

# Small X-ray scattering: problems and solutions

Hatem M. Titi

McGill University

Canadian Powder Diffraction Workshop 18 (CPDW18)



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MCGILL CHEMISTRY  
CHARACTERIZATION

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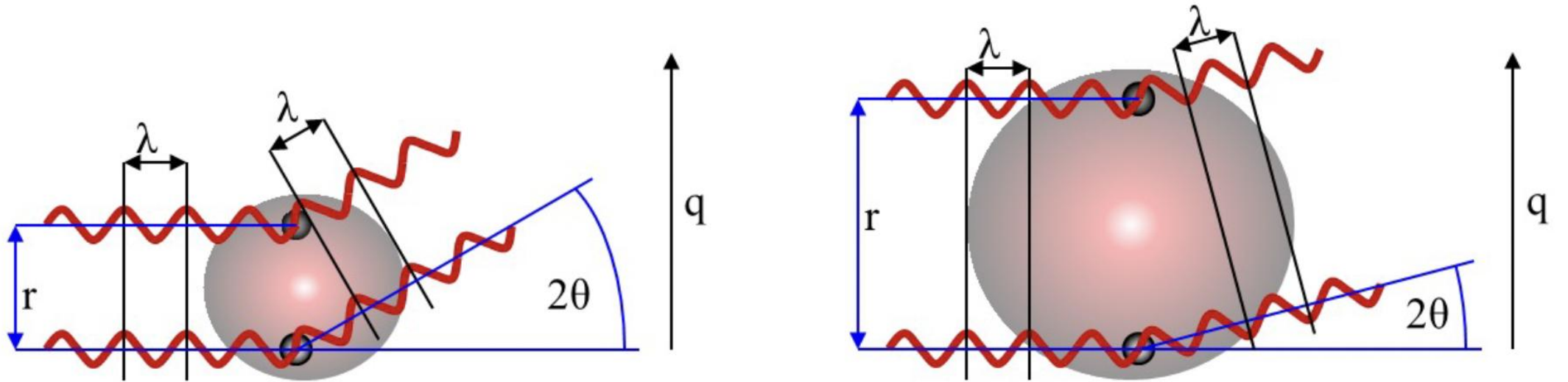


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X-RAY  
DIFFRACTION

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# Why SAXS?

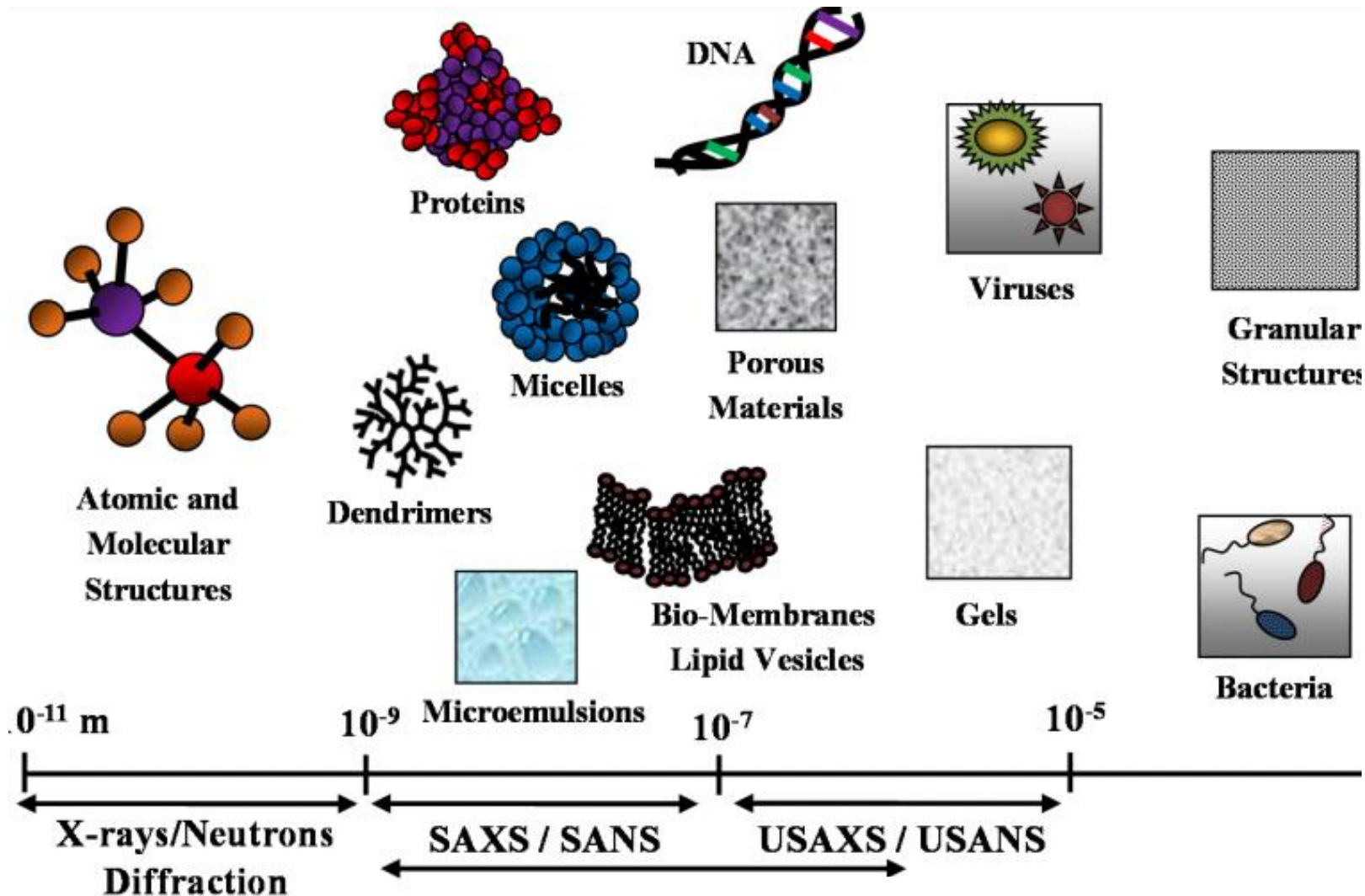


Small size  $\Leftrightarrow$  Large angle

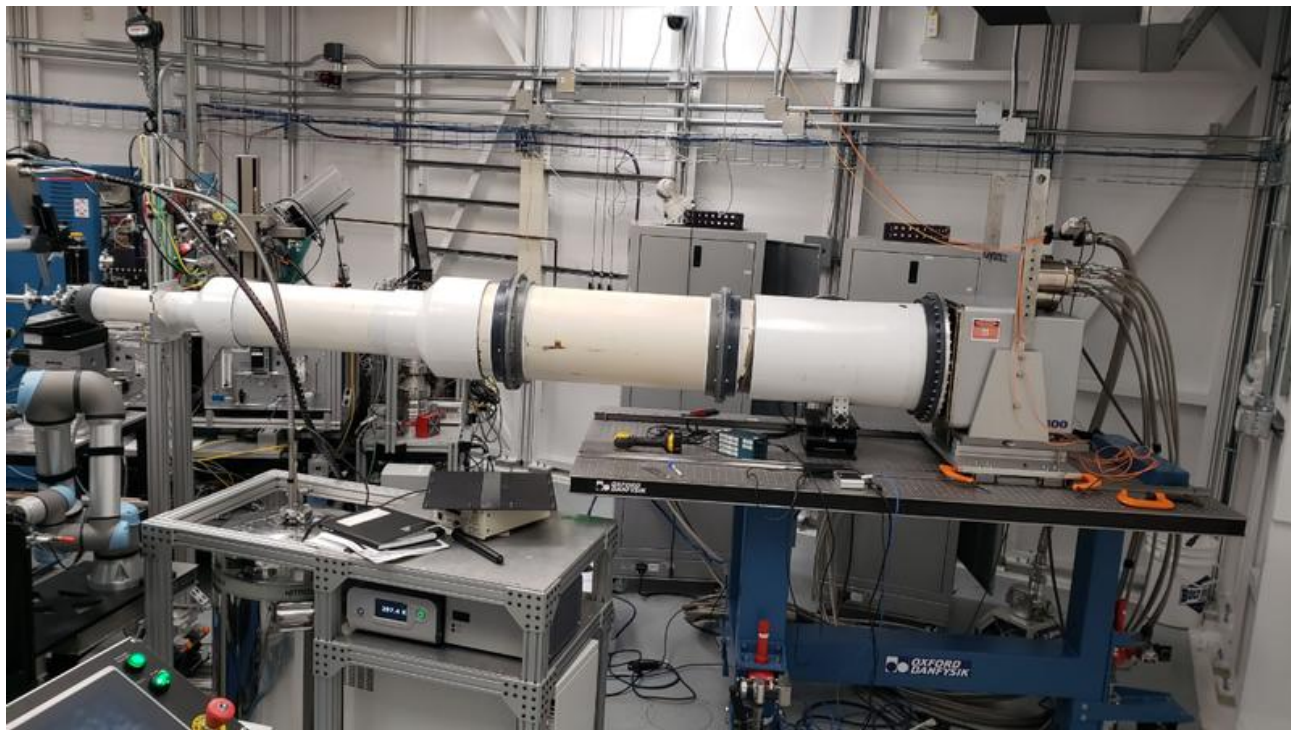
Large size  $\Leftrightarrow$  Small angle

Scattering for two points of a wave front on small and large particles.

# X-ray diffraction and scattering

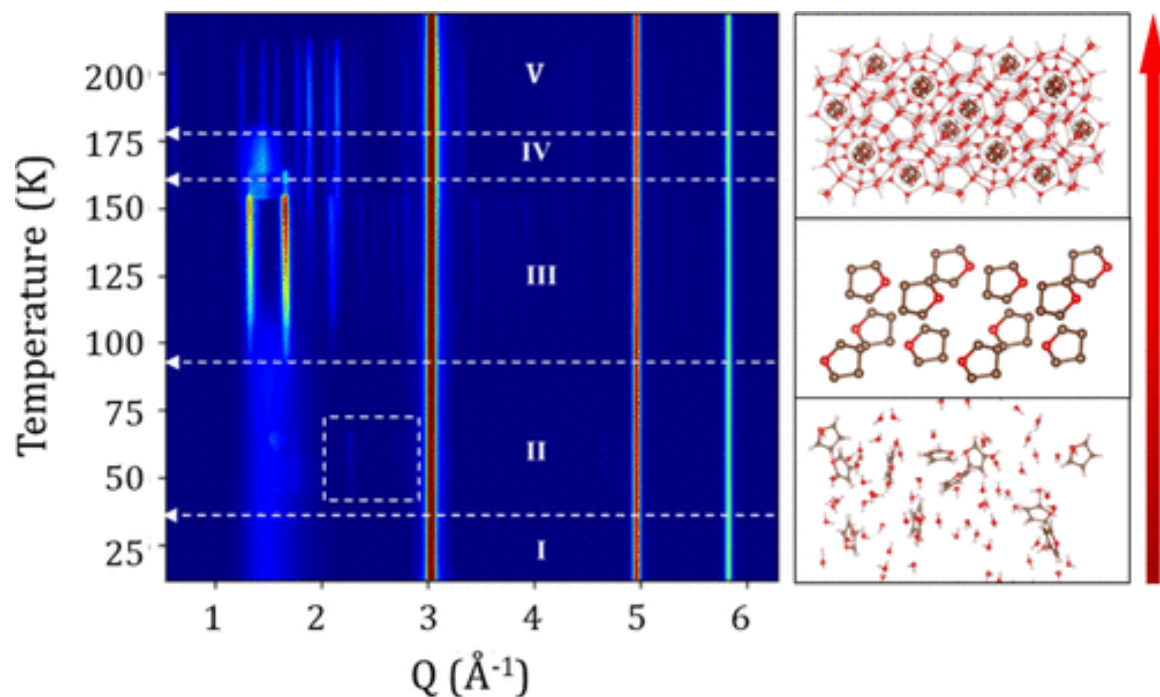


# X-ray scattering and diffraction in CLS



<https://brockhouse.lightsource.ca/about/low-energy-wiggler-beamline/>

Tse *et al.* reported the Clathrate hydrate behaviour



*J. Phys. Chem. Lett.* 2025, 16, 20, 5160-5167



# Small-angle X-ray scattering – Anton Paar SAXSPoint 2.0



The typical setup can resolve particle size range  
from 1 to 300 nm  
q-resolution as low as  $0.02 \text{ nm}^{-1}$

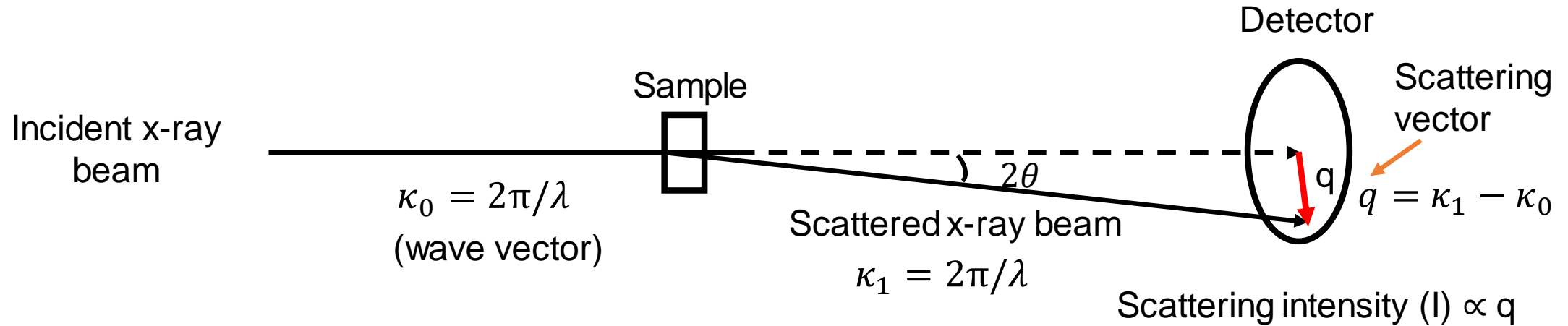
## Applications

- Protein analysis
- Polymers
- Fibers
- Structural biology
- Colloidal dispersions
- Emulsions
- Pharmaceuticals
- Nanoparticles
- Liquid crystals
- Porous materials
- Thin films

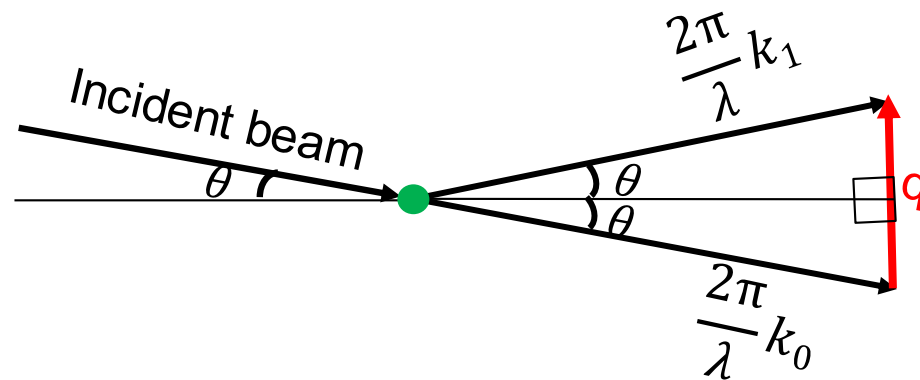
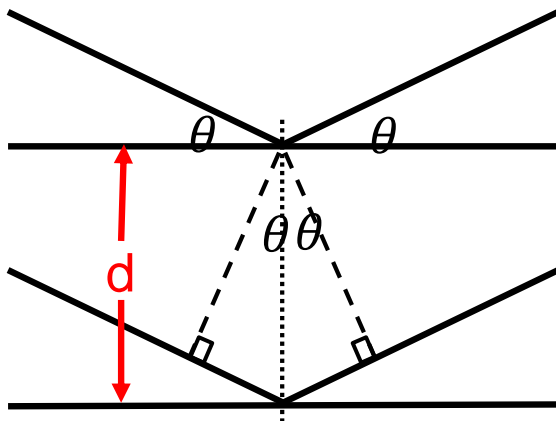
## Results

- Size distribution
- Shape
- Surface
- Internal structure
- Interaction
- Porosity
- Crystallinity
- Orientation
- Molecular weight

# X-ray scattering: basics



$\lambda$  of incident x-ray



**Bragg's law**

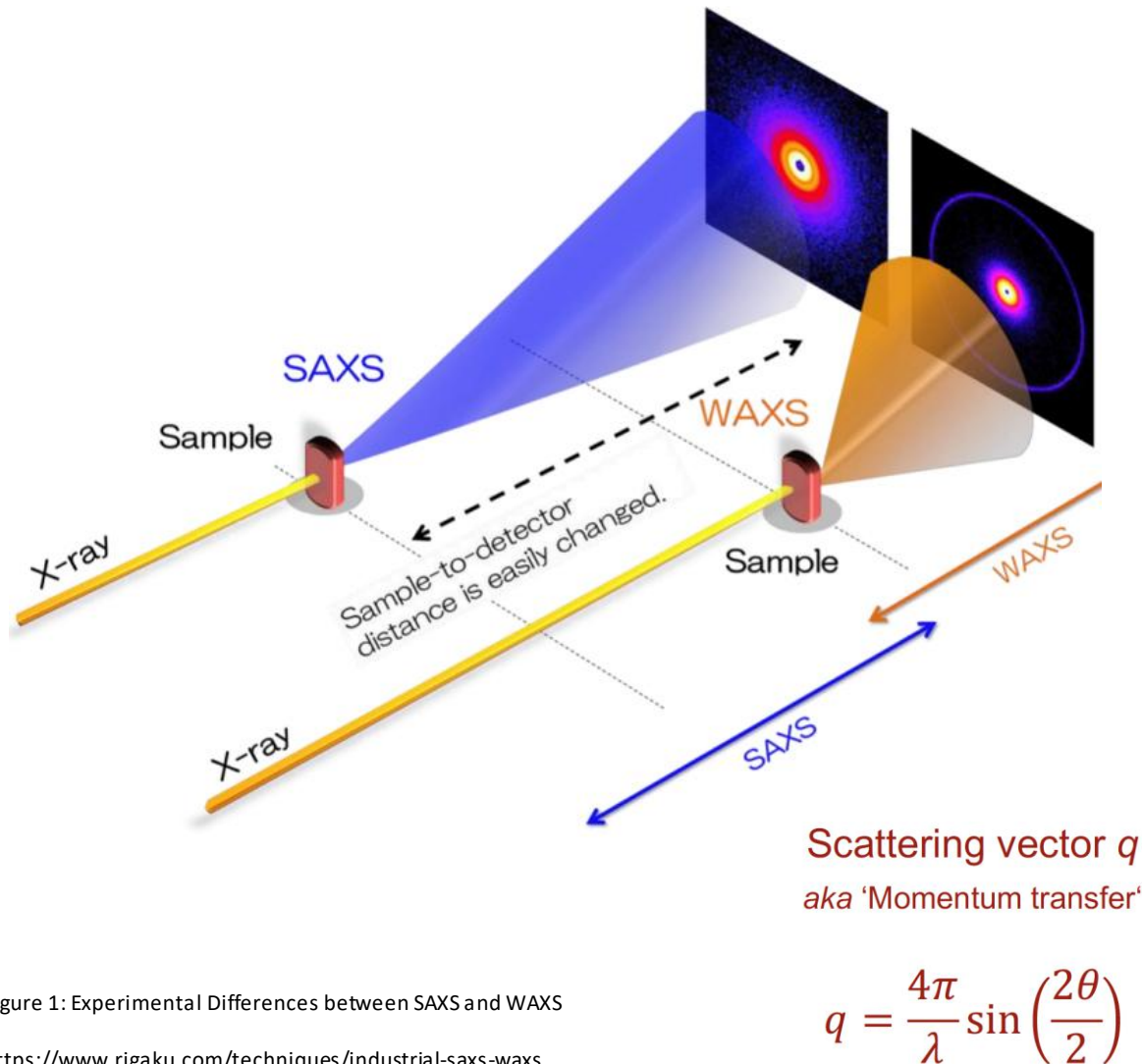
$$\lambda = 2d \sin(\theta)$$

$$d = \frac{2\pi}{q}$$

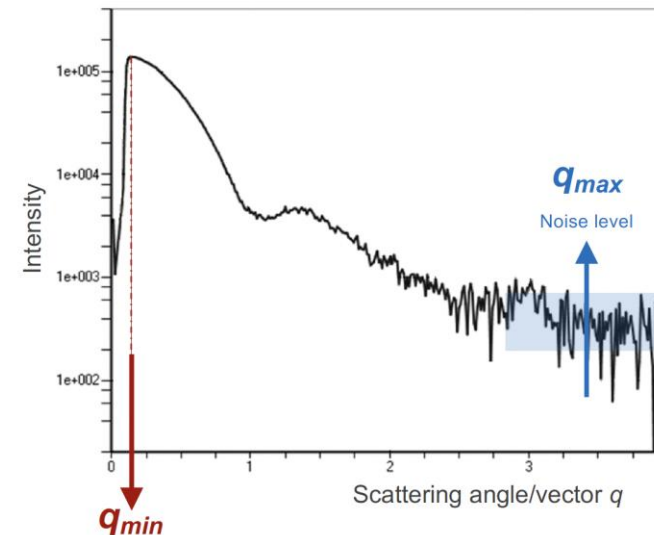
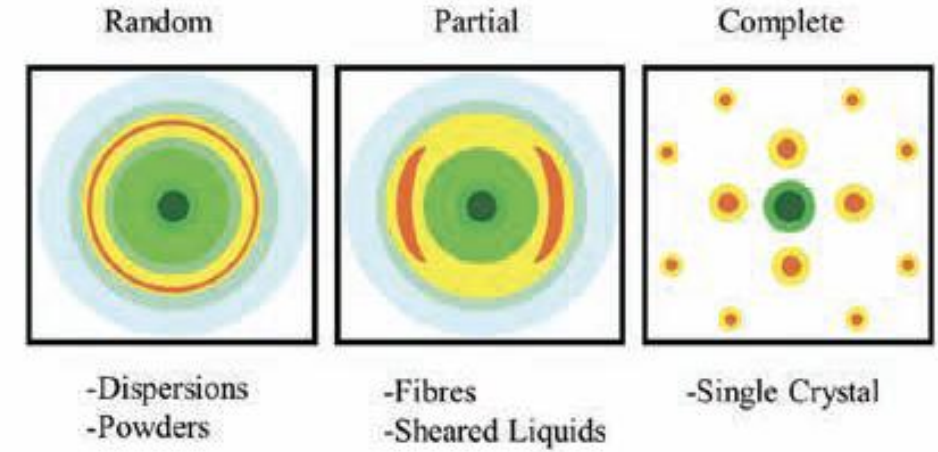


$$q = \frac{4\pi}{d} \sin(\theta)$$

# X-ray scattering: basics



Sample Orientation:



