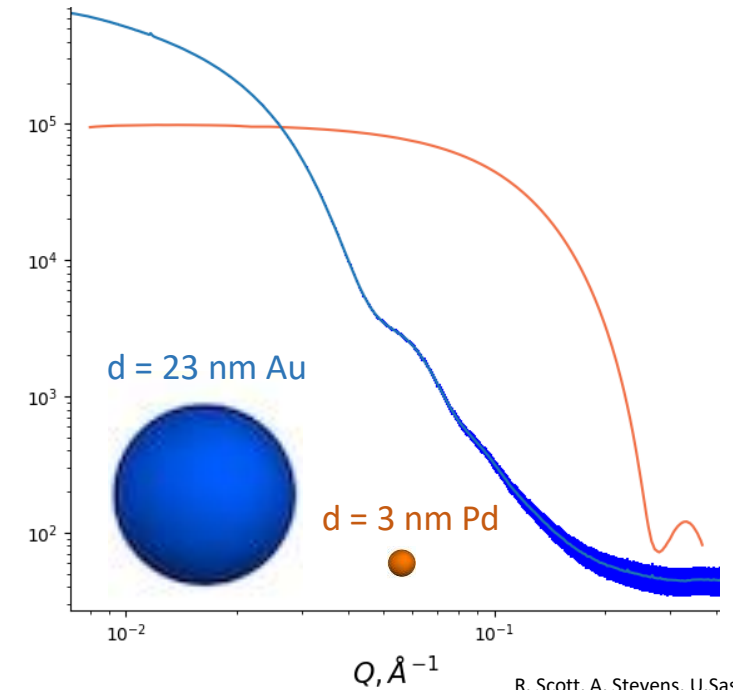
$$P(q) = \left[3 \frac{\sin(qR) - qR \cos(qR)}{(qR)^3} \right]^2$$



“Porod” region - Clues to the nature of the scattering objects



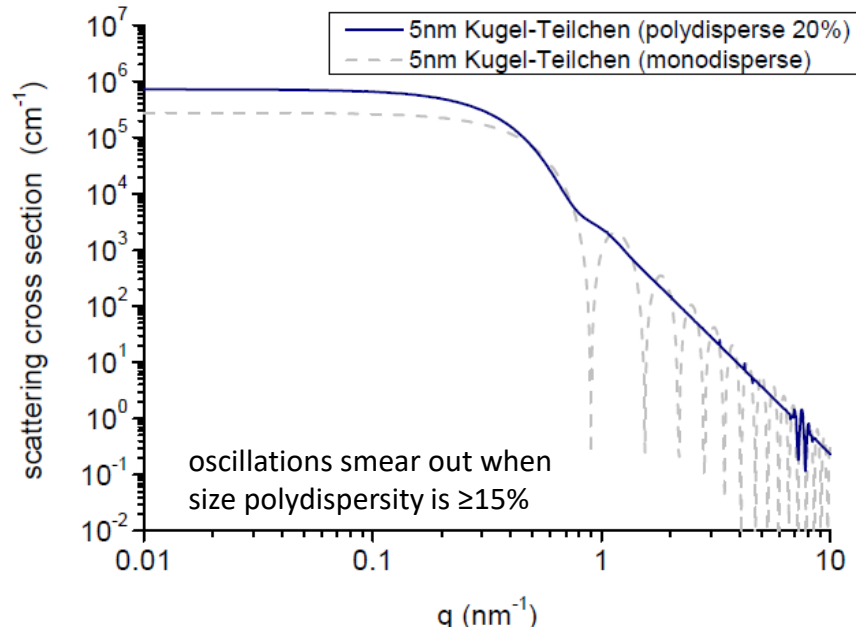
Real samples, spherical metallic NPs



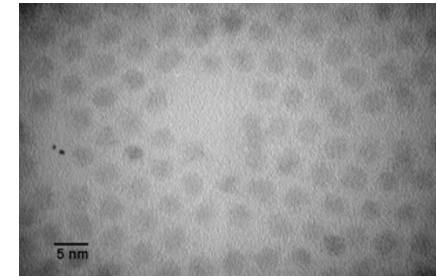
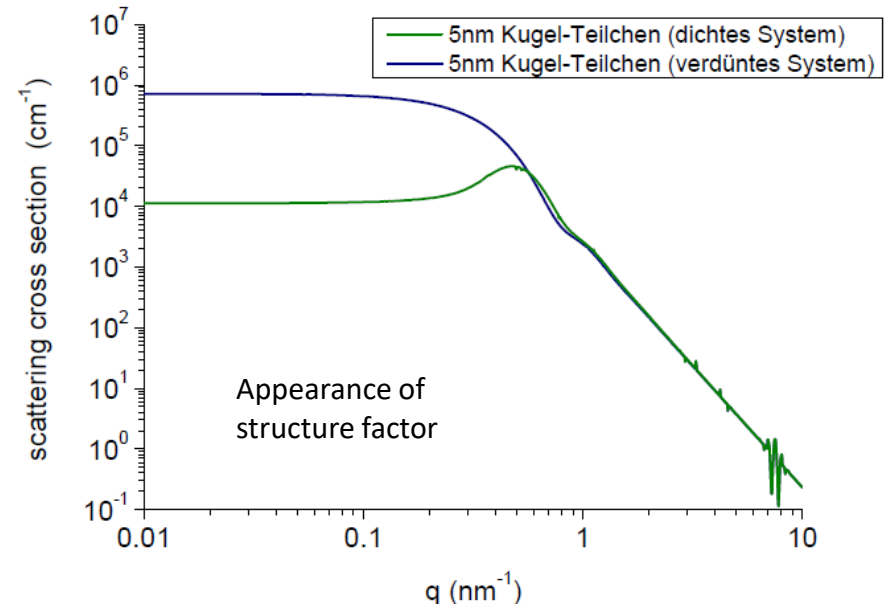
R. Scott, A. Stevens, U.Sask

