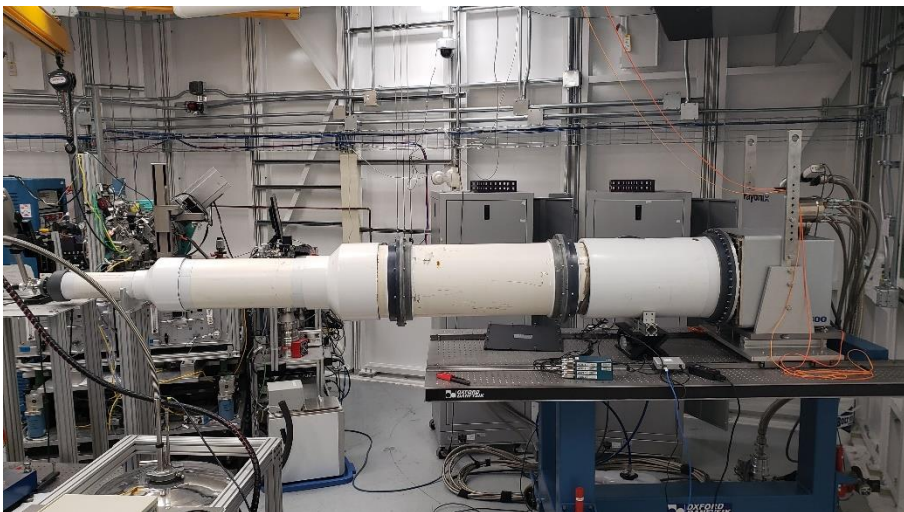
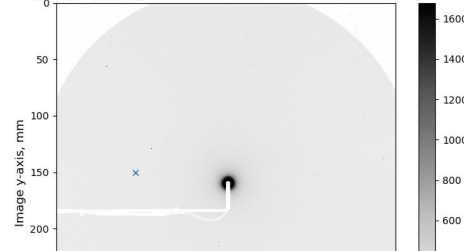


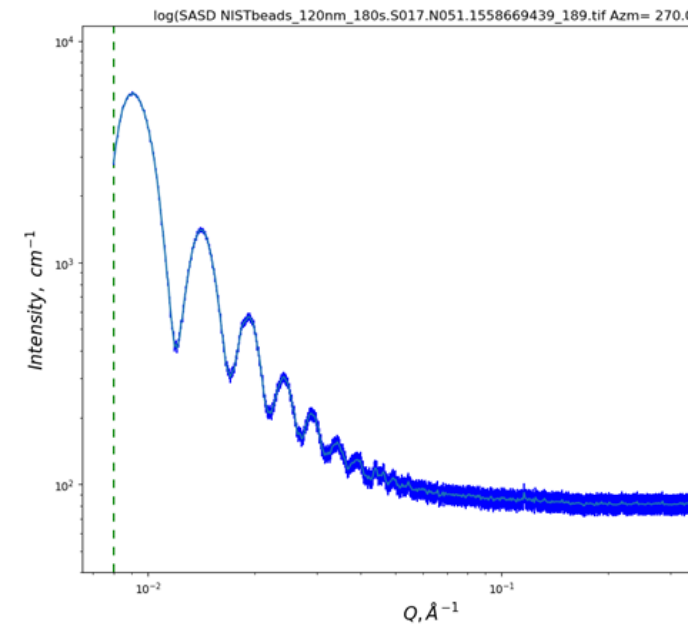
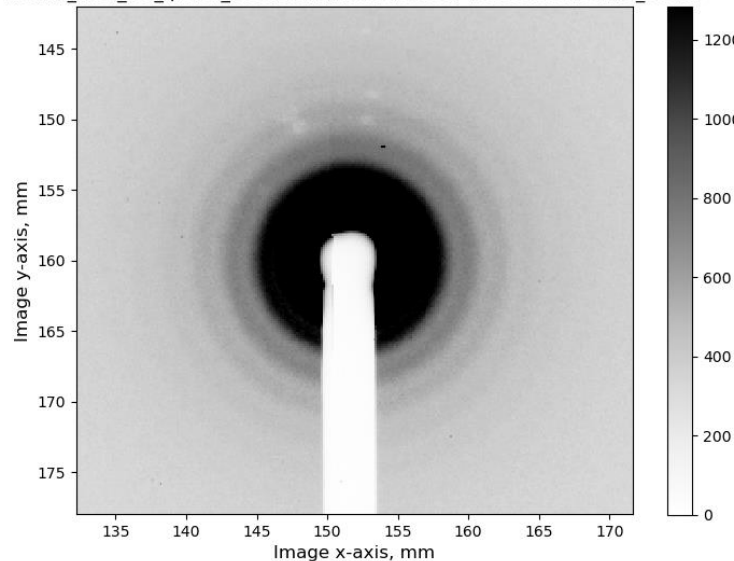
# An Introduction to Small Angle X-ray Scattering (SAXS)