Largest resolvable  
dimension (particle)

$$D_{max} = \frac{\pi}{q_{min}}$$

Smallest resolvable  
dimension (particle)

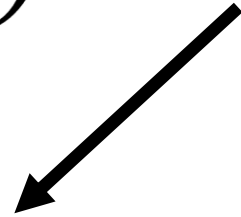
$$D_{min} = \frac{\pi}{q_{max}}$$

Figure 1: Experimental Differences between SAXS and WAXS

<https://www.rigaku.com/techniques/industrial-saxs-waxs>

# Scattering from particles

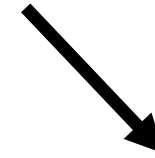
$$I(q) = n |F(q)|^2 S(q)$$



Number density  
of particles



Form factor  
(particle shape)

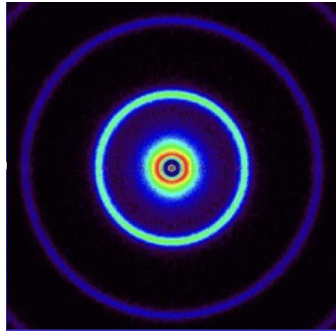


Structure factor  
(particle interactions)

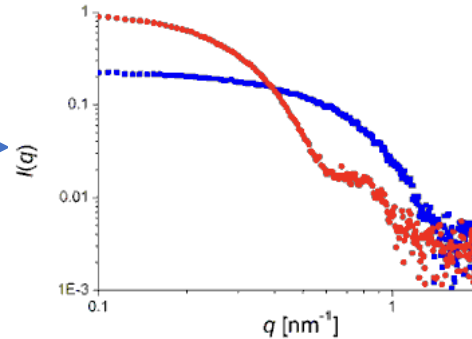
Discrete particles are often the easiest to understand and analyze



# General flow chart for data analysis

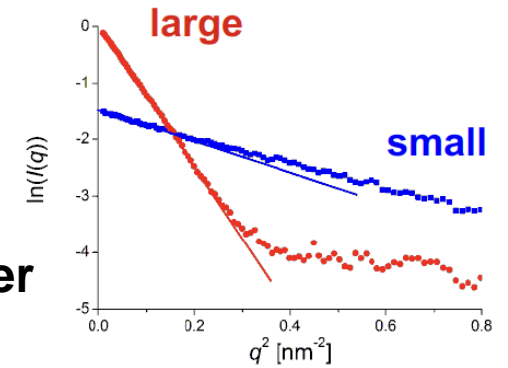


Reduction  
2D to 1D



Basic calculations

Guinier



PDDF

Peak analysis

Slope analysis

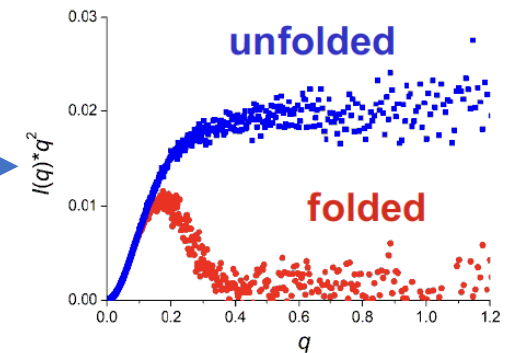
Orientation/crystallinity

Porod  
 $I(q) \sim q^{-n}$

Molecular  
weight

Model fitting

And more



Kratky plot

SASBDB

Small Angle Scattering Biological Data Bank

sign in | Register

Advanced search  Search

E.g. P00921, BSA, Nucleic Acids Res

Home Browse Submit data About SASBDB Help

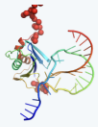
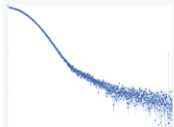
Curated repository for small angle scattering data and models

Small angle scattering (SAS) of X-ray and neutrons provides structural information on biological macromolecules in solution at a resolution of 1-2 nm. SASBDB is a fully searchable curated repository of freely accessible and downloadable experimental data, which are deposited together with the relevant experimental conditions, sample details, derived models and their fits to the data.

SASBDB currently contains:  
4809 experimental data sets  
5988 models  
570 experimental data sets on hold  
794 models on hold

Recent depositions:

SASDWS5 – 2A protein from Theiler's murine encephalomyelitis virus (TMEV) bound to stimulatory element RNA

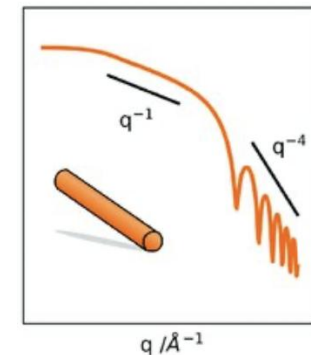
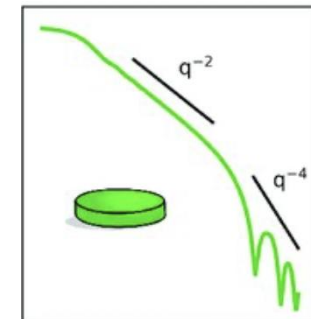
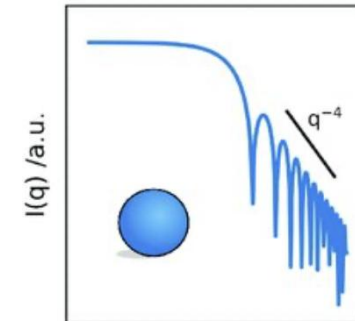
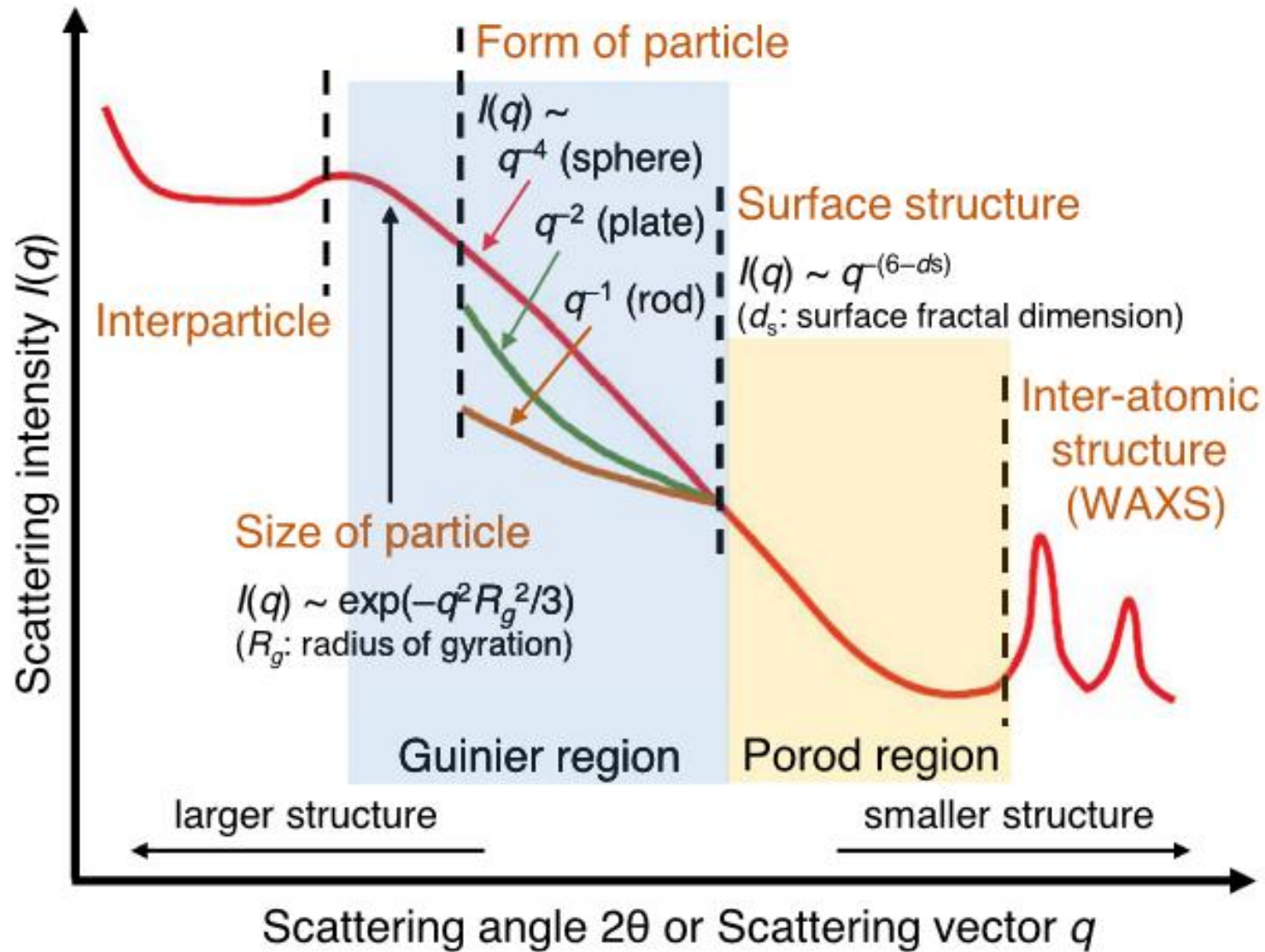


Sample: Genome polyprotein monomer, 17 kDa *Theiler's murine encephalomyelitis* ... protein  
Stimulatory element RNA monomer, 12 kDa *synthetic RNA* RNA  
Buffer: 20 mM HEPES pH 7.4, 400 mM KCl 2% v/v glycerol, 1 mM DTT, pH:  
Experiment: SAXS data collected at EMBL P12, PETRA III on 2021 Apr 26

$R_g^{Guinier}$  2.2 nm  
 $D_{max}$  7.5 nm  
 $Volume^{Porod}$  46 nm<sup>3</sup>

A new protein-dependent riboswitch activates ribosomal frameshifting (2025)  
Betts J, Jeffries C, Passchier T, Kung H, Graham S, Abdelhamid M, Howard J, Craggs T, Graham S, Brierley I, Leake M, Quinn S, Hill C

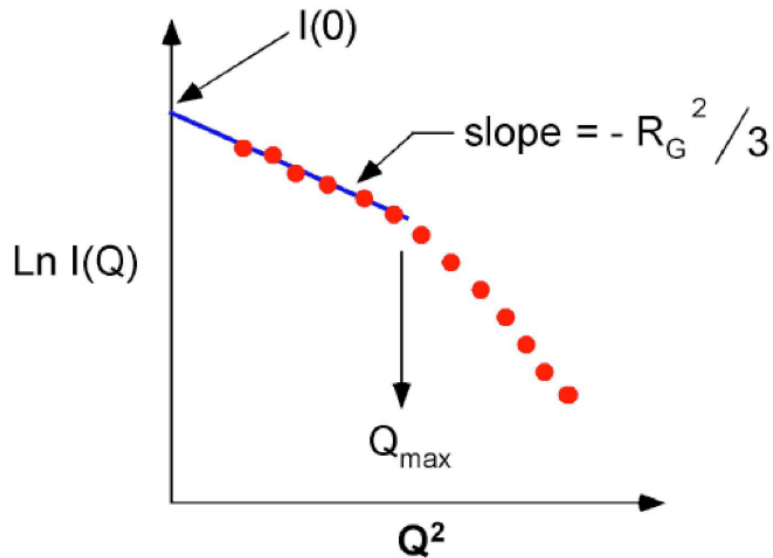
# q-plot (or s) and what type of information can we obtain?



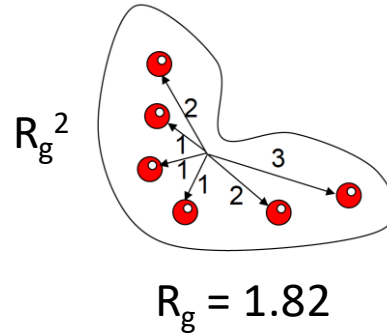
# Guinier region: defines the radius of gyration $R_g$ , the overall dimension of the particle

## Guinier Plot:

$$\ln[I(Q)] = \ln[I(0)] - Q^2 R_G^2 / 3$$

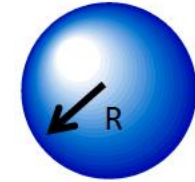


Quarterly Reviews of Biophysics 2007, 40, 191–285.



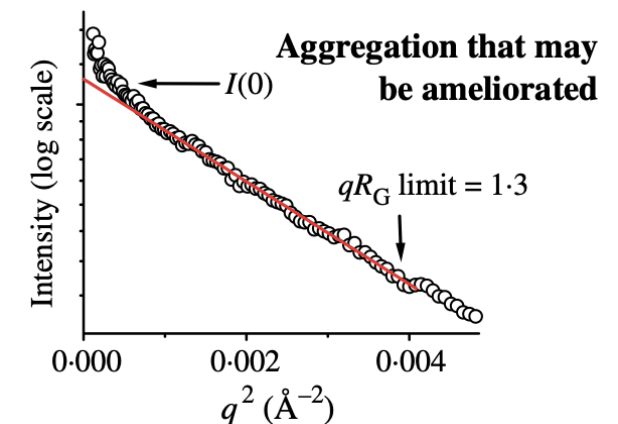
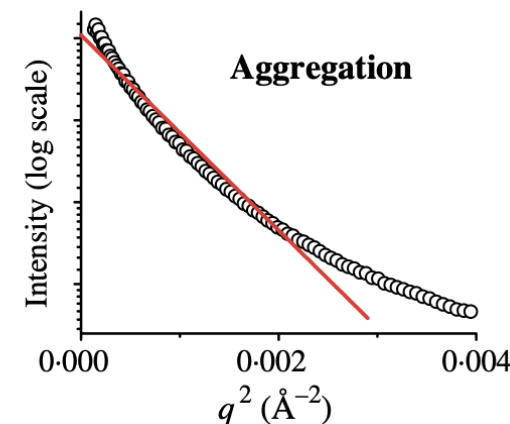
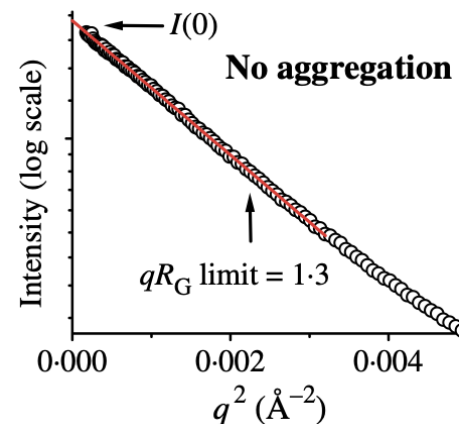
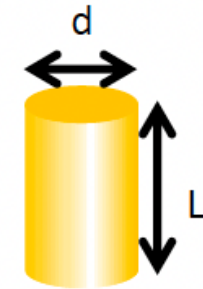
Sphere:

$$R_g^2 = \frac{3}{5} R^2$$



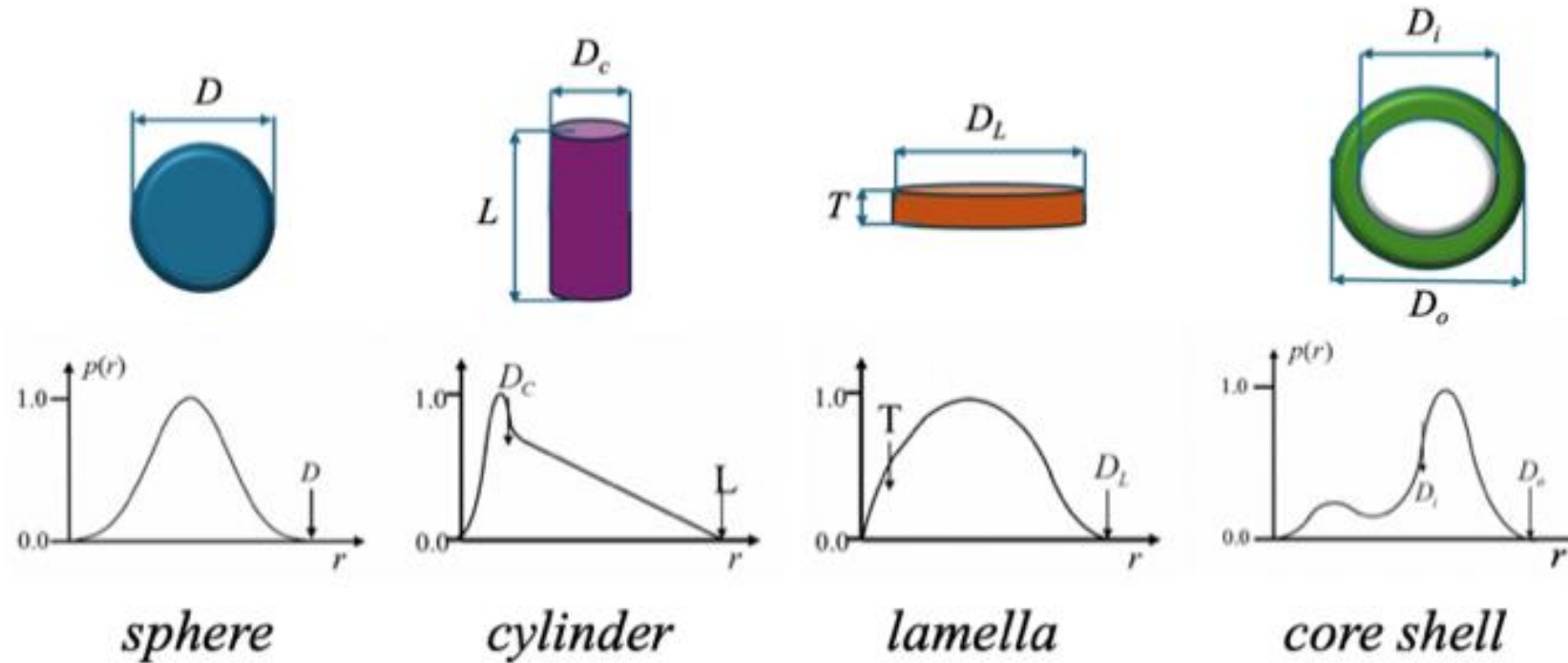
Cylinder:

$$R_g^2 = \frac{L^2}{12} + \frac{d^2}{8}$$



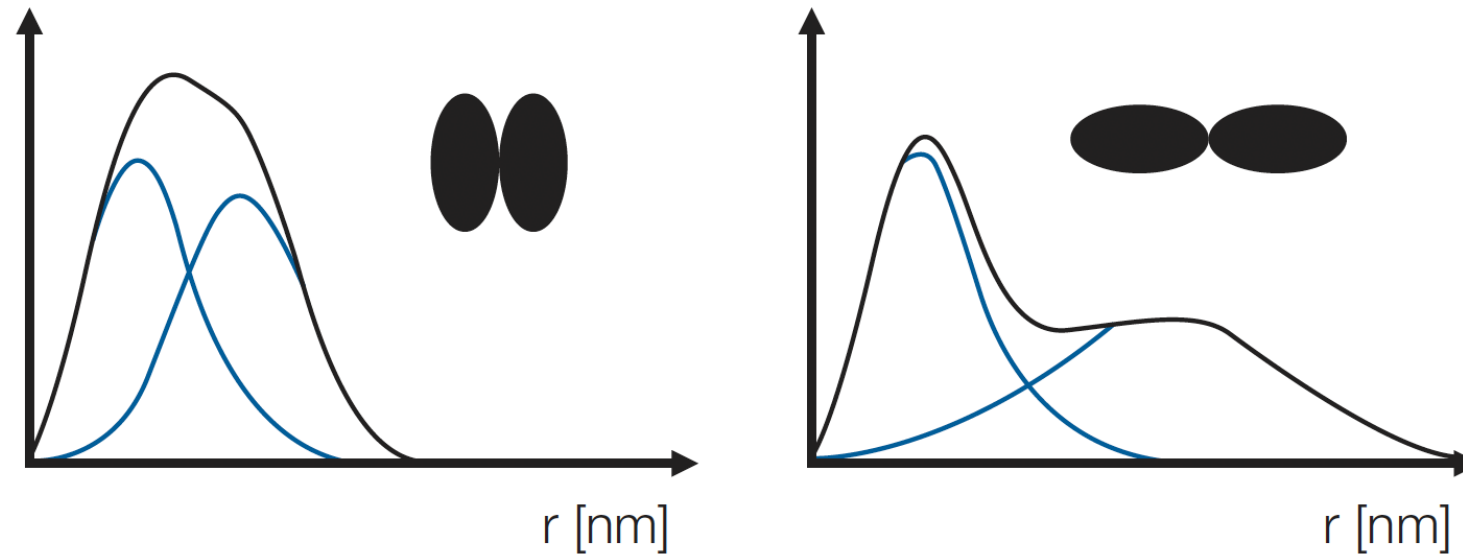
The radius of gyration is a distribution of components of an object around an axis. Note that  $R_g$  does not contain information about particle shape

# Pair distance distribution function $P(r)$ ; PDDF



- Calculated from form factor
- Appearance of the resulting plot enables prediction of molecular size and shape
- $P(r)$  is the inverse Fourier Transform of  $I(q)$  and represents the histogram of electron-pair distances in compounds.