Complications...

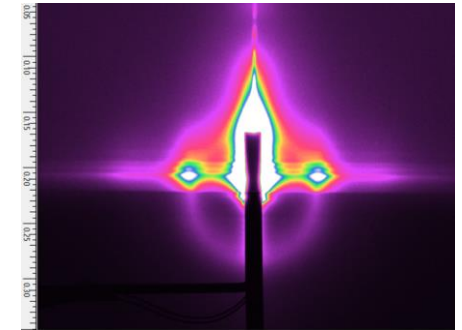
Polydispersity?



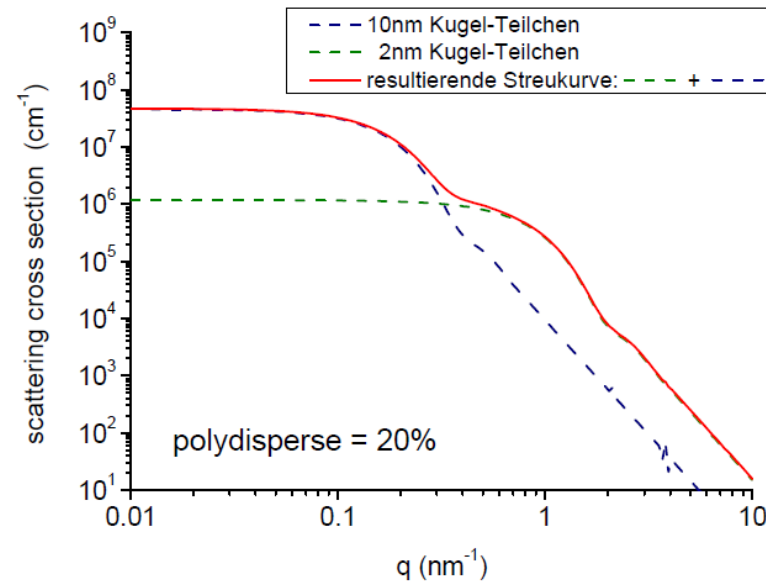
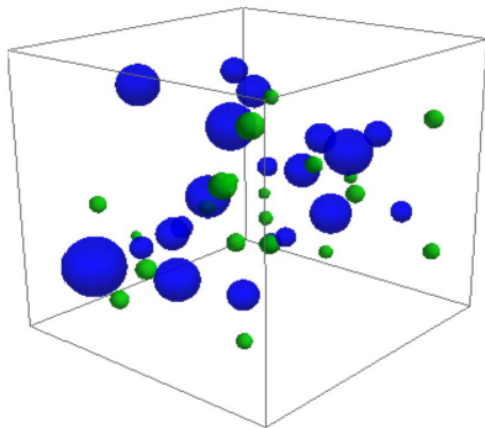
Too concentrated?



M. Yuan et al., Adv. Mater. 2014, 26, 3513–3519



Several populations?



SAXS data interpretation can be ambiguous, especially if the sample is not pure, not dilute and/or not monodisperse

'Particle' size: Guinier plots

Guinier's approximation:

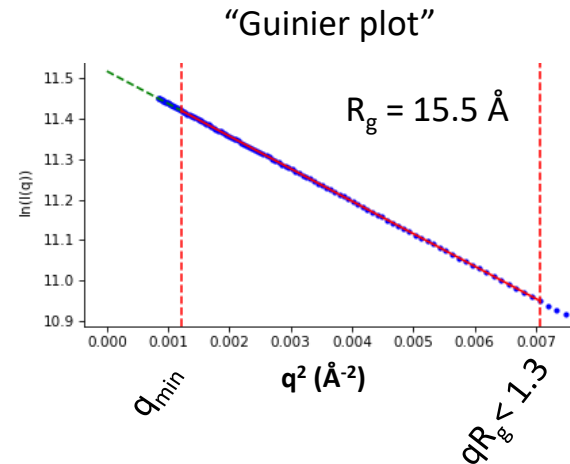
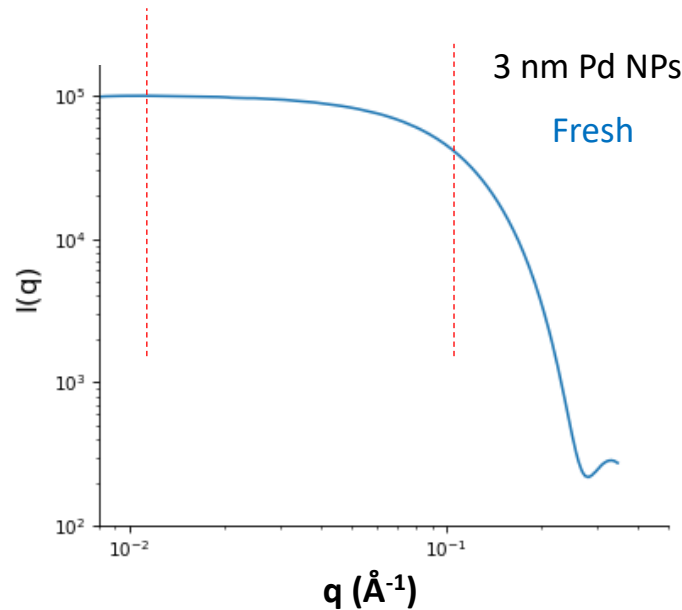
$$I(q) \approx I(0)e^{-q^2 R_g^2/3}$$

R_g : Radius of gyration
 $I(0)$: Intensity at $q=0$

R_g is the square root of the average squared distance of each scatterer from the particle center

RMS distance of the objects parts from its center of mass

➤ Particle radius



Slope: $-R_g^2/3$
 Intercept: $\ln I_0$

I_0 contains information about molecular weight of the particle

Sphere		Thin rod	Thin disc	Cylinder	
$R_g^2 = \frac{3}{5} R^2$		$R_g^2 = \frac{L^2}{12}$	$R_g^2 = \frac{R^2}{2}$	$R_g^2 = \frac{d^2}{8} + \frac{L^2}{12}$	

How big of an object?

q range required

Globular, disc: $q_{\min} * R_g < 1.0$ to ~ 1.3
 Linear: $q_{\min} * R_g < 0.65$ to ~ 1.0

If $q_{\min} = 0.005 \text{ Å}^{-1}$

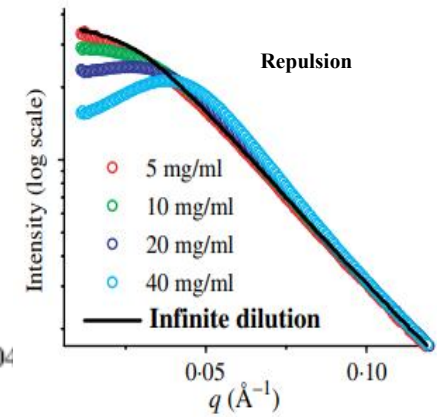
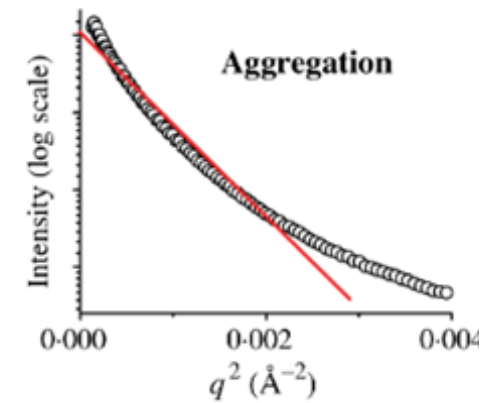
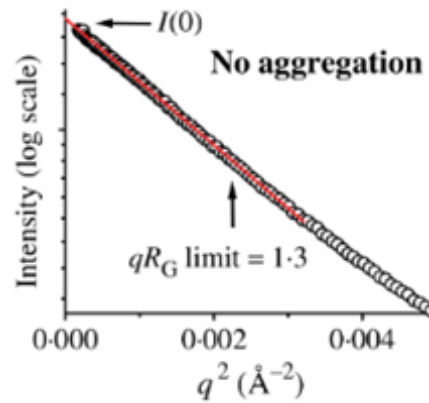
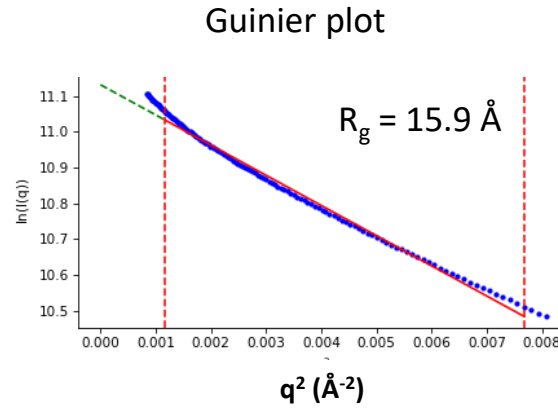
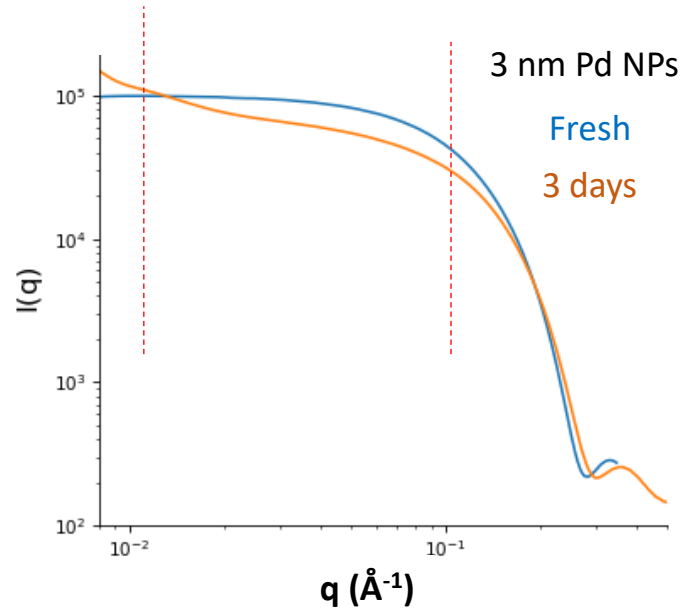
$R_{g \text{ max}} = 20 \text{ nm}$
 $R_{g \text{ max}} = 13 \text{ nm}$