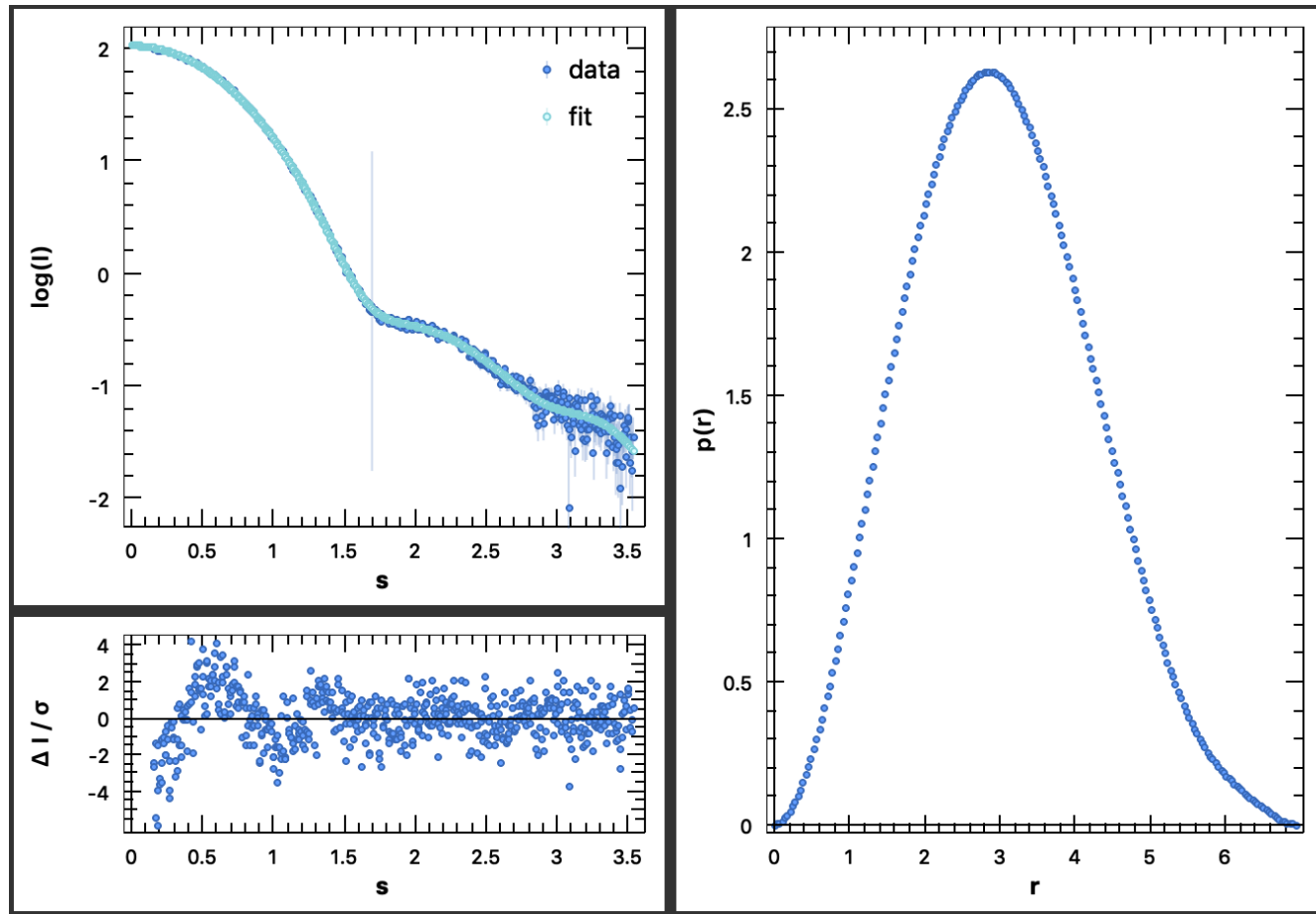
# Aggregates and PDDF



*The aggregate of two subunits makes a PDDF, which a second peak can recognize.*



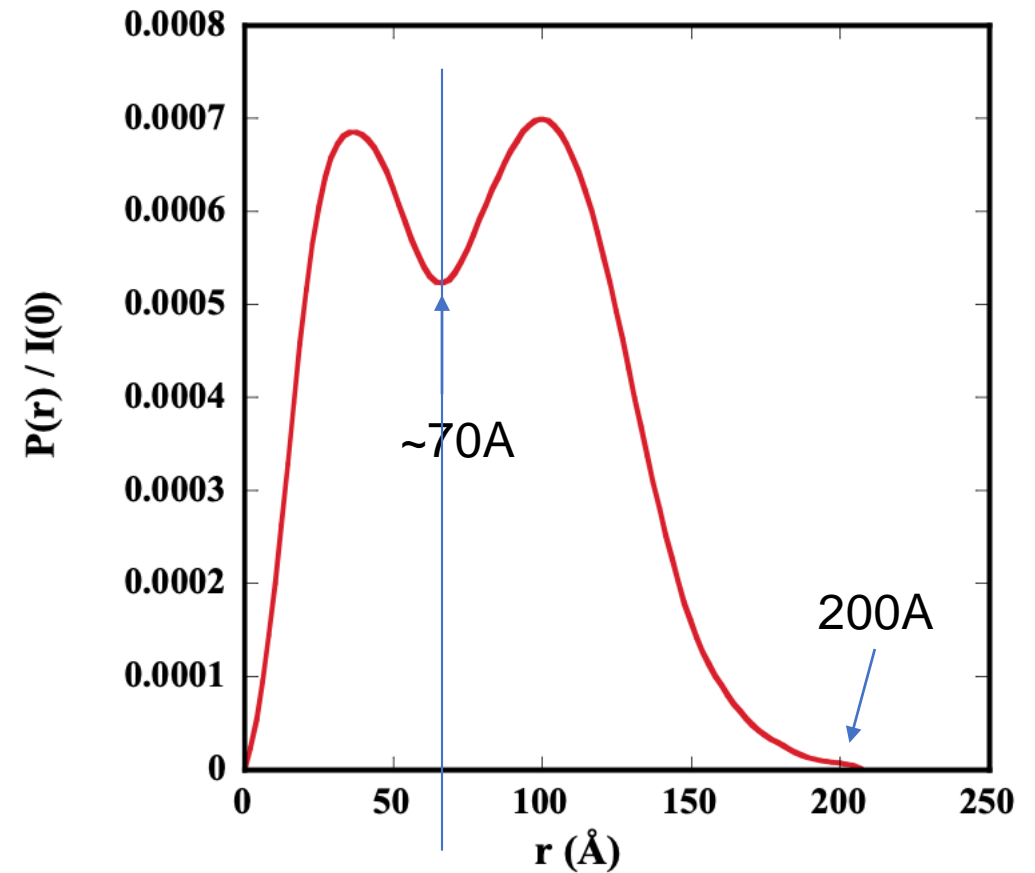
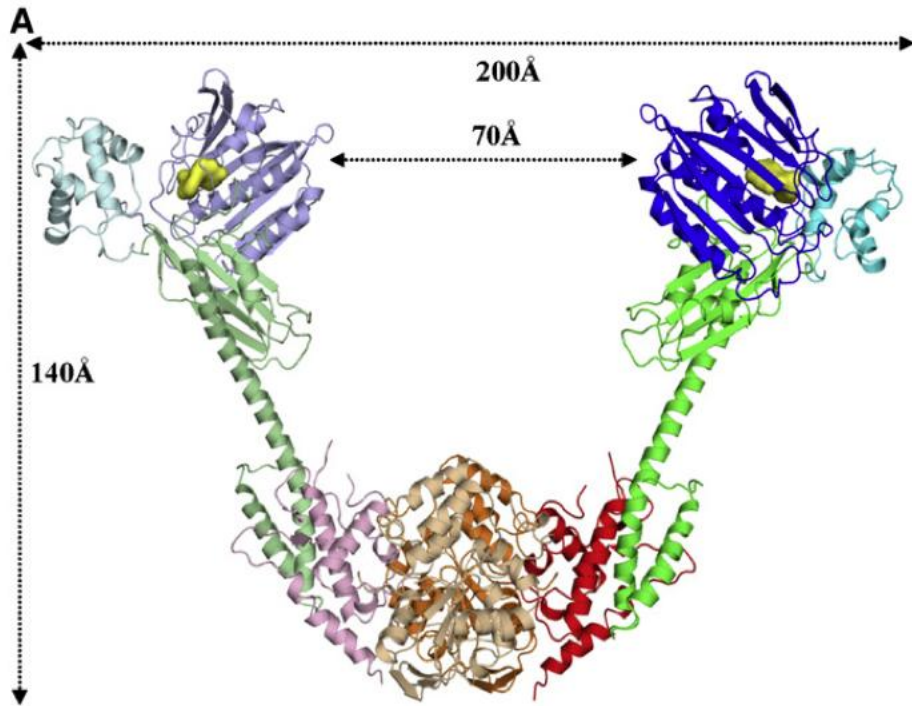
# The $R_g$ and $P(r)$ of quantum dots



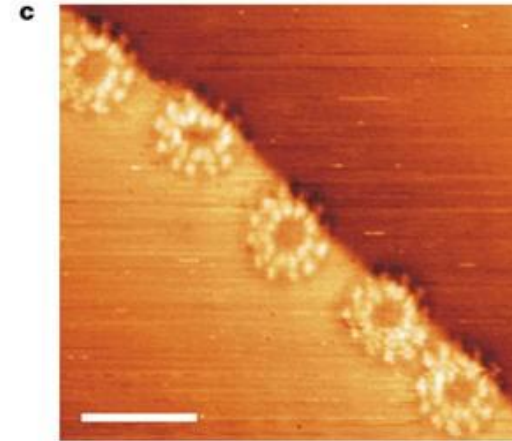
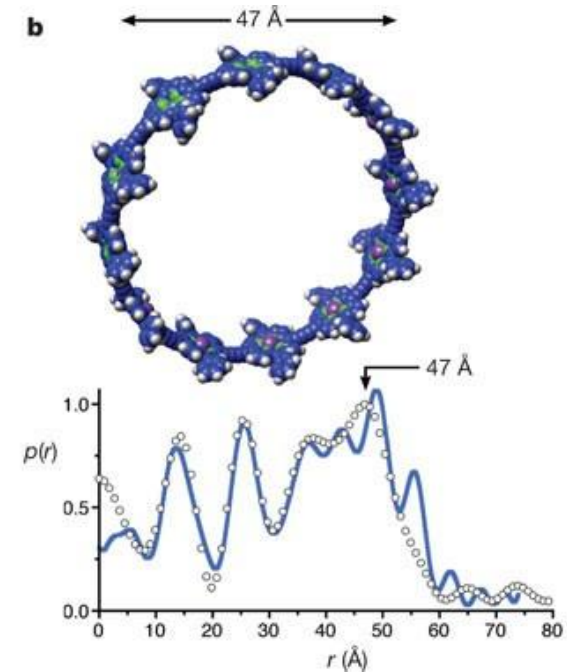
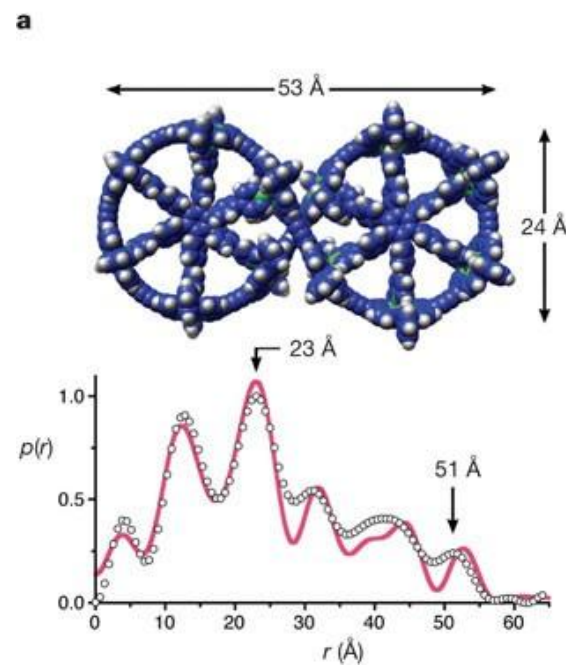
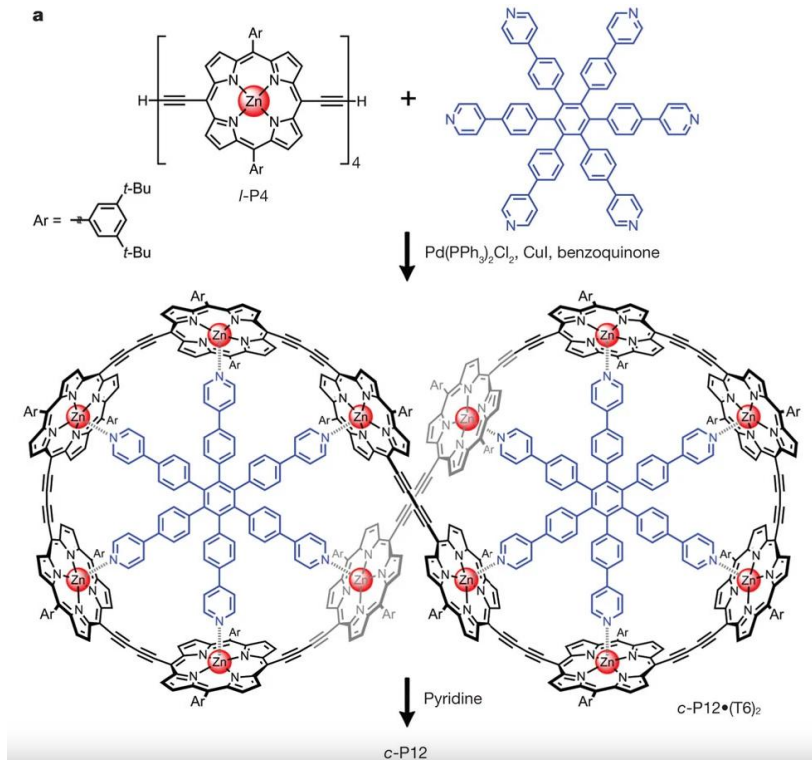
Monodisperse spheres of  
CdS quantum dots  
 $R_g = 2.26$  nm  
(solved using ATSAS).

The results are in good  
agreement with the TEM

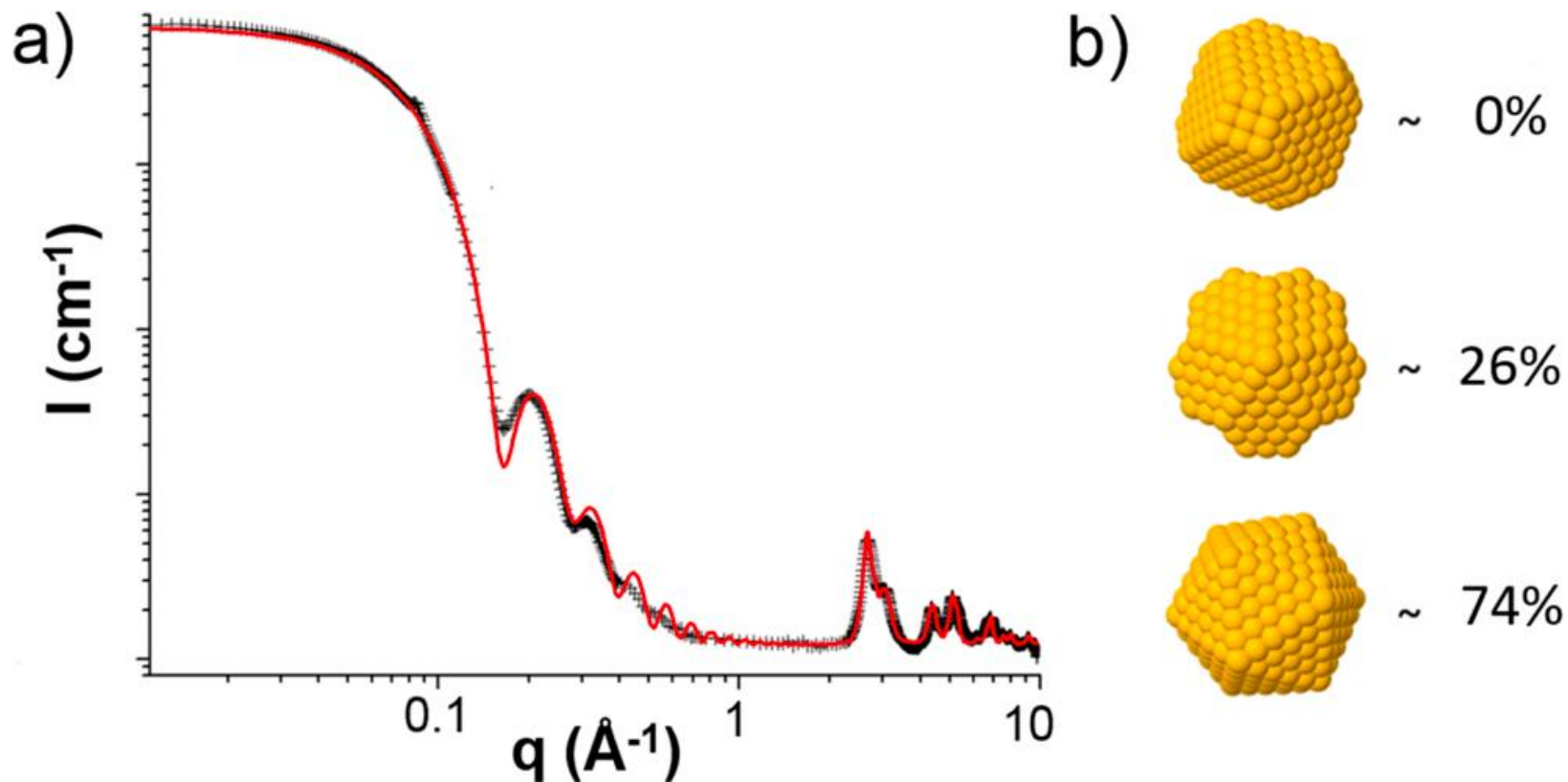
# PDDF example of biomolecular structure



# Pair distance distribution function $P(r)$ : examples




# Modeling different external shapes using SAXS and WAXS




*The best fit is obtained from a linear combination of the calculated scattering intensities of three different external shapes, including icosahedra, decahedra, and truncated octahedral structures*

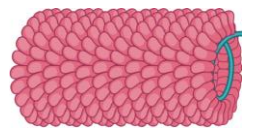
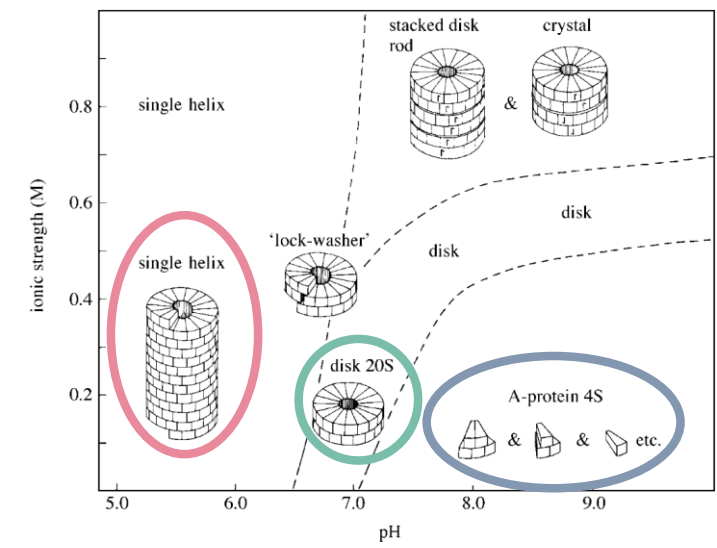


# SAXS – Lab X-ray source vs Synchrotron

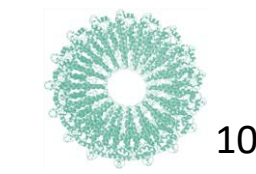


Guido Merino ([guido.merino@mail.mcgill.ca](mailto:guido.merino@mail.mcgill.ca))  
 Dr. Harrington Lab, McGill University.  
[Matt.harrington@mcgill.ca](mailto:Matt.harrington@mcgill.ca)

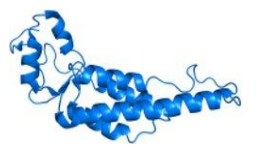




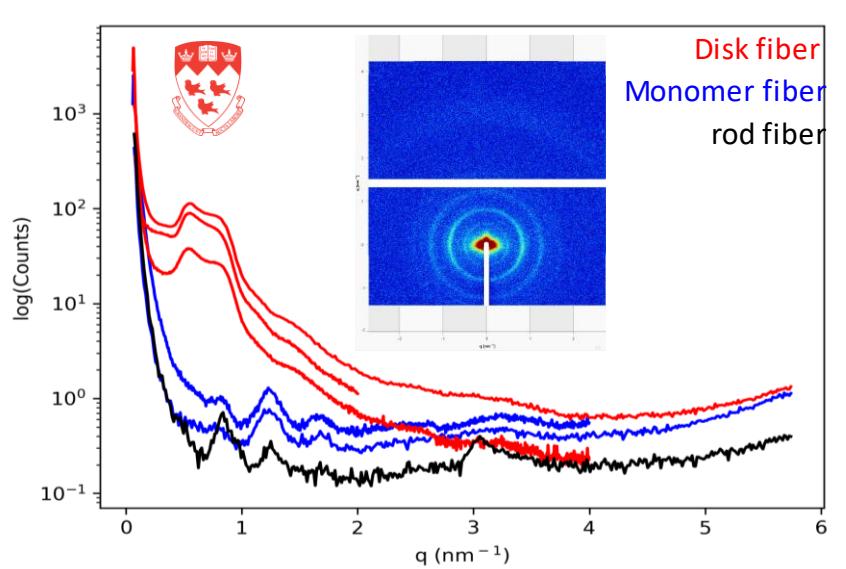
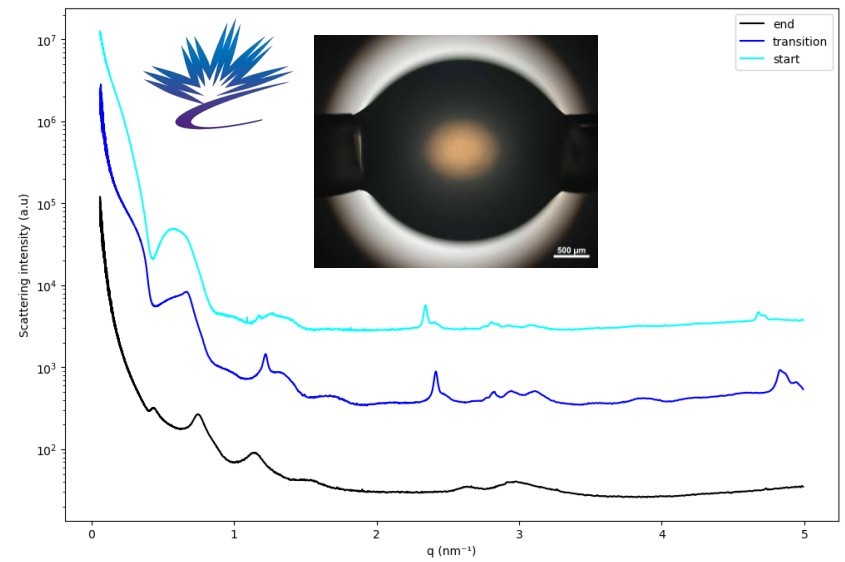
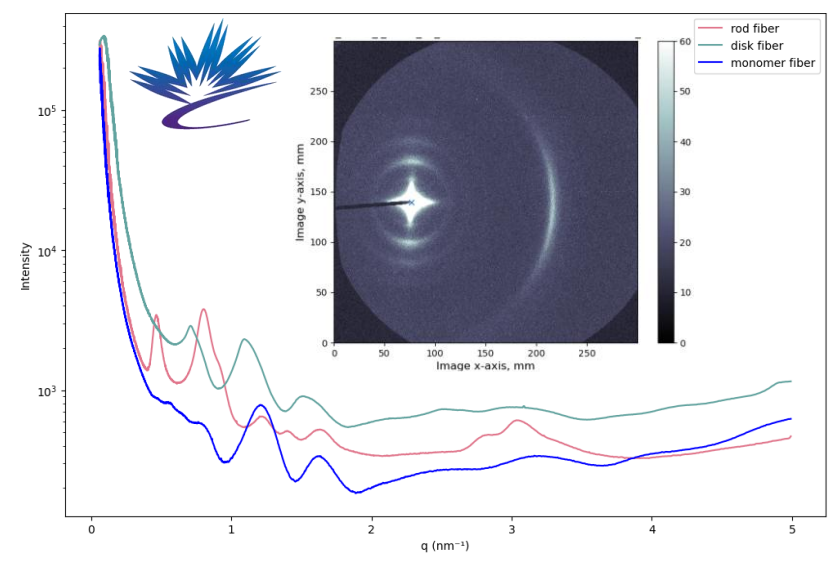
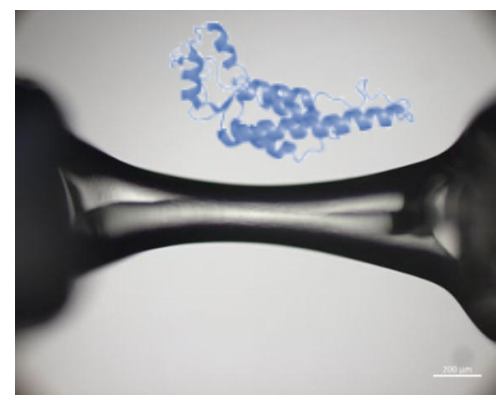
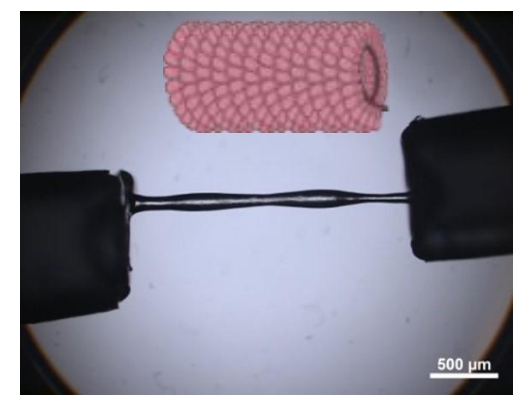
Rod, pH 5.0  
10mM Acetate



Disk, pH 6.8  
10mM Potassium Phosphate

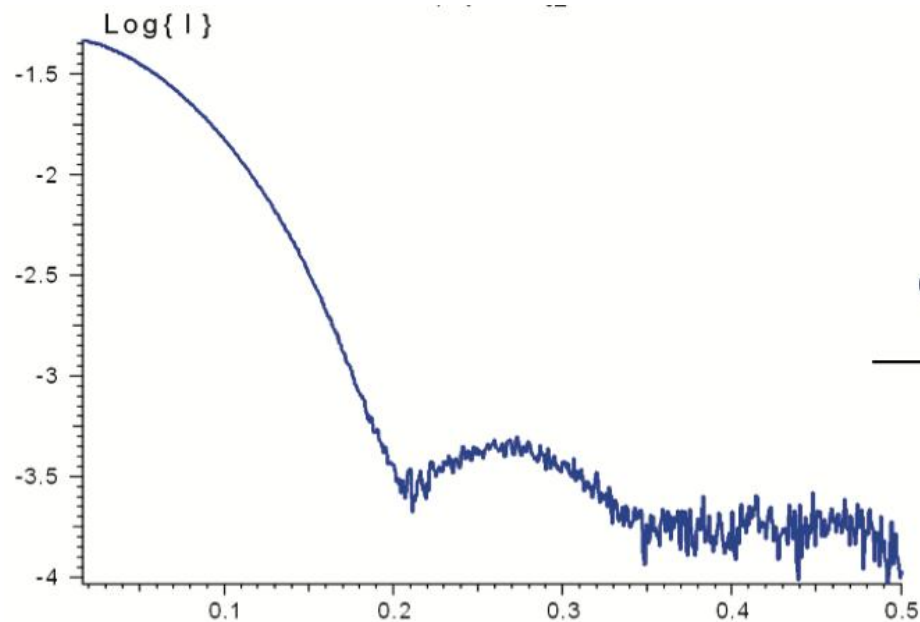


Monomer, pH 8.0  
10mM Triethanolamine



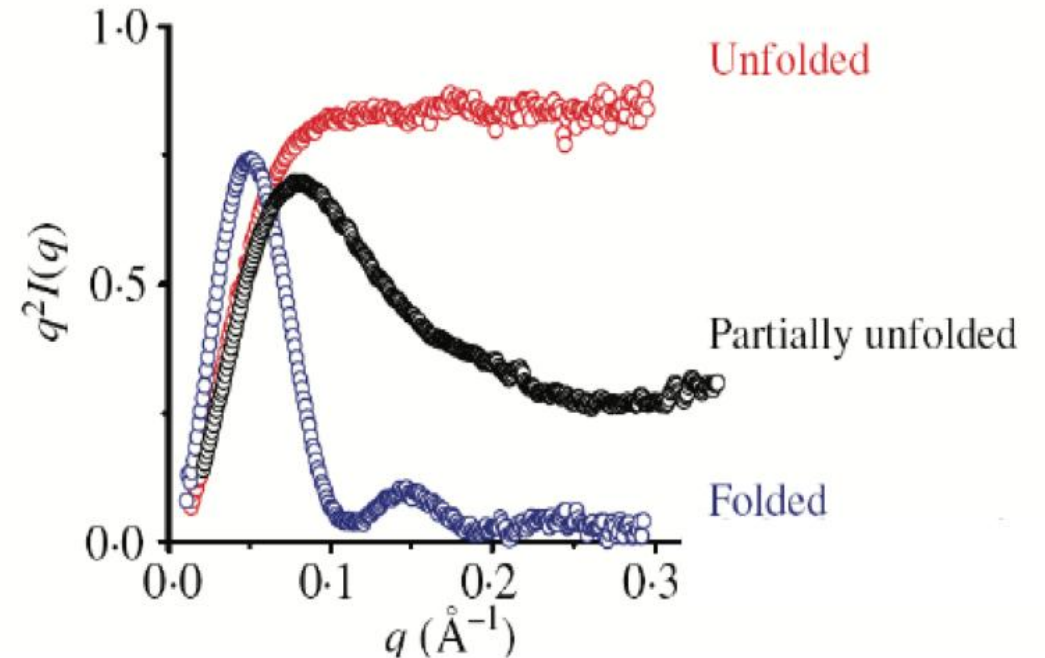


# Kratky plot: protein analysis



A protein sample

$q^2 I(q)$  versus  $q$



*Putnam, D., et al. (2007) Quart. Rev. Biophys. 40, 191-285.*

- Kratky plot:  $I \cdot q^2$  vs.  $q$
- Sensitive to morphology of the particle
- Sensitive to the compactness of a protein
- Unfolded and folded states of proteins are easy to distinguish

# Supramolecular structures and their corresponding peak ratios observed in SAXS

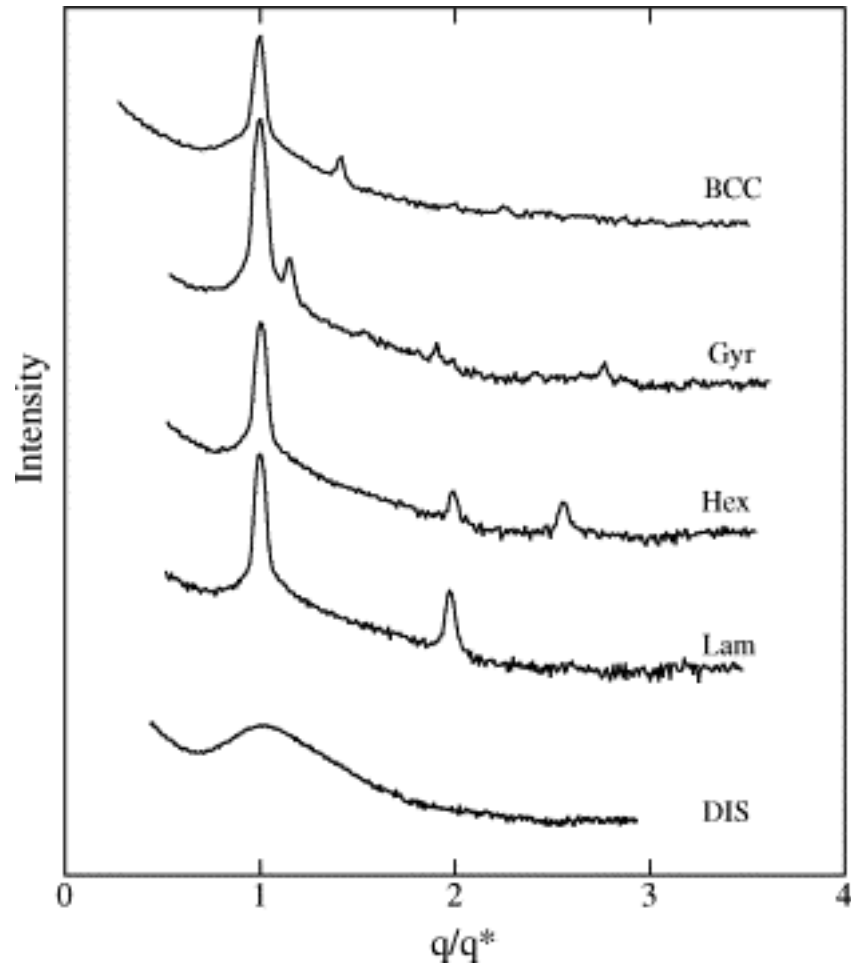
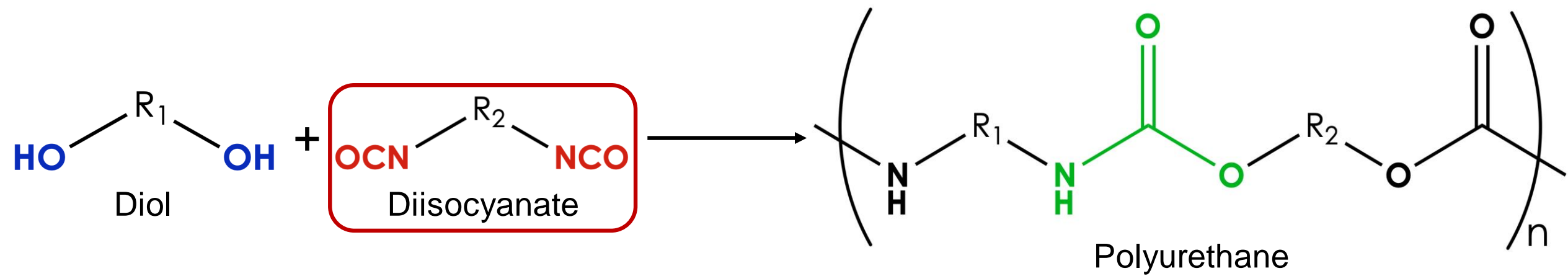


Table 1. Peak positions (expressed as  $q/q^*$ ) of Bragg reflections for various structures

Structure	Ratio $q/q^*$
Lam	1, 2, 3, 4, 5, 6,...
Hex ( $p6mm$ )	1, $\sqrt{3}$ , $\sqrt{4}$ , $\sqrt{7}$ , $\sqrt{9}$ , $\sqrt{12}$ ,...
BCC ( $Im\bar{3}m$ )	1, $\sqrt{2}$ , $\sqrt{3}$ , $\sqrt{4}$ , $\sqrt{5}$ , $\sqrt{6}$ ,...
FCC ( $Fm\bar{3}m$ )	1, $\sqrt{4/3}$ , $\sqrt{8/3}$ , $\sqrt{11/3}$ , $\sqrt{12/3}$ , $\sqrt{16/3}$ ,...
Gyr ( $Ia\bar{3}d$ )	1, $\sqrt{4/3}$ , $\sqrt{7/3}$ , $\sqrt{8/3}$ , $\sqrt{10/3}$ , $\sqrt{11/3}$ ,...

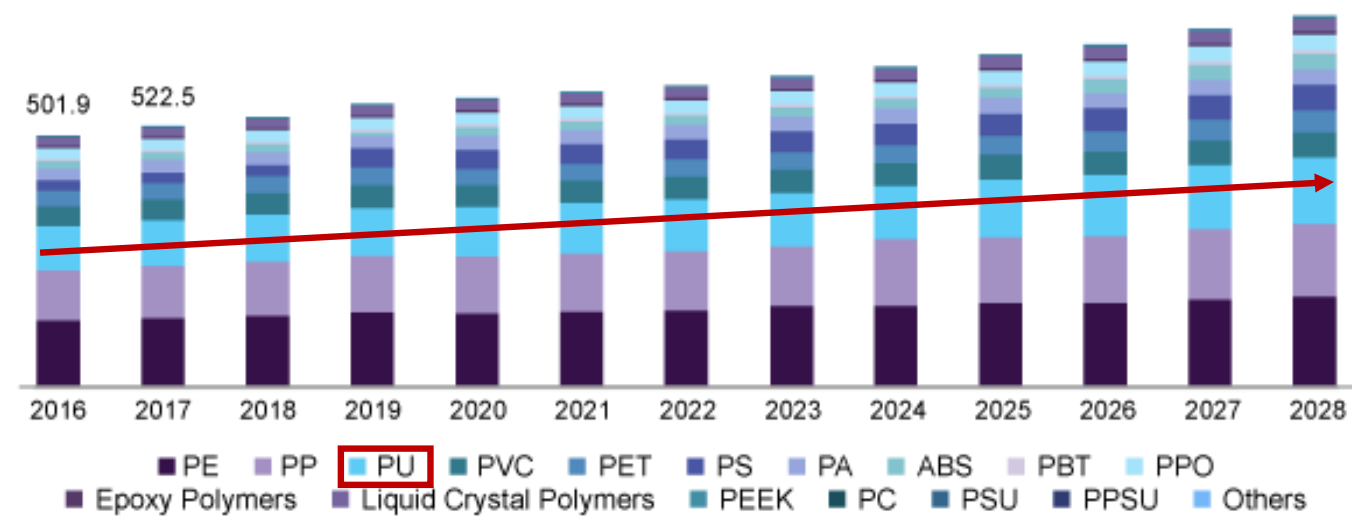
# Conventional polyurethanes (PUs)



Formulations of polyurethanes without isocyanates:

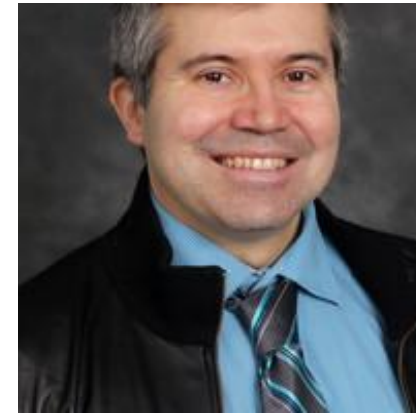
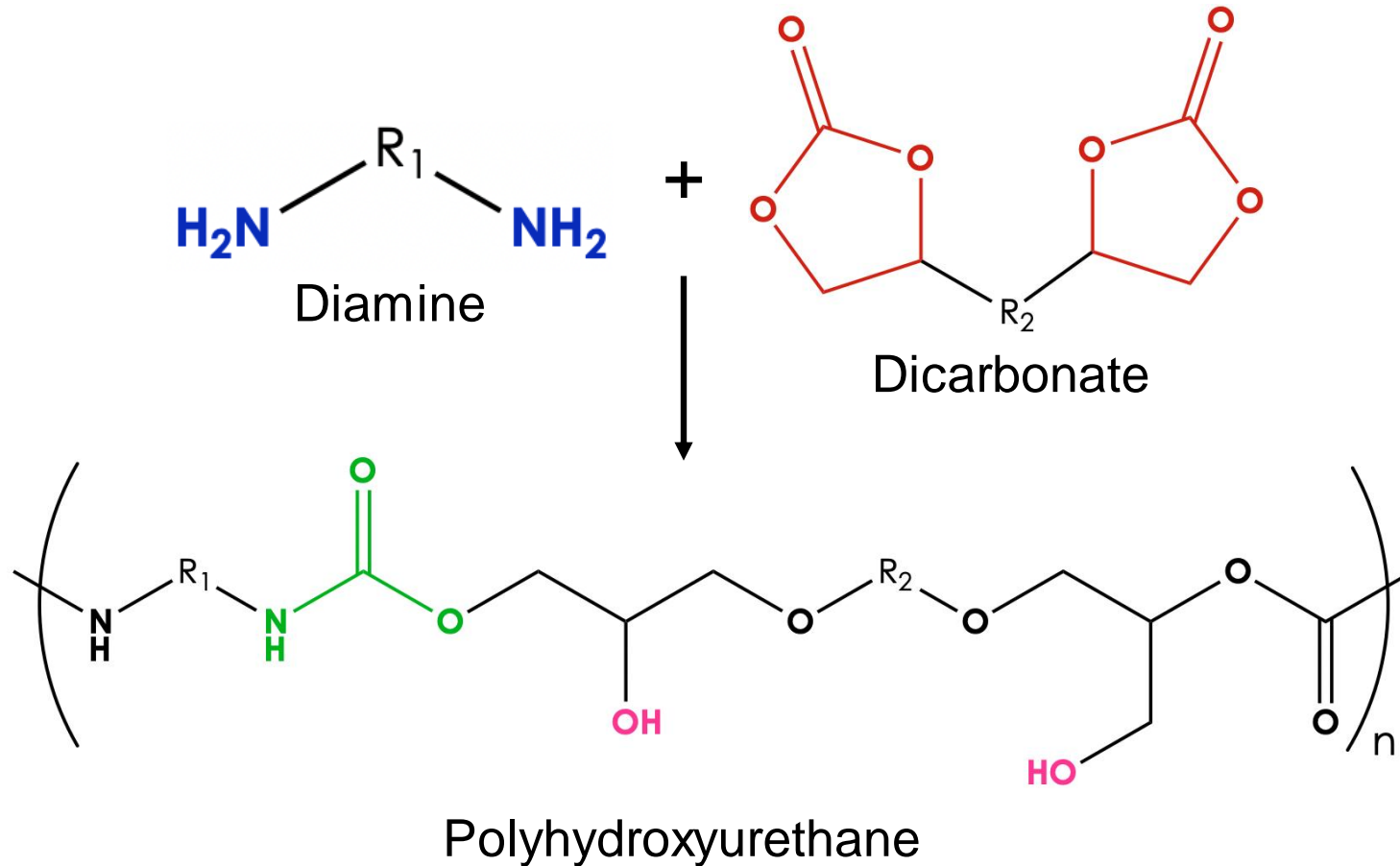
**Non-Isocyanate Polyurethanes (NIPUs)**

Global plastic market size, by product, 2016 - 2028 (USD Billion)



<https://www.grandviewresearch.com/industry-analysis/global-plastics-market>

# Non-isocyanate polyurethanes (NIPUs): Green chemistry



Prof. Milan Marić



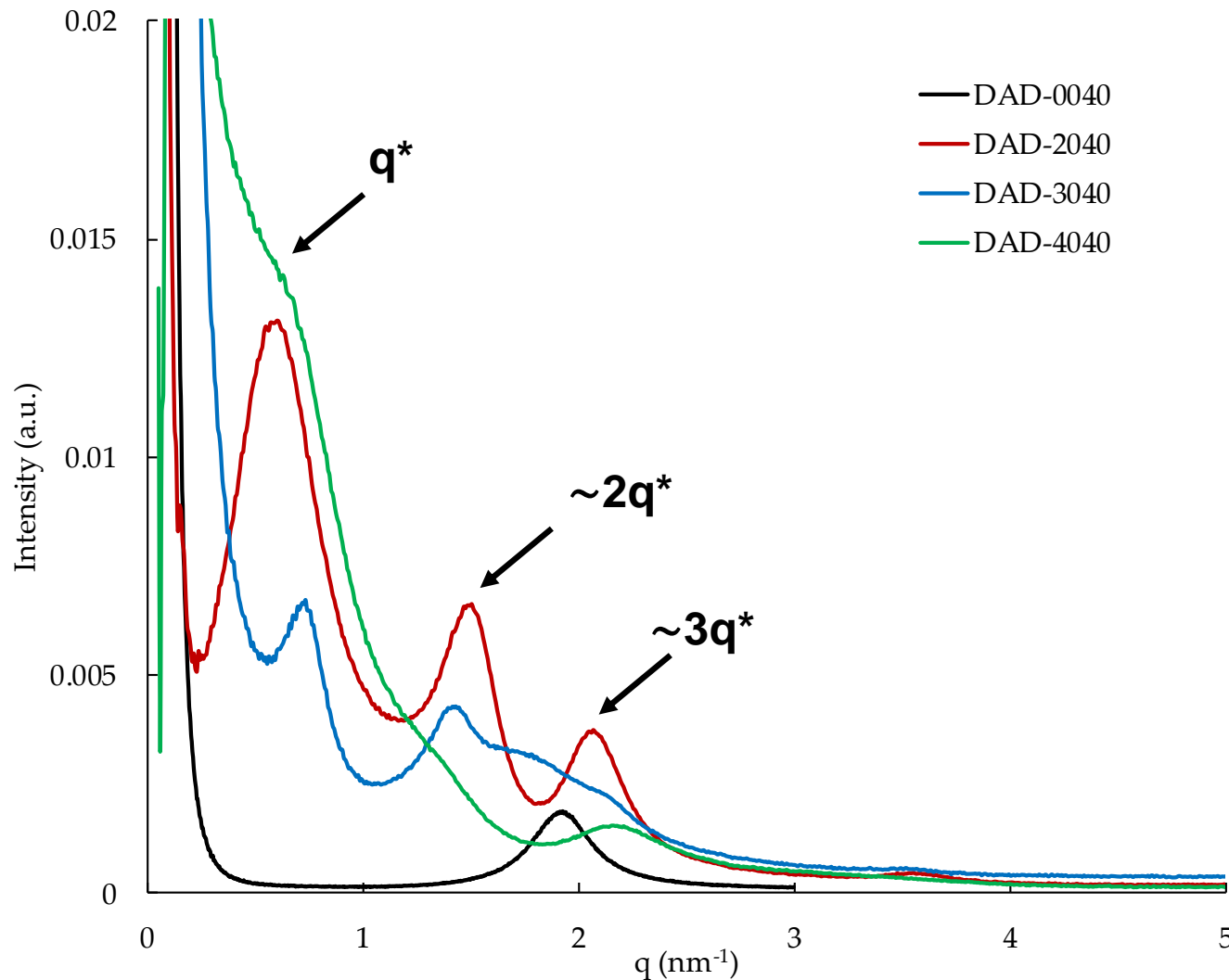
Dr. Georges Younes

McGill University  
Department of Chemical Engineering

Safe and green reagents, bulk polymerization, moderate conditions

Hydroxyl groups: hydrogen bonding and post-functionalization

# Thermoplastic PHUs microstructure: SAXS

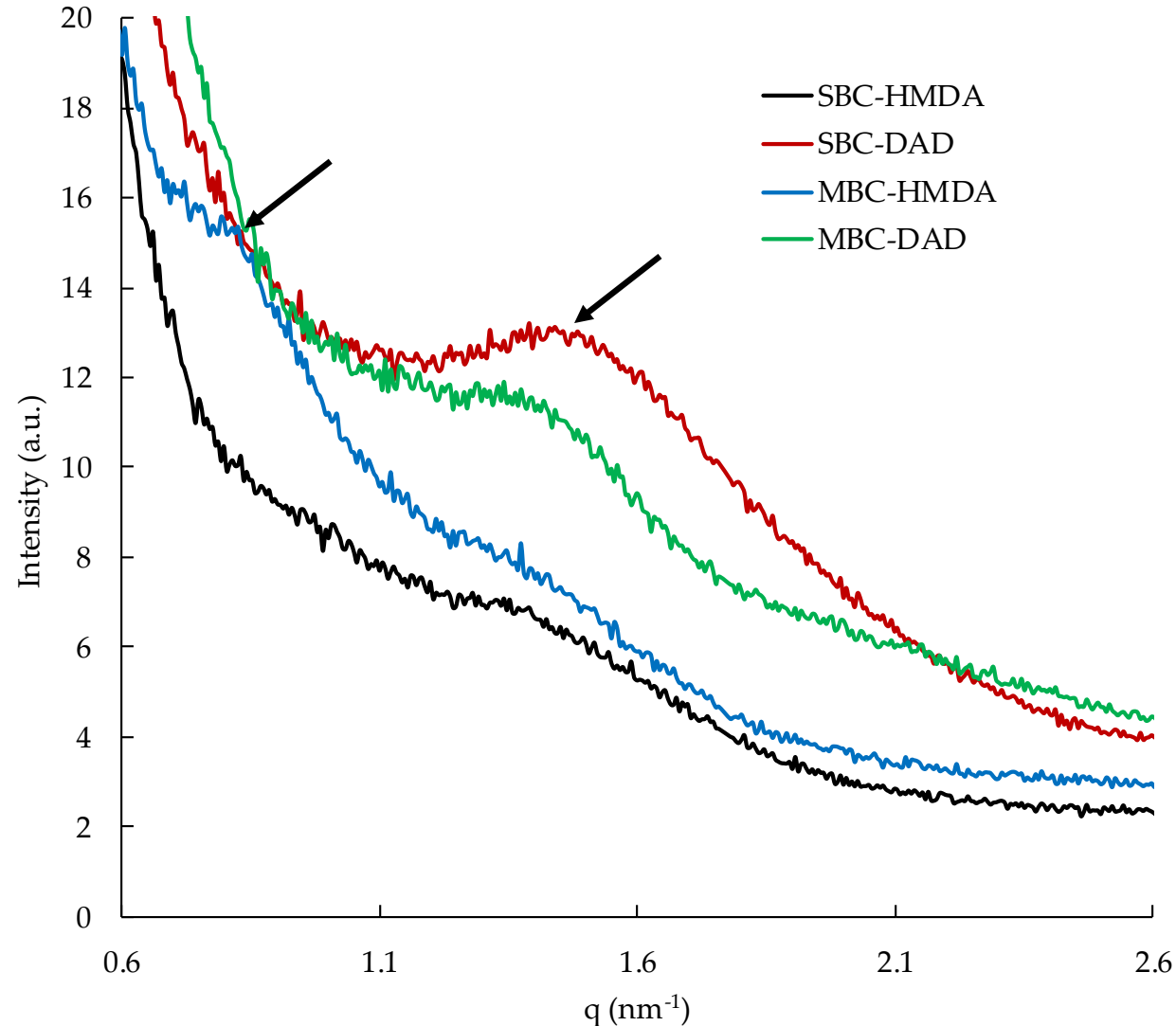


TPHUs from DGC and Priamine 1074 show **disordered nanophase separation** between hard and soft segments

Addition of DAD introduced **crystallinity** into the microstructure of TPHU - **ordered segmented lamellae** pattern (polymer chains are parallel to one another at room temperature)



# SBC/MBC-based TPHUs microstructure: SAXS



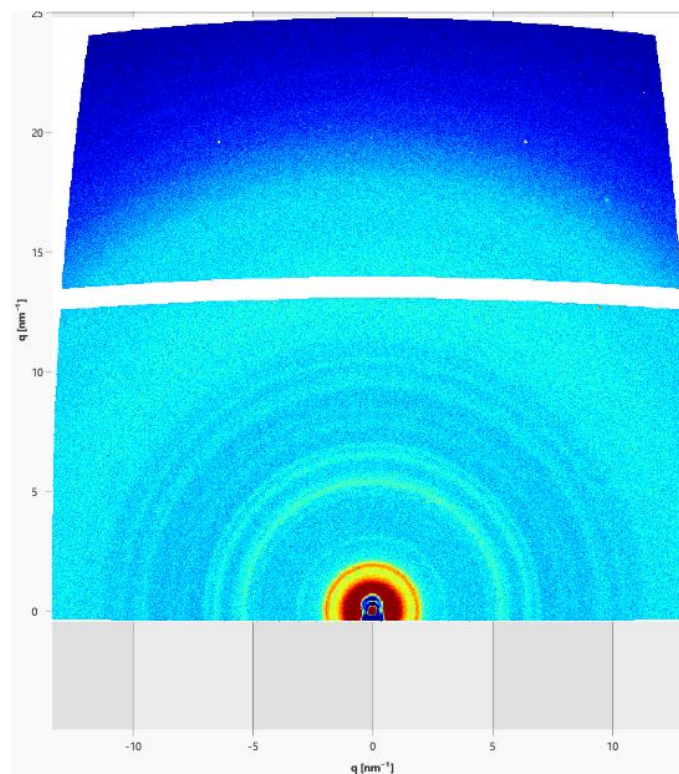
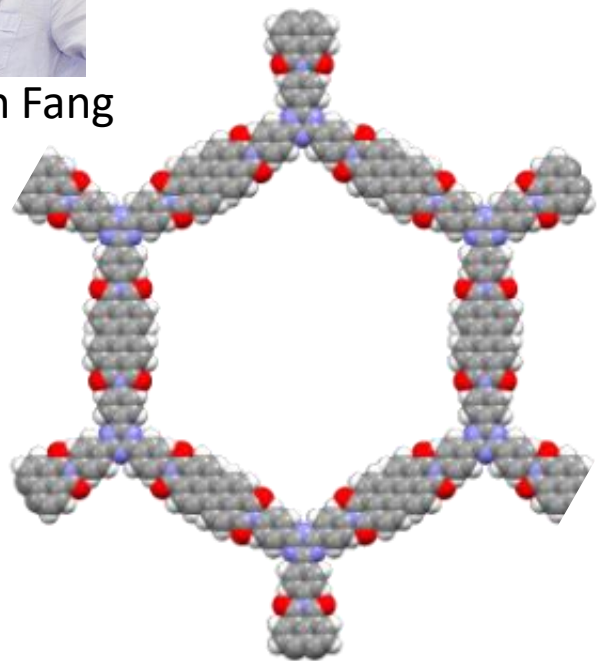
PHU	$q_{\max}$ (nm <sup>-1</sup> )	$d$ (nm)
SBC-HMDA	1.40	4.49
SBC-DAD	1.48	4.27
MBC-HMDA	0.83/1.42	7.57/4.42
MBC-DAD	1.42	4.42

- **Nanophase separation** exists in PHUs (hard segments versus soft segments)
- **Random ordering** of the PHU chains except for MBC-HMDA chains (ordered in a hexagonal  $P6mm$  ( $1, \sqrt{3}...$ ) spacial geometry)

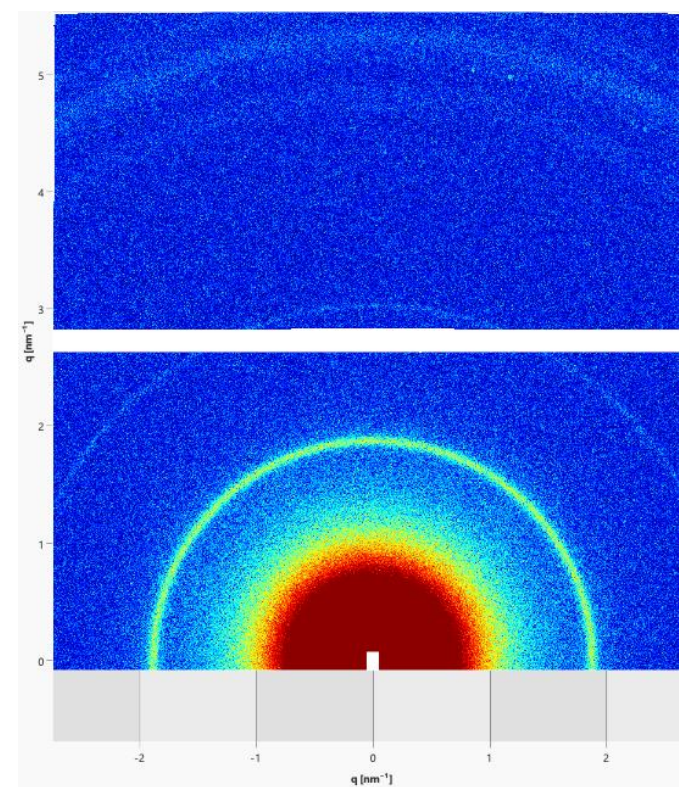


Prof. Yuan Fang

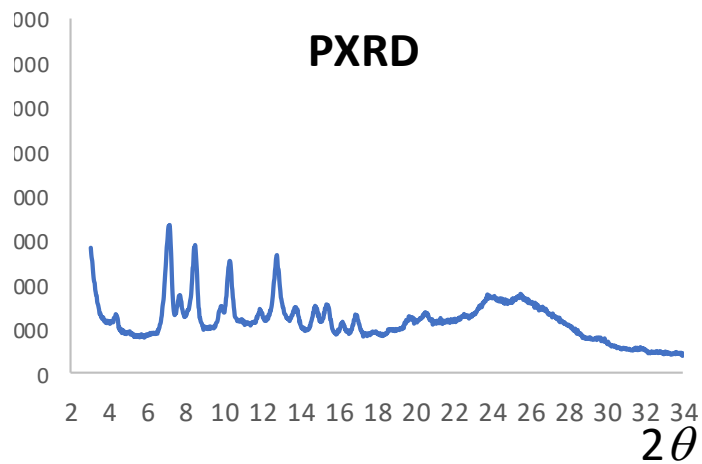
# Covalent Organic Framework



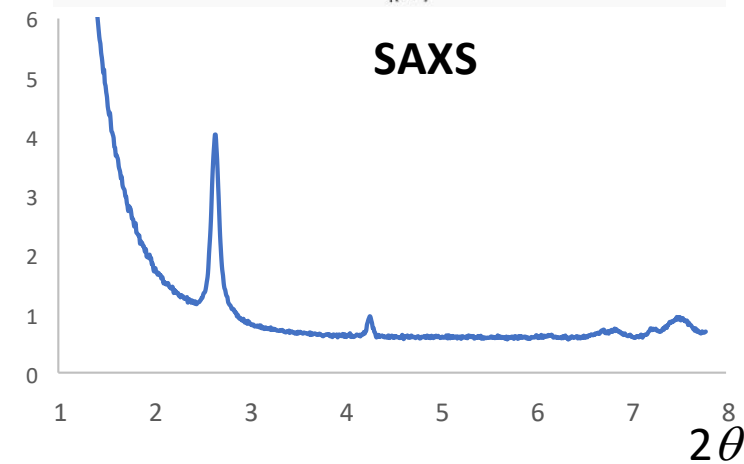
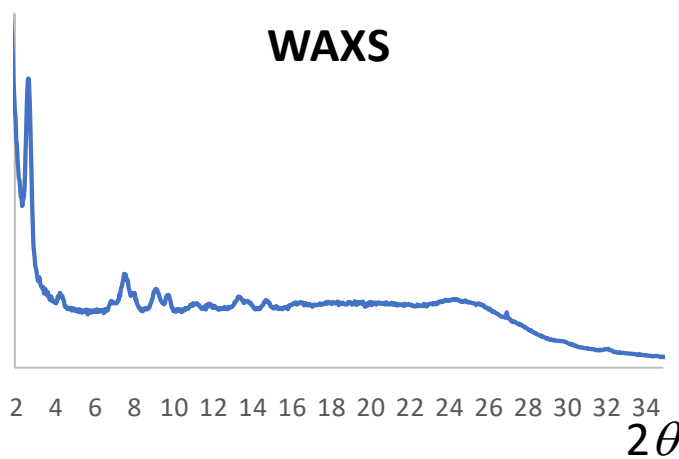
WAXS