'Particle' size: Guinier plots

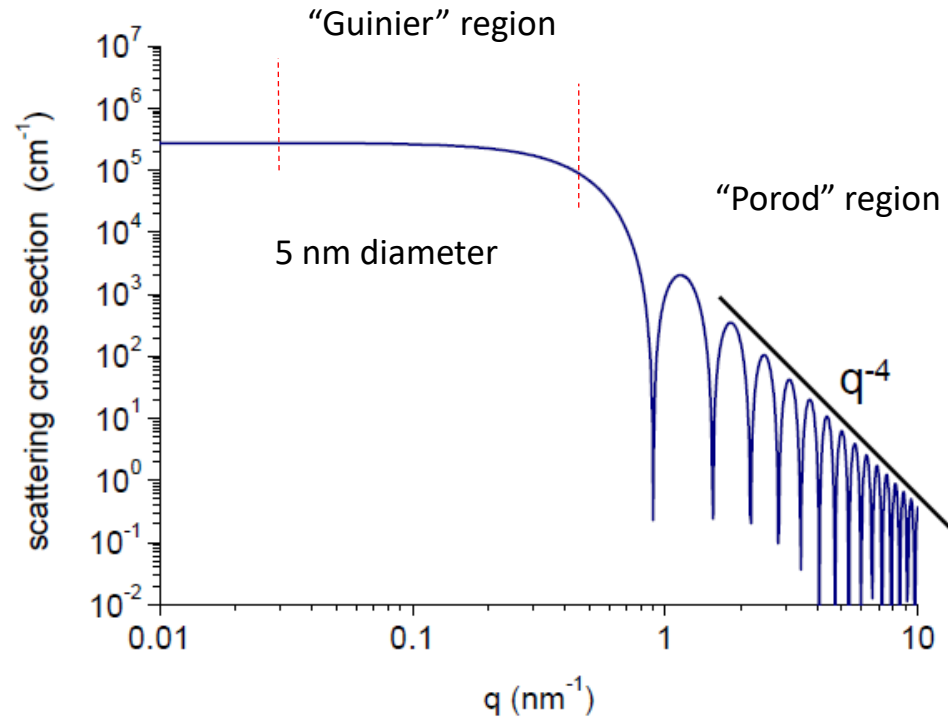
Guinier's approximation:

$$I(q) \approx I(0)e^{-q^2 R_g^2/3}$$

➤ Sample quality: non-linearity can indicate aggregation, repulsion, radiation damage



Porod exponent



Scattering intensity in this region arises from surfaces or interfaces, and is related to the particle surface, shape, and/or flexibility

Porod's law states that at high q scattering intensity decays following a power law,

$$I(q) \propto q^{-D}$$

D is the Porod exponent, and can give some evidence of particle shape, surface, and/or flexibility

4

Hard sphere/sharp interfaces, 3D object, or a very polydisperse sample

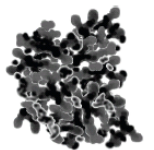


3 – 4

A rougher surface, 3D object

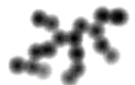
2 – 3

Thin disc, or sheet, 2D object, a more open or porous microstructure



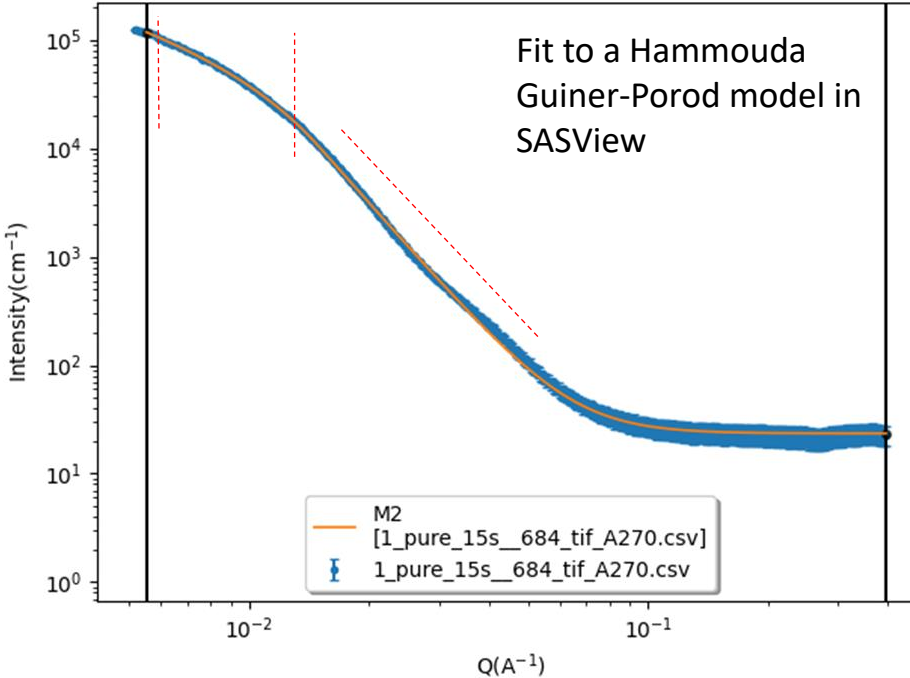
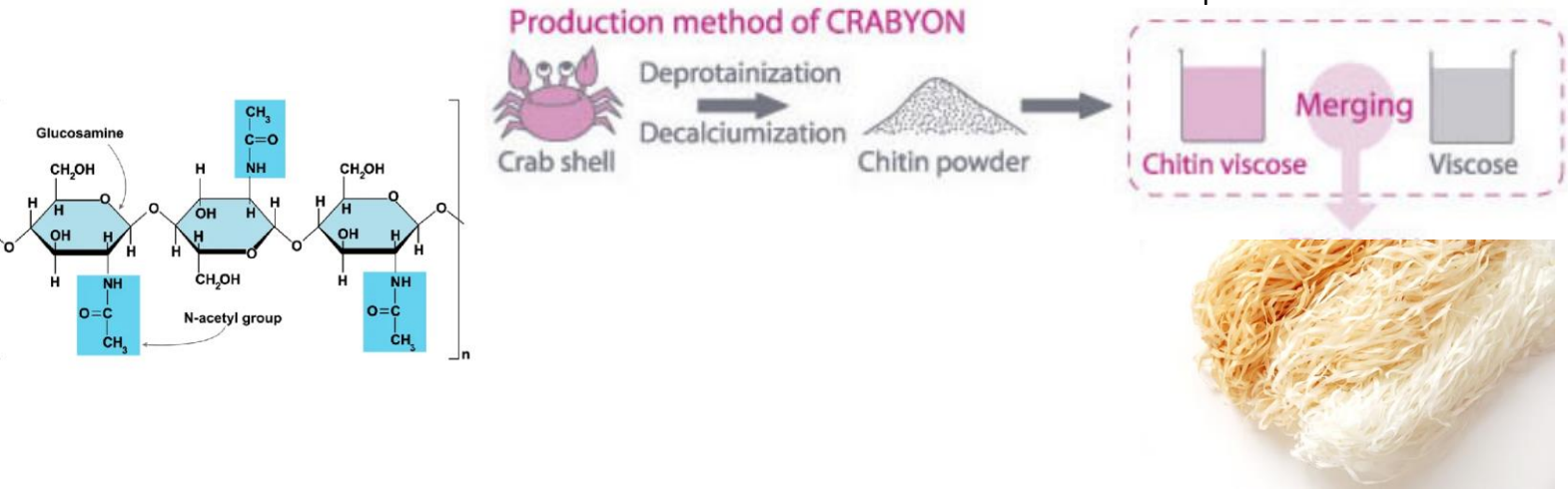
1 – 2

Needle-like, stiff rods, a fully extended 1D structure



Biomaterialization phenomena in biofilaments based on nanochitin

Tianyu Guo, Rojas group, UBC



Chitin is a polysaccharide, found in the exoskeletons of crustaceans and insects.