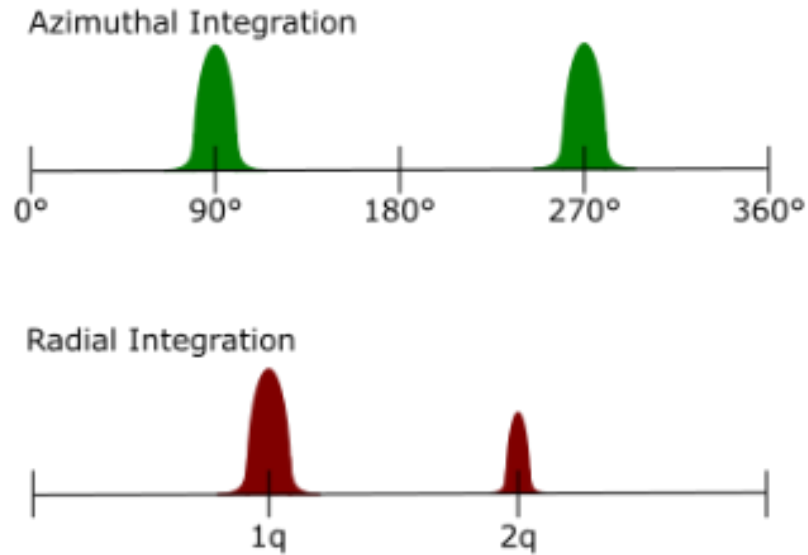
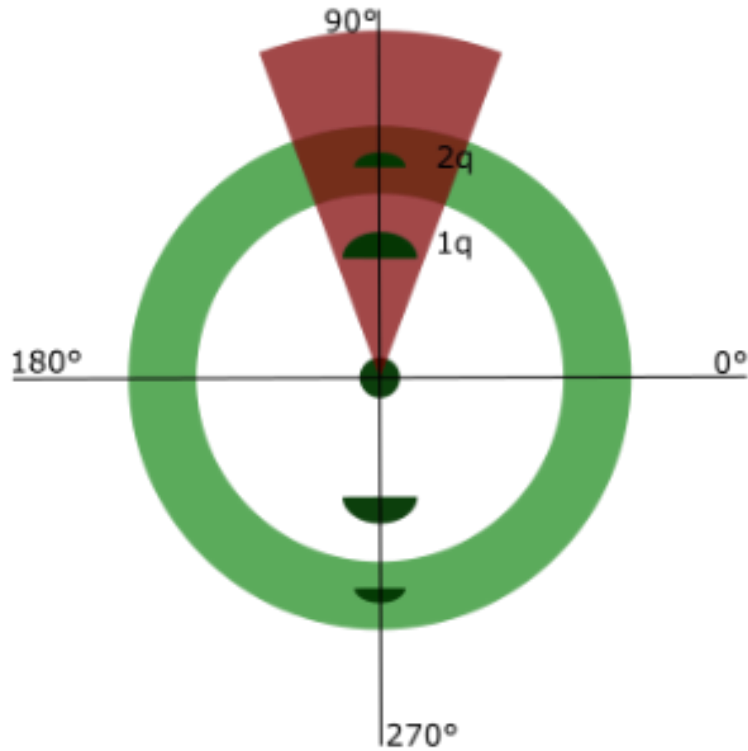
SAXS



PXRD



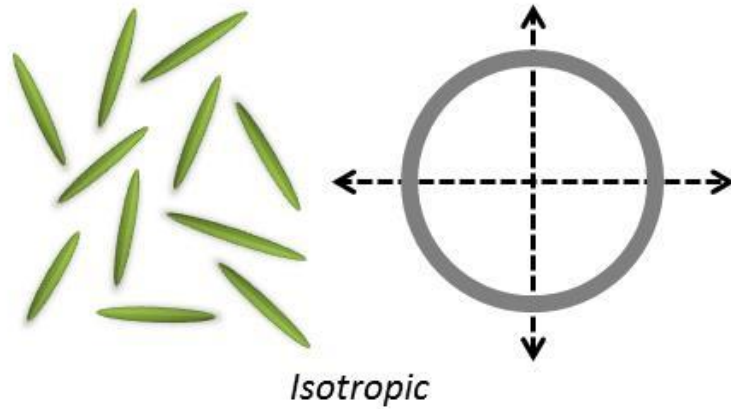
# Azimuthal vs radial integration of the 2D and their resulting 1D scattering profile



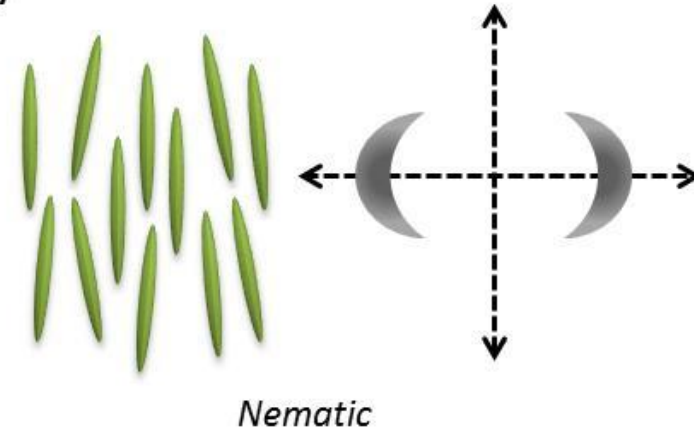
- Azimuthal integration of the scattered intensities provides a distribution of LC preferential orientations
- Full width at half maximum, FWHM, is used to calculate the crystallite orientation

# X-ray scattering of liquid crystals

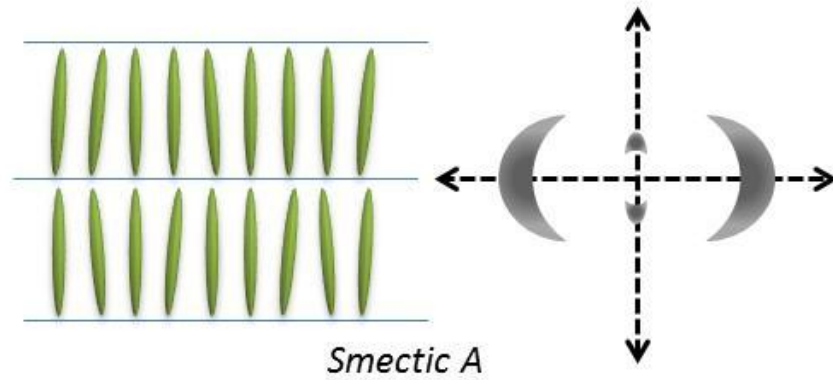
(a)



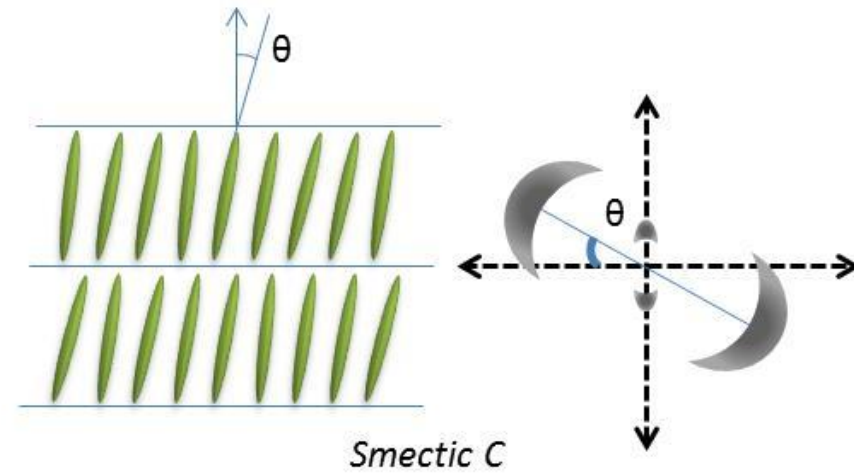
(b)



(c)



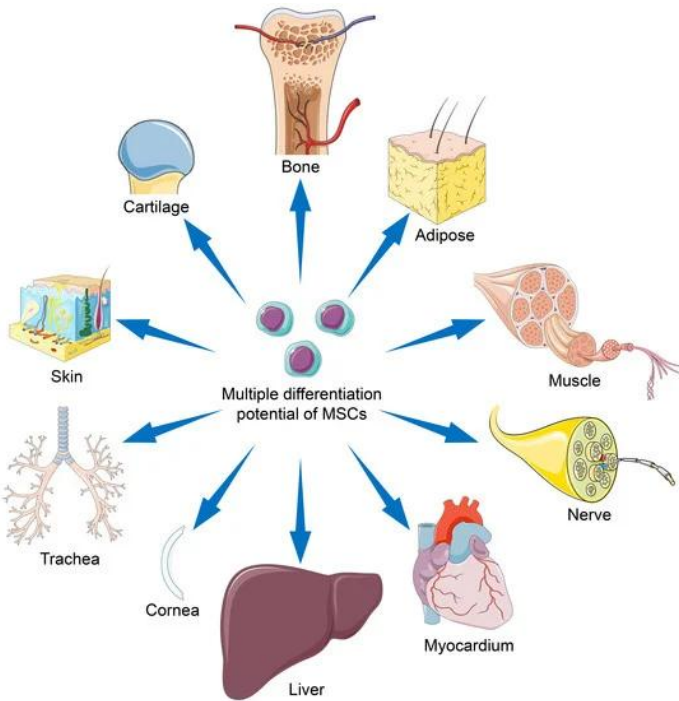
(d)





# In search of a tunable scaffold

Stem cell differentiation is a highly regulated process, but the ECM plays a key role



Han et al., *Cells*. (2019)



Nathalie Singh



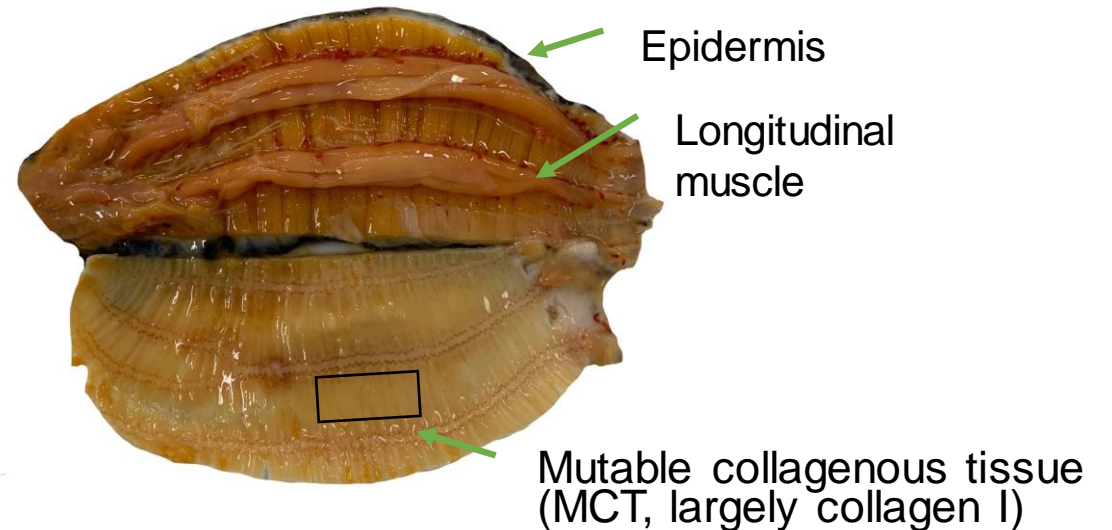
Prof. Matt Harrington



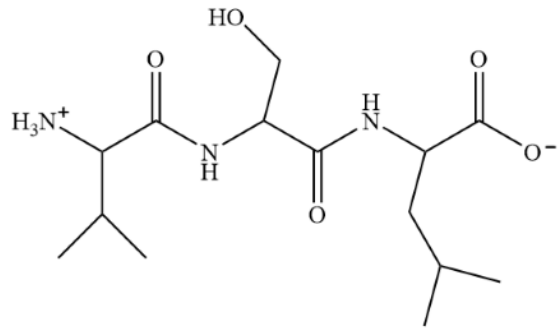
Low stiffness



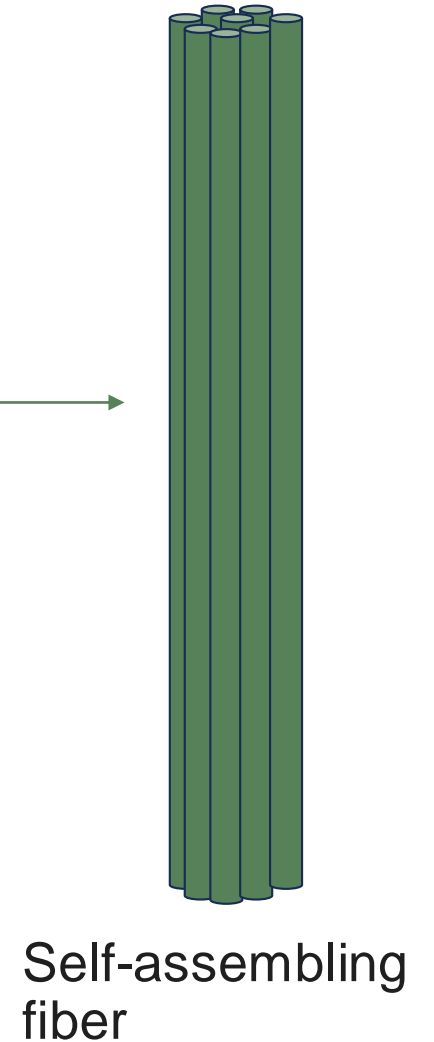
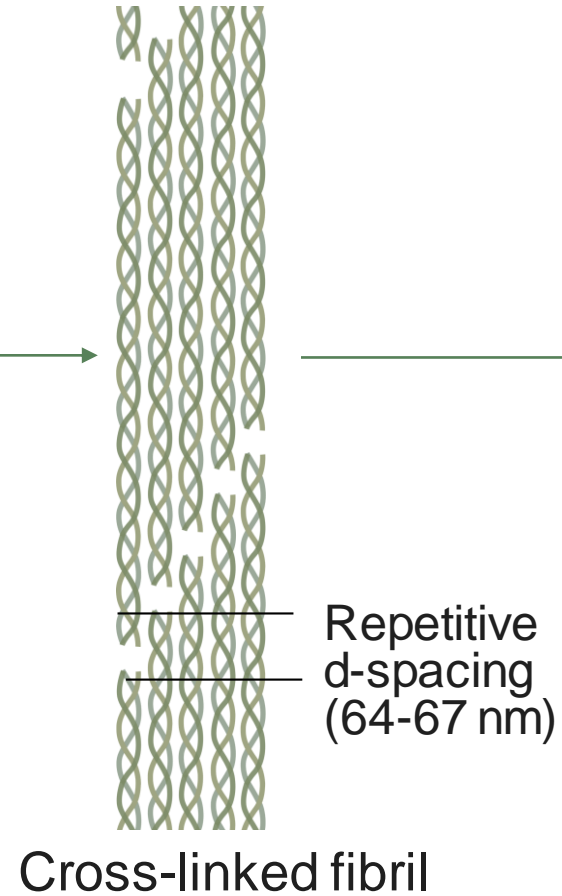
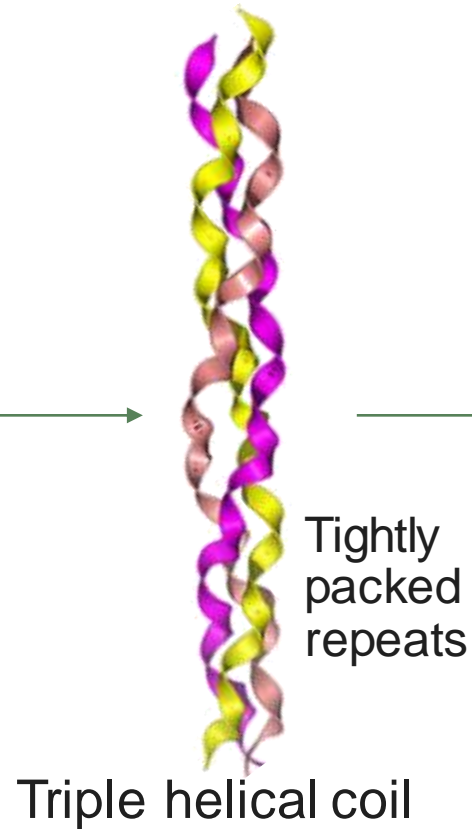
High stiffness



# Collagen I structure



1° aa sequence:  
G-X-Y repeats



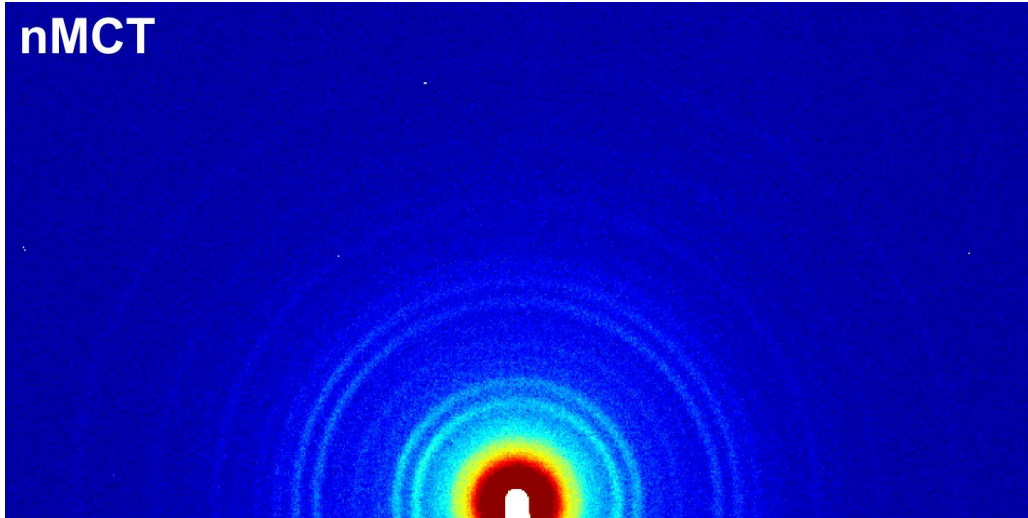


# SAXS of MCT (SDD: 576 mm)

SAXS probes the d-spacing (gap & overlap periodicity) between collagen I triple helices as they assemble into triple helices

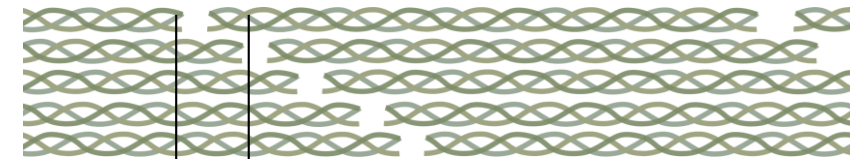
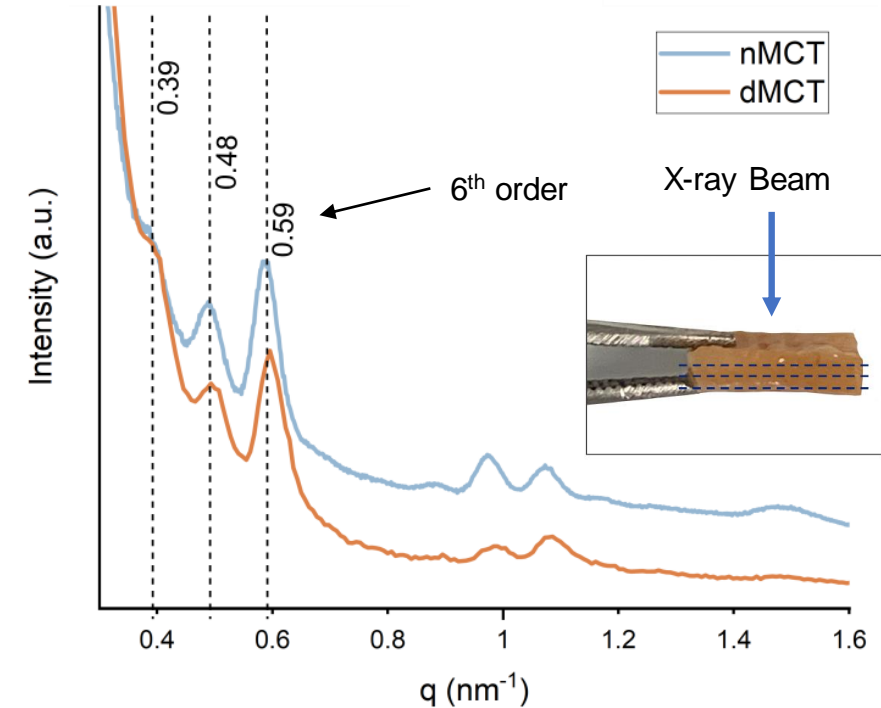
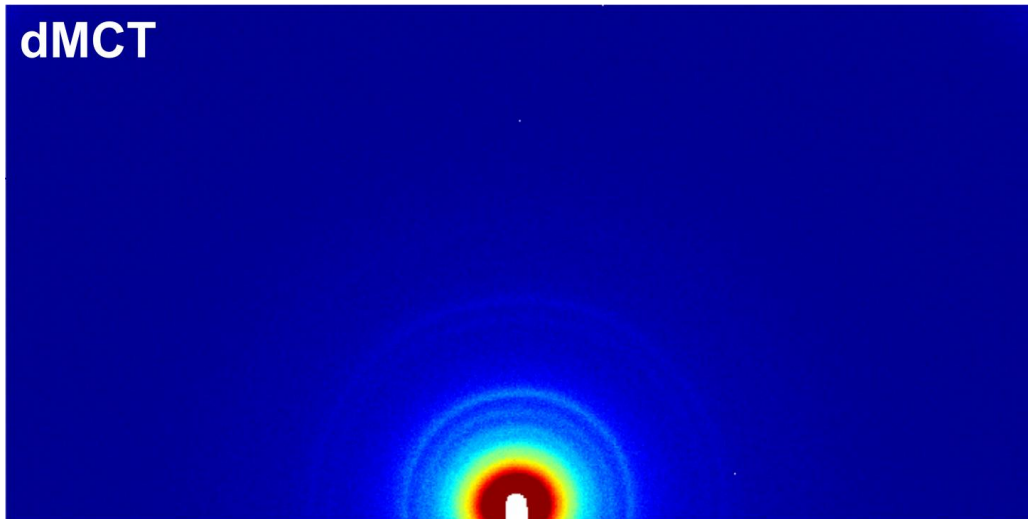
native tissue

nMCT



decellularized tissue

dMCT

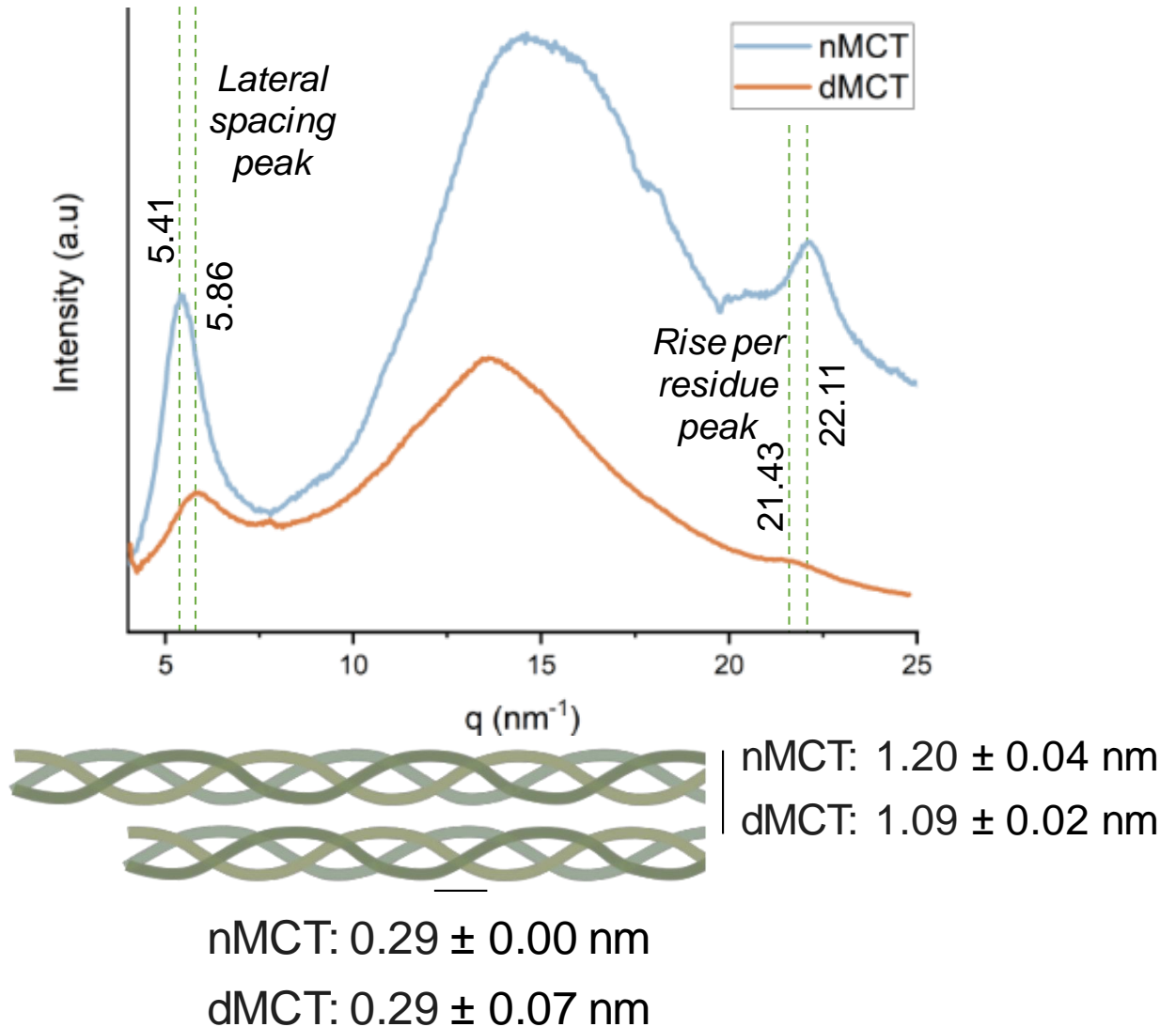
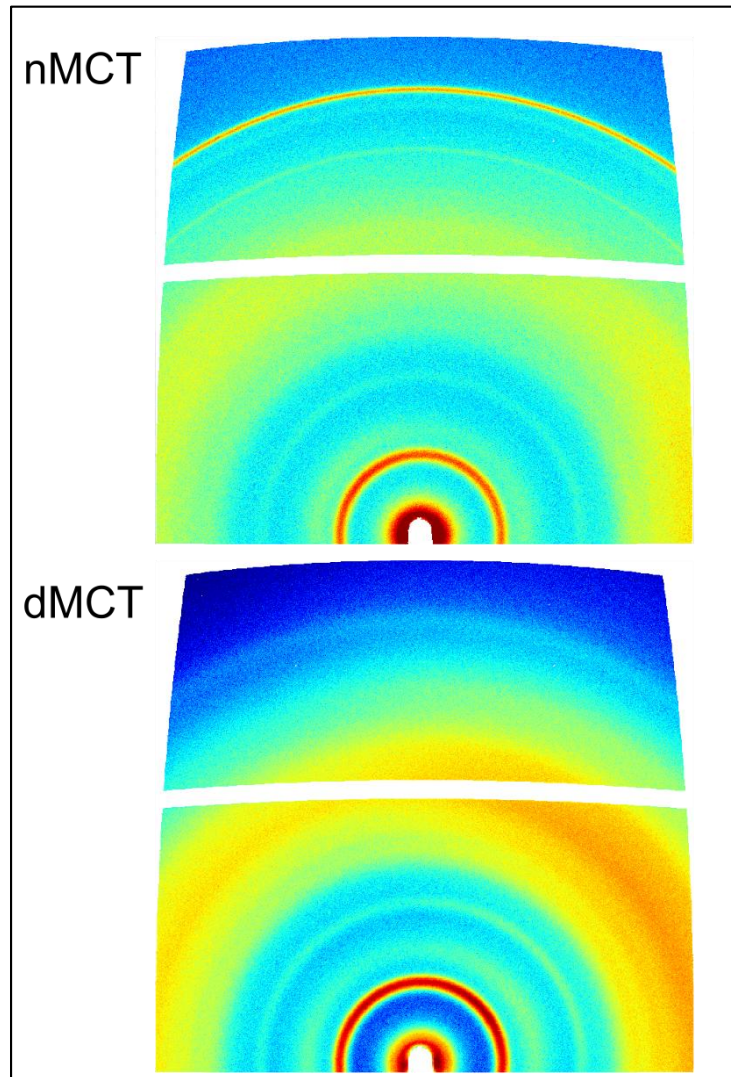


*d*-spacing: nMCT:  $65.38 \pm 0.28$  nm

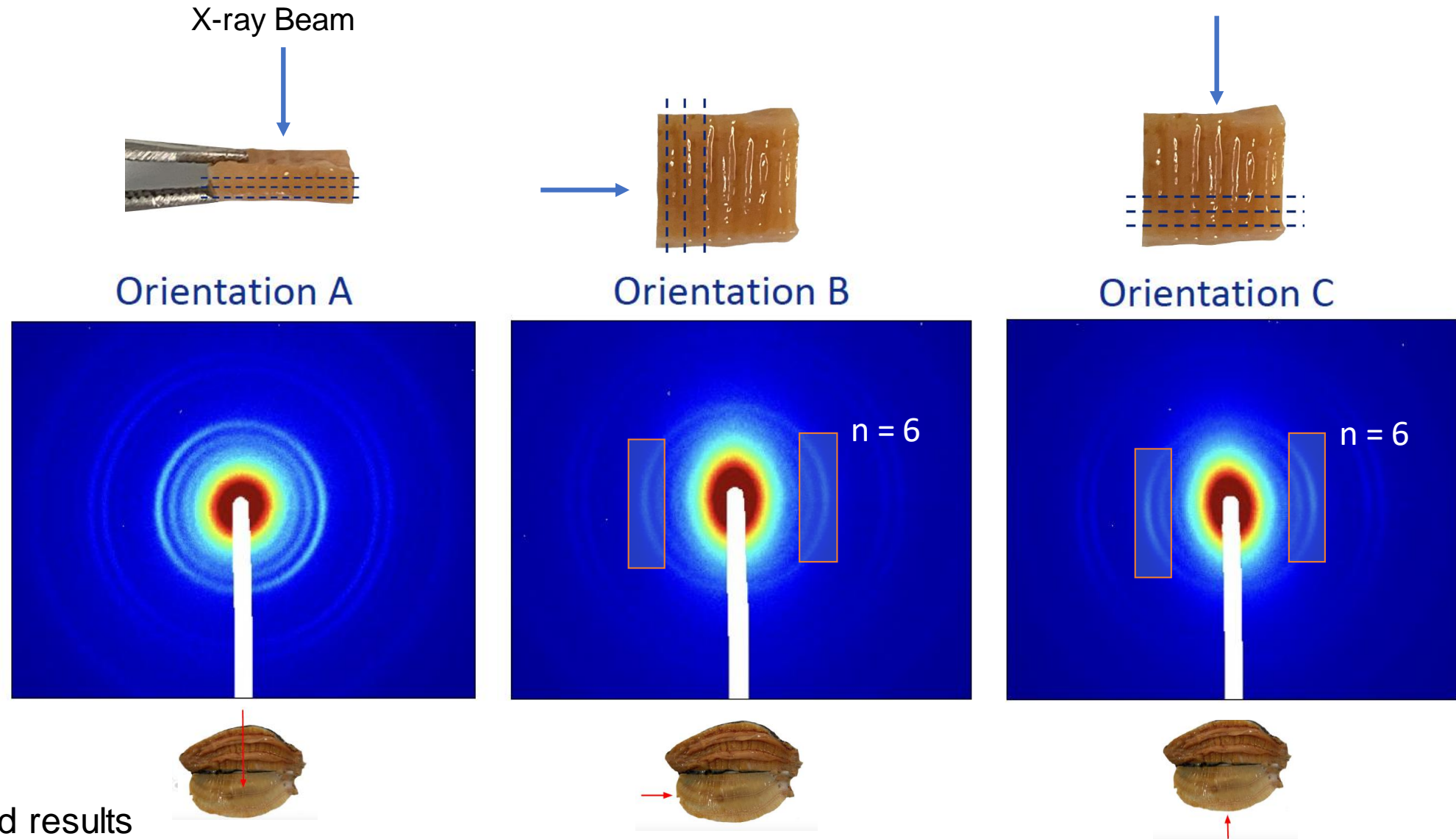
dMCT:  $63.48 \pm 0.26$  nm

# WAXS of MCT (SDD 113 mm)

WAXS probes the rise per residue spacing **within** the triple helix and the lateral spacings **between** triple helices

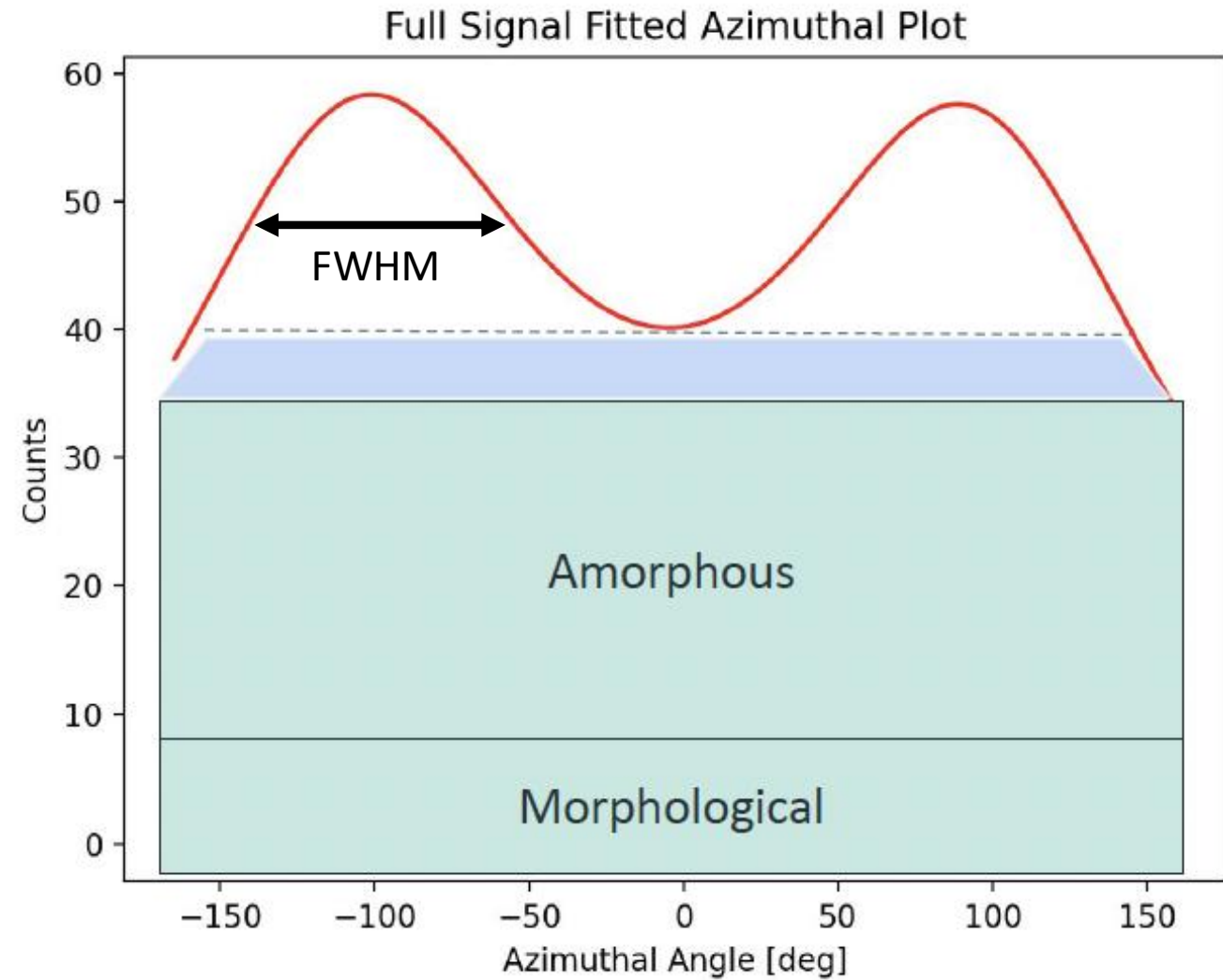
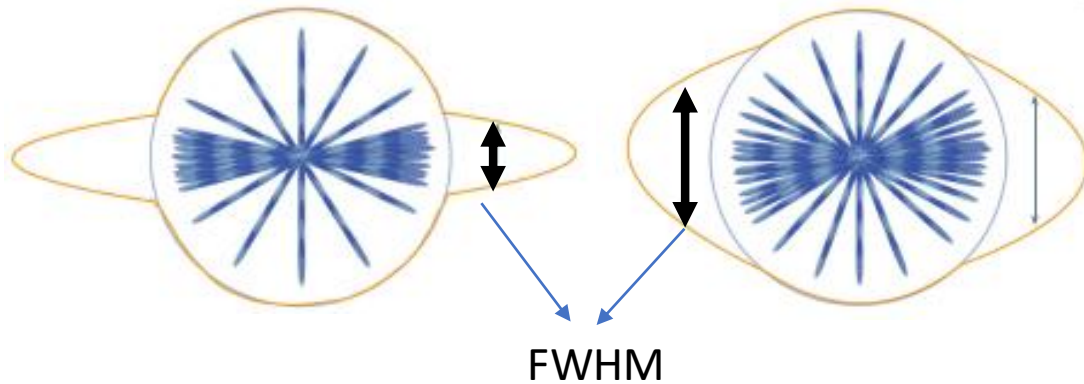
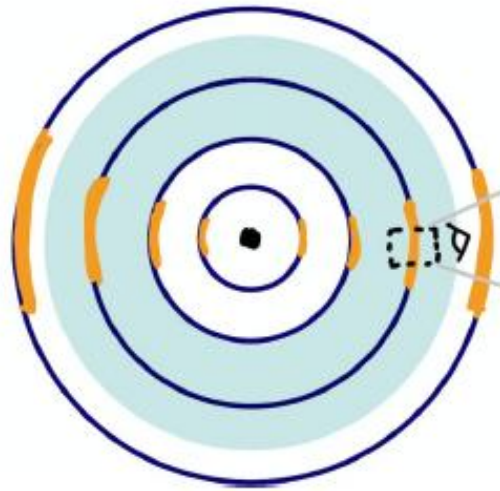
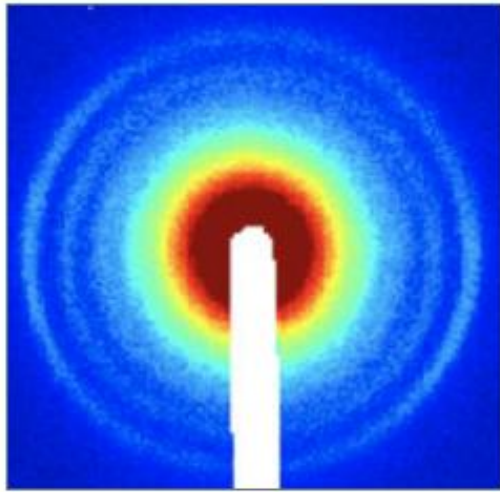