It can be spun into fibers and thread or made into textiles. These materials could replace synthetic fibers like nylon and polyester made from petroleum and tend to shed microplastics.

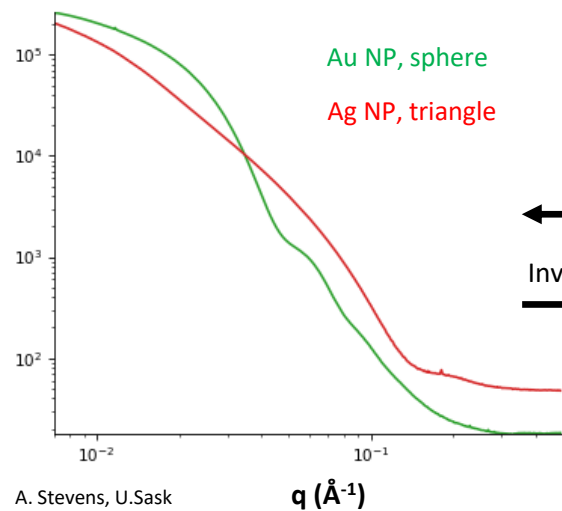
Moisture and humidity weakens the mechanical strength and durability.

Nanochitin composite, bio mineralized with calcium carbonate (CaCO_3) nanocrystals.

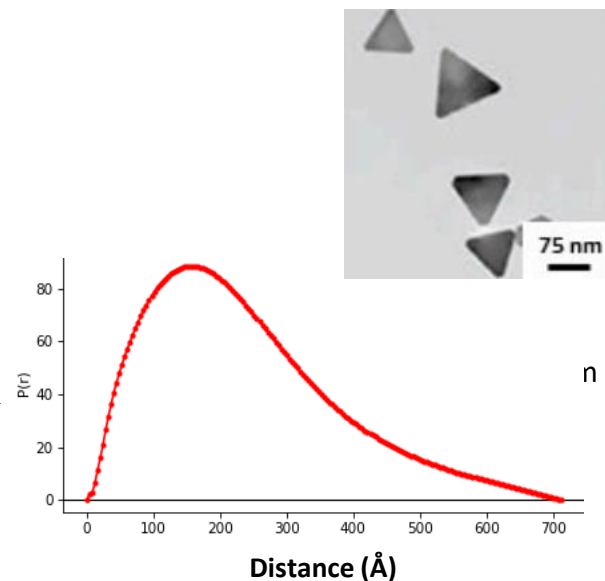
Use SAXS to gain a deeper understanding of the water-resistance mechanism of our manufactured bio-fibers.

- CaCO_3 is reducing R_g (fibers are compacting? shrinking? stiffening?)
- Porod exponent indicates smooth surfaces, not swollen
- Nano-chitin is likely tightly coiled in solution

Shape information: Pair distance distribution function (PDDF)

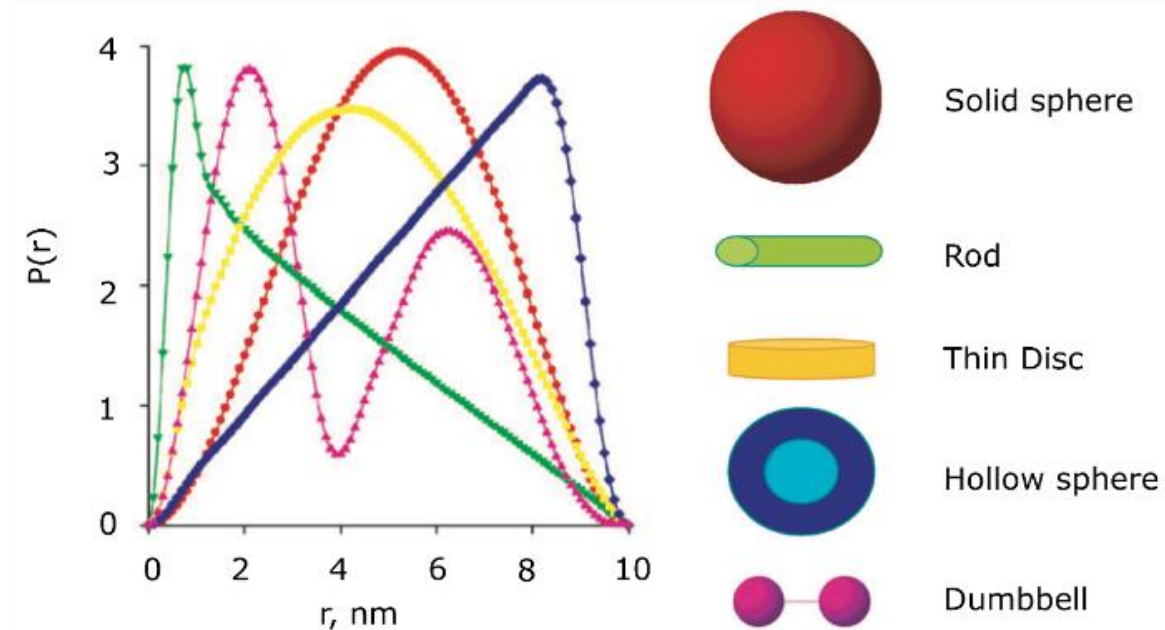


Reciprocal space intensity



Real space distance frequency

Histogram of atom-atom distances within a single particle, weighted by respective electron densities



D. Svergun, M.H.J. Koch, Rep. Prog. Phys. 66 1735 (2003)

$D_{\max}$ – **Largest interatomic distance** in the scattering particle

Shape of the $P(r)$ function is related to the **shape** of the particle, with low spatial resolution

q range required

$$q_{\min} \leq \pi / D_{\max} \quad (\pi/0.005 \text{ \AA}^{-1} = 62 \text{ nm})$$

$$q_{\max} \geq 2\pi / D_{\max}$$

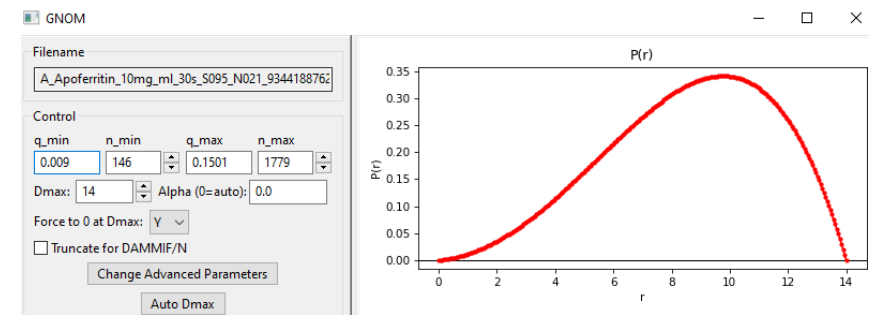
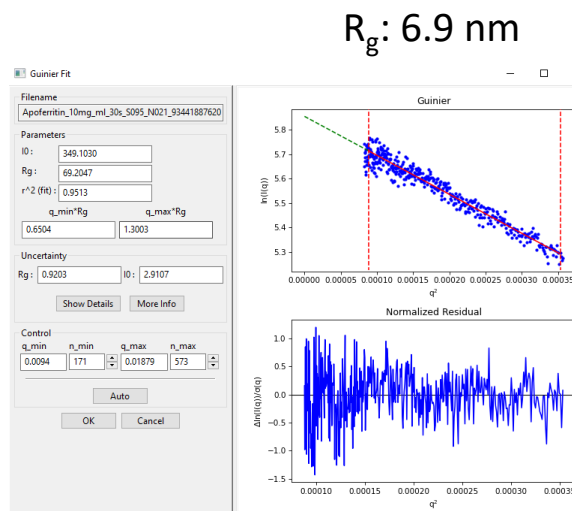
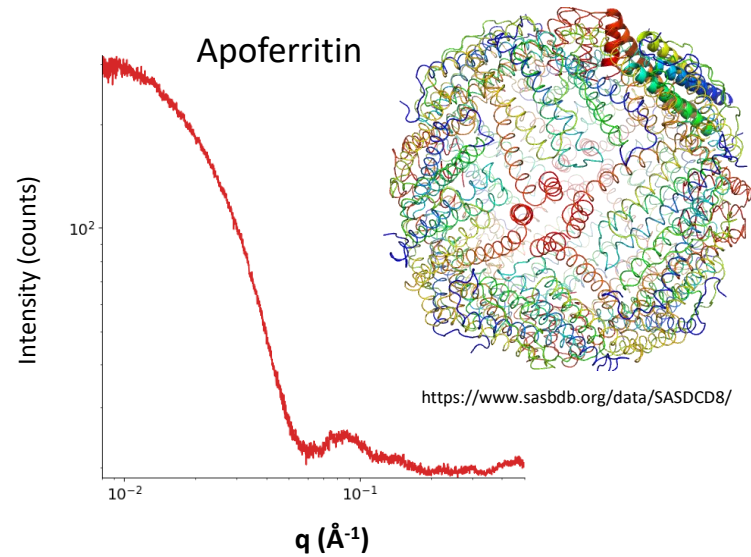
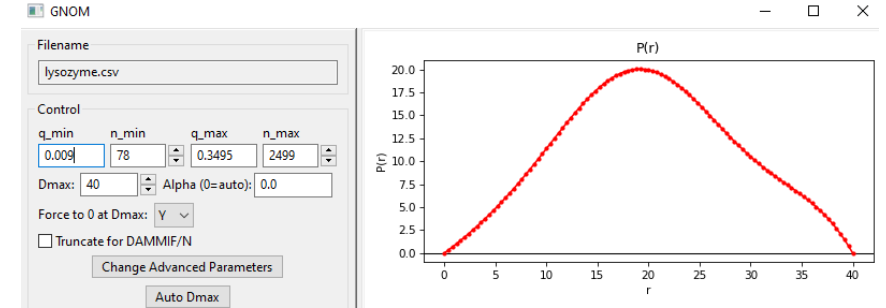
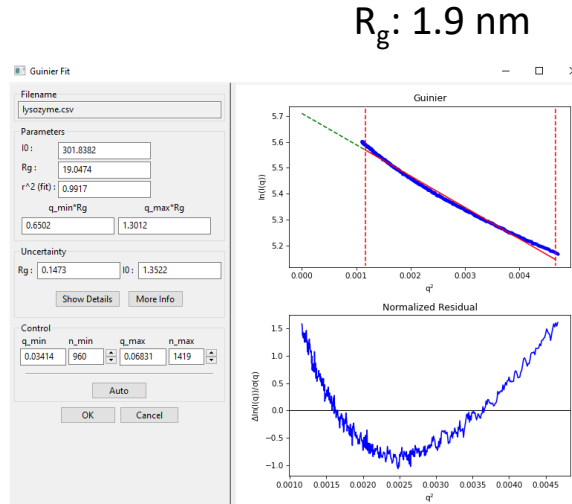
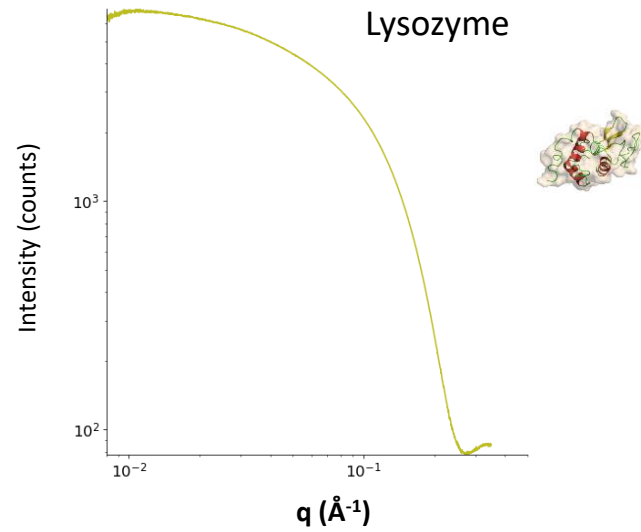
D. Putnam et al. Quart. Rev. Biophys. 40, 191-285 (2007)