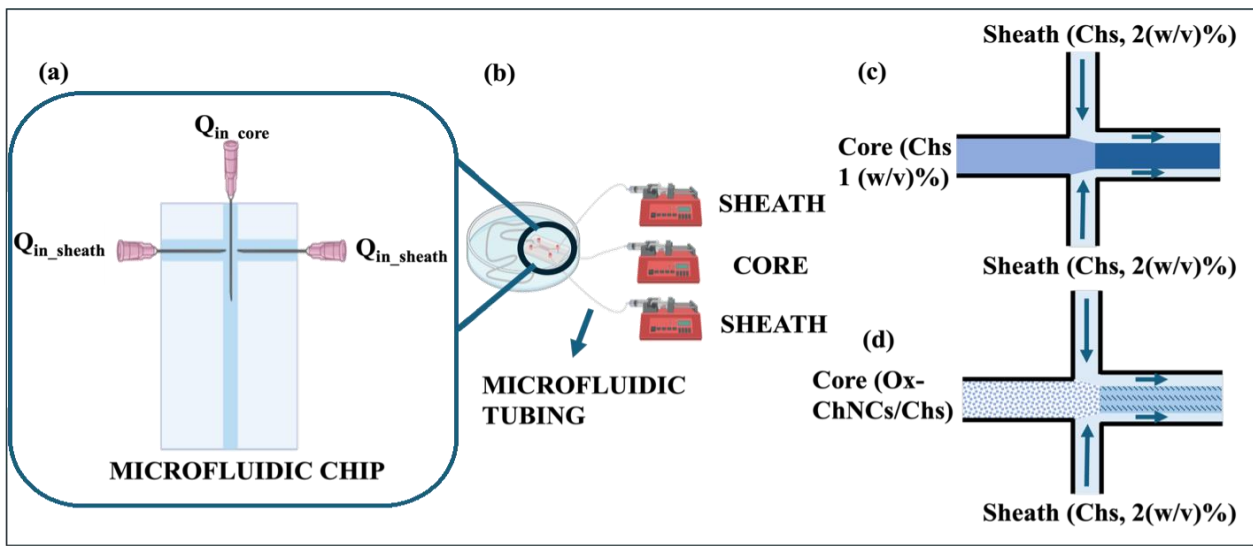
# Preferential orientation of Fibrils



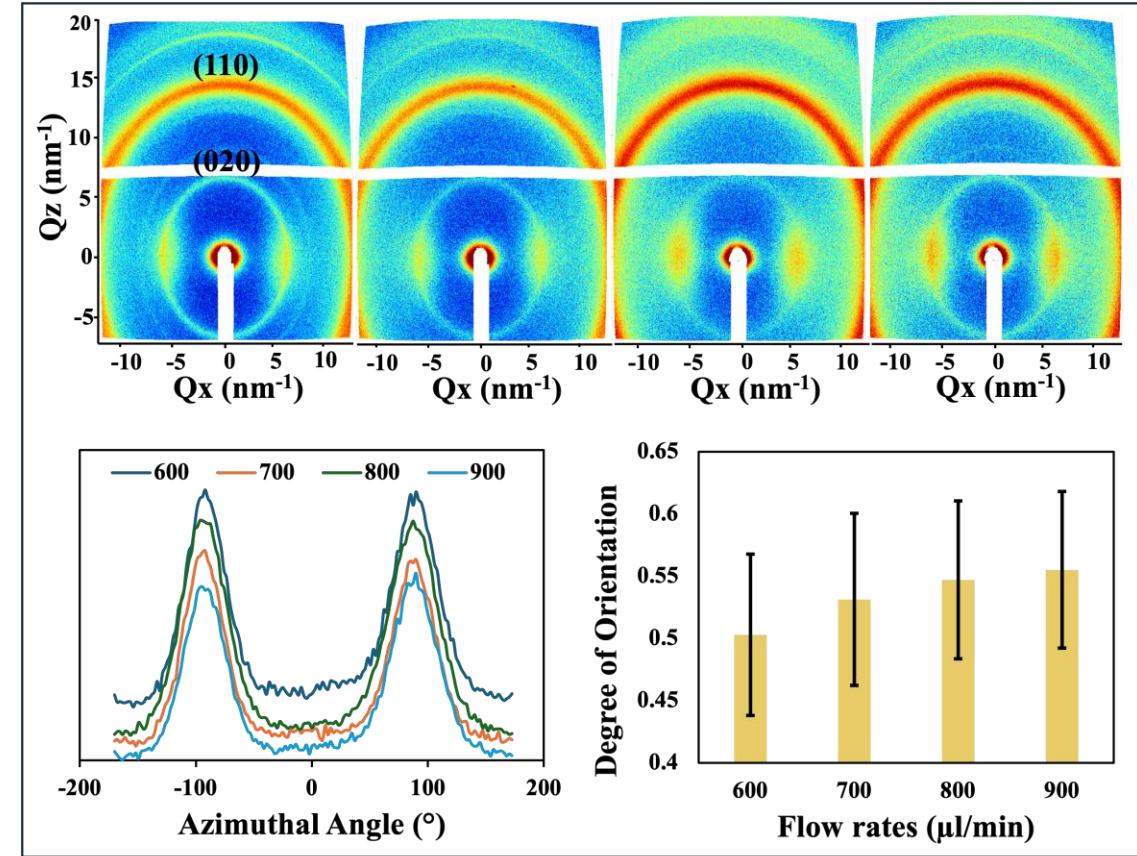


# Radial and Azimuthal SAXS data





# Chitosan Fibers



Ishneet Kaur



Prof. Audrey Moores



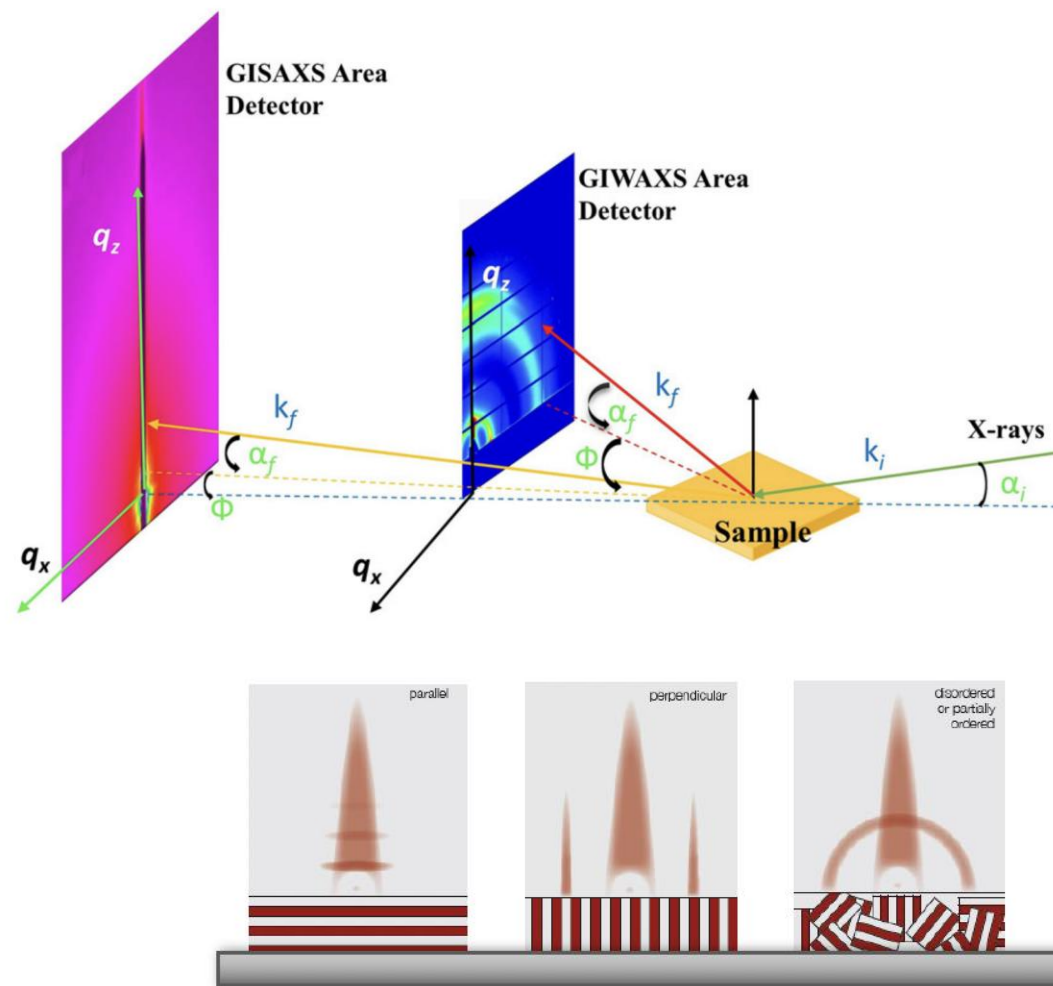
# Grazing-incidence small- & wide-angle X-ray scattering (GISAXS & GIWAXS)

## GISAXS:

- Domain size
- Domain shape
- Phase separation
- Phase purity

## GIWAXS:

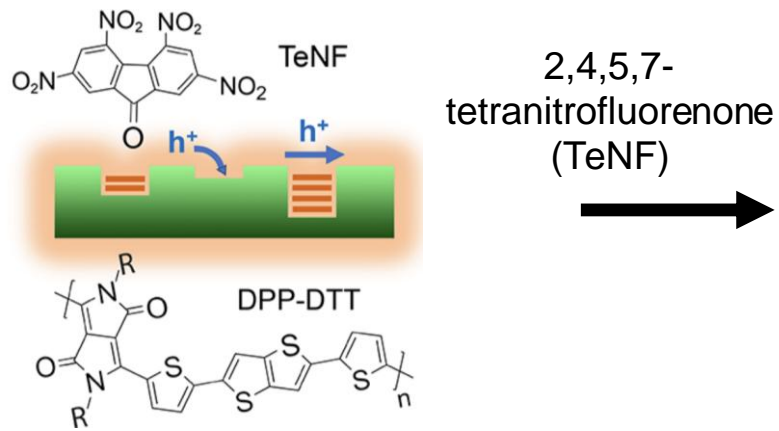
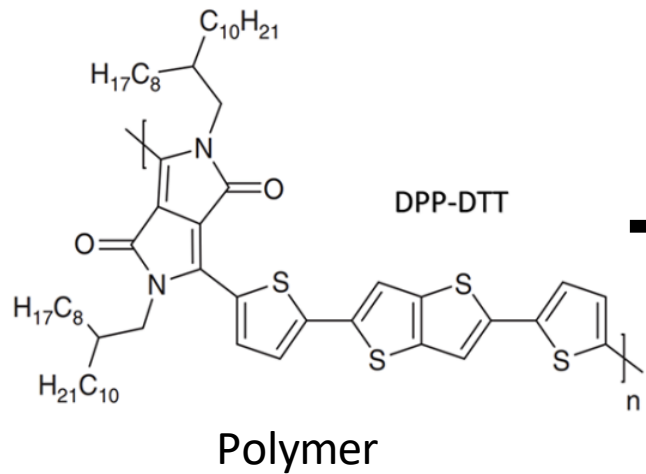
- Crystallinity
- Crystal orientation
- Crystal size
- Interlayer spacing



Sample holder is tilted between  $0.05$ - $0.5^\circ$



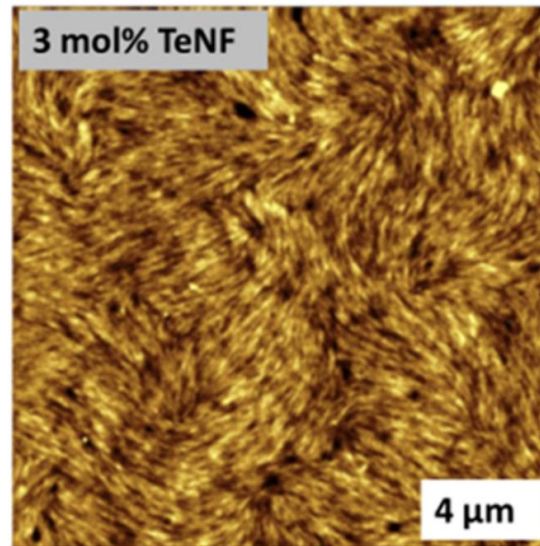
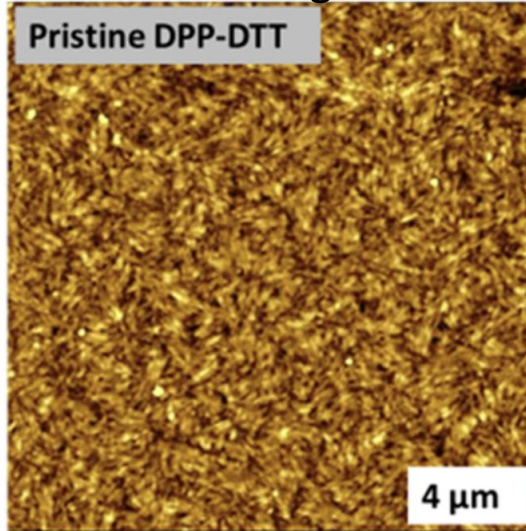
# Effect of doping nitromolecules on thin film: AFM



Doped transistor (thin film)

Che *et al.* *Chem. Mater.* **2021**, 33, 2937–2947

AFM images

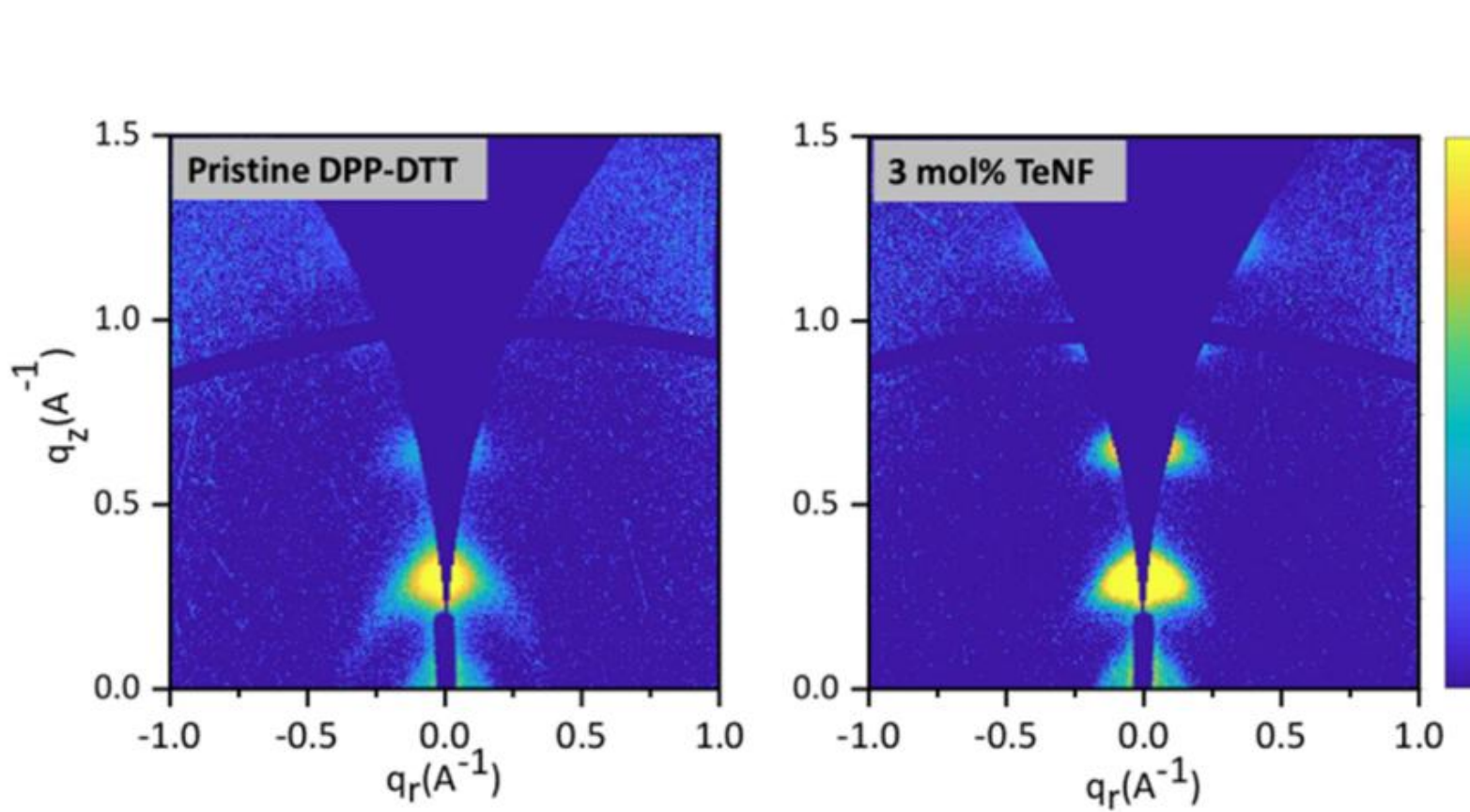


Prof. Dmytro  
Perepichka

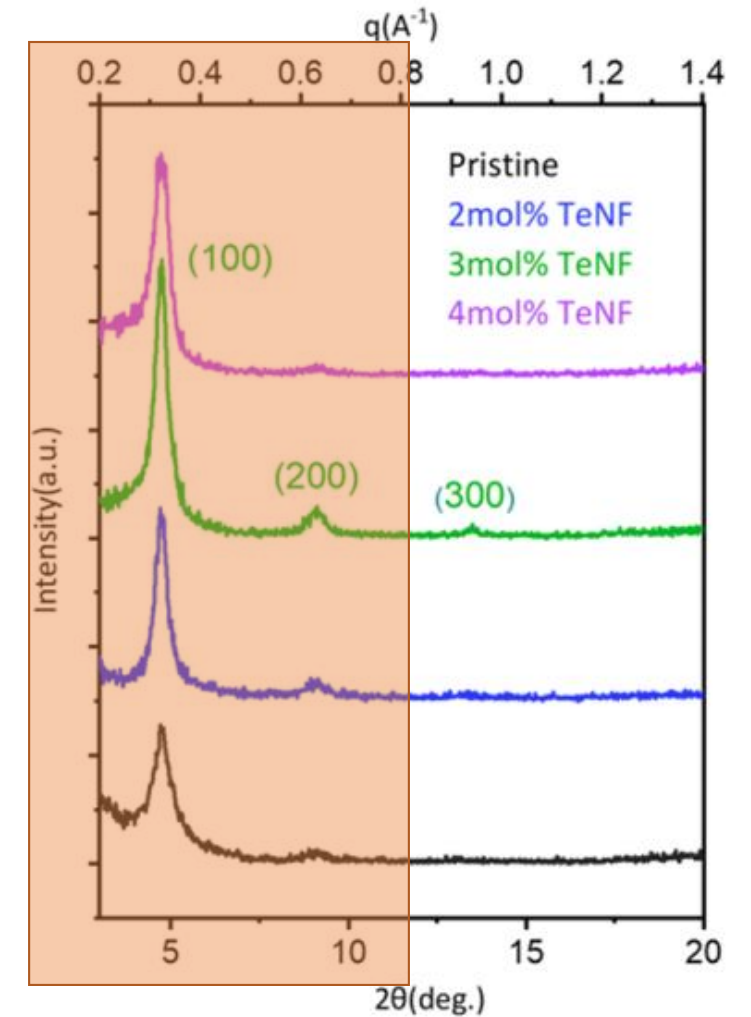


Dr. Pegah Ghamari

# The effect of doping on the thin film morphology: GIWAXS

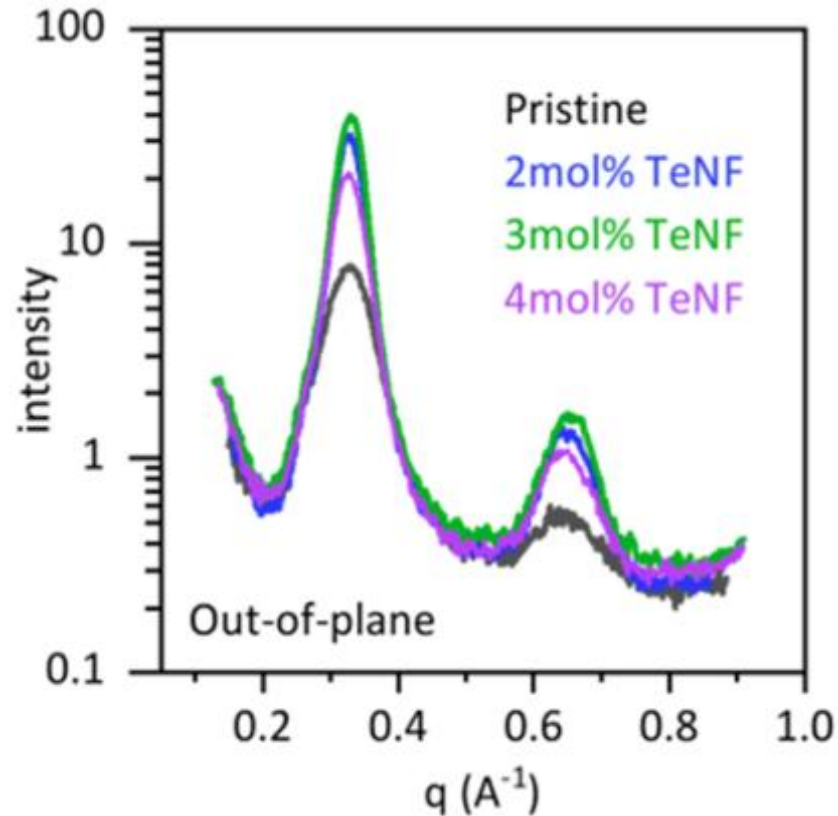


Out-of-plane GIWAXS patterns extracted along the  $q_z$  direction



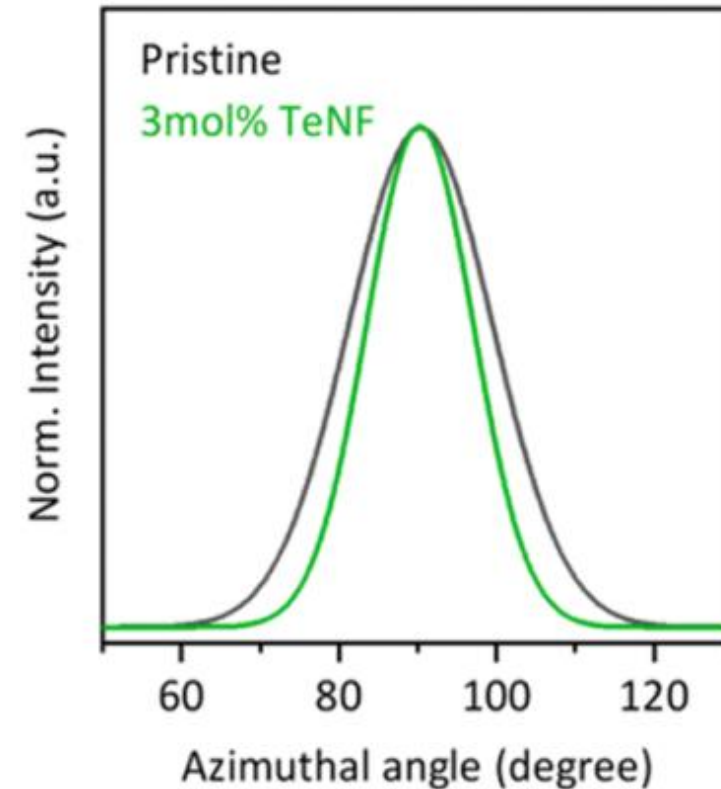
# The effect of doping on the thin film morphology: GIWAXS

Selected region



An increase in crystallinity was achieved by adding 3mol% TeNF

Azimuthal angle to determine size and shape



A higher degree of parallelism



# Acknowledgements



Elaa Ben Fredj



Jean-Luc Brousseau



Semih Gulec

Harrington's lab  
Marić's lab  
Perpichka's lab  
Andrews's lab  
Robert's lab  
Howarth's lab

Thank  
you



**Anton Paar**



**McGill**  
UNIVERSITY



**MCGILL CHEMISTRY**  
CHARACTERIZATION