Canadian
Light
Source

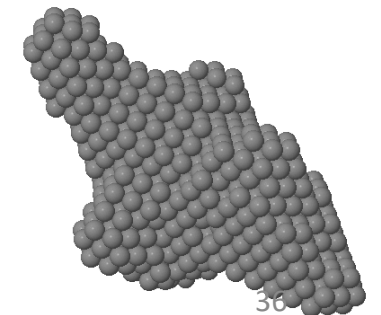
Centre canadien
de rayonnement
synchrotron

Proteins in solution



Low resolution protein structure

ATSAS package
DAMMIF/DAMMIN
GASBOR



Summary

The CLS has a SAXS endstation available... Call for General User Proposals opens **July 22, closes August 19, 2026**

Probe nano-scale dimensions and ordering (1 - 150 nm) with X-rays by measuring elastic scattering at small angles

Can learn about packing, particle size, and particle shape

Limited structural information, and data analysis can be ambiguous with numerous interpretations

Complementary information, that should be supported with supplementary techniques such as microscopy



Further reading

Basic SAXS concepts

H. Schnablegger, Y. Singh, “The SAXS Guide: Getting Acquainted With the Principles”, Anton Parr, Austria. (2017)

Making a good measurement

B.R. Pauw, “Everything SAXS: small-angle scattering pattern collection and correction” J. Phys.: Condens. Matter 25 (2013) 383201

Basic data work up

J.B. Hopkins, R.E. Gillilan, S. Skou
https://bioxtas-raw.readthedocs.io/en/latest/saxs_tutorial.html

BioSAXS, data workup, introduction to structure modelling

C.D. Putnam, M. Hammel, G.L. Hura, J.A. Tainer, “X-ray solution scattering (SAXS) combined with crystallography and computation: defining accurate macromolecular structures, conformations and assemblies in solution” Quarterly Reviews of Biophysics 40, 3 (2007), pp. 191–285.



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Hands on SAXS data analysis

Update CPDW19 tutorials here: https://drive.google.com/drive/folders/1x1noj2g9FmPB5CQIF7ISQoMC6g_W6Fn4?usp=sharing

Data reduction

GSAS-II

- Detector calibration
- Masking
- Subtract background
- Multi-image pixel mask search
- Integration

Data manipulation

BioXTAS RAW

- Merge
- Crop
- Convert q scale from $\text{\AA}^{-1}$ to nm^{-1}

Data analysis

BioXTAS RAW

- Guinier
- PDDF

SASView

- Fitting to models
- Power law