120nm\_NIST\_std\_quartz\_300s.S014.N051.4056907486906665264173\_365.tif



120nm\_NIST\_std\_quartz\_300s.S014.N051.4056907486906665264173\_365.tif



Adam Leontowich  
CLS XRD school, Thursday, June 20, 2024



Canadian Light Source  
Centre canadien de rayonnement synchrotron

**The small angle X-ray scattering (SAXS) region**

**Instrumentation, sample prep, data collection and reduction**

**Basic data processing with examples from BXDS**



# Brockhouse X-ray Diffraction and Scattering (BXDS) sector

## The lower energy diffraction and scattering side-bounce beamline for materials science at the Canadian Light Source

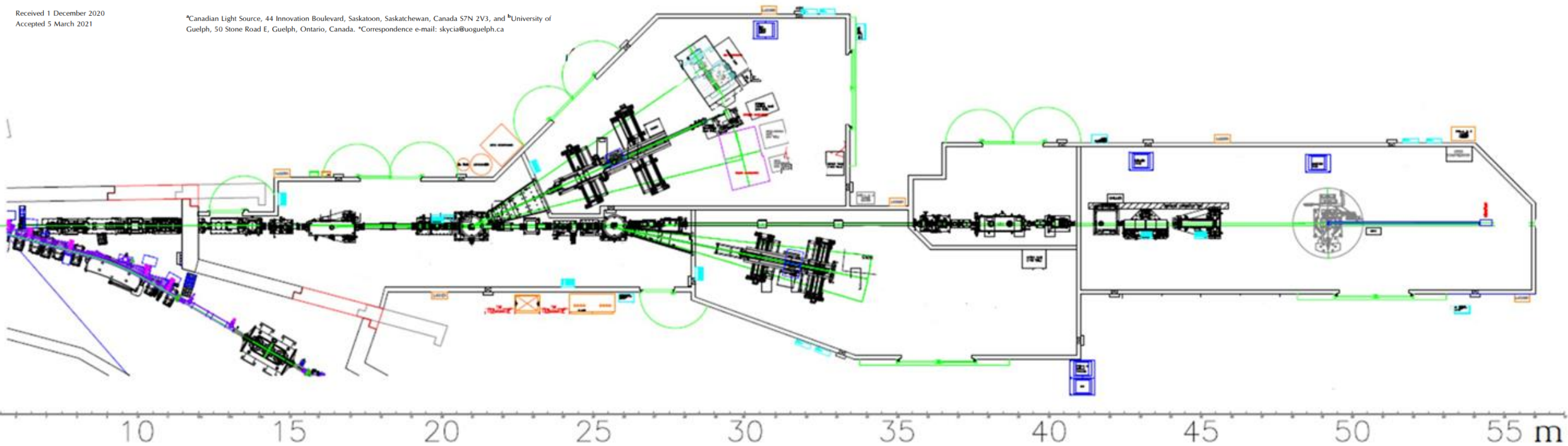
Adam F. G. Leontowich,<sup>a</sup> Ariel Gomez,<sup>a</sup> Beatriz Diaz Moreno,<sup>a</sup> David Muir,<sup>a</sup> Denis Spasyuk,<sup>a</sup> Graham King,<sup>a</sup> Joel W. Reid,<sup>a</sup> Chang-Yong Kim<sup>a</sup> and Stefan Kycia<sup>b\*</sup>



Received 1 December 2020  
Accepted 5 March 2021

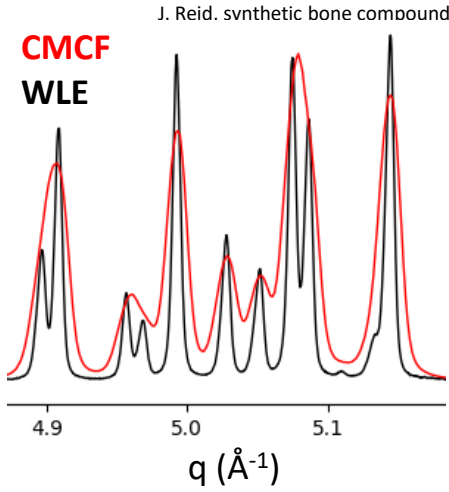
<sup>a</sup>Canadian Light Source, 44 Innovation Boulevard, Saskatoon, Saskatchewan, Canada S7N 2V3, and <sup>b</sup>University of Guelph, 50 Stone Road E, Guelph, Ontario, Canada. \*Correspondence e-mail: skycia@uoguelph.ca

3 “hard X-ray” beamlines (5 – 100 keV)



# WLE beamline

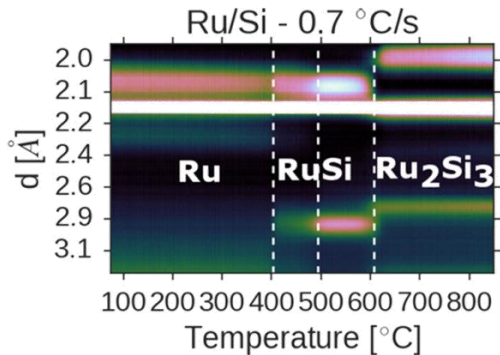
## High-resolution powder diffraction



- Peak shape analysis
- Complex mixtures
- Complex structures

IBM (rapid thermal annealing to 1100 °C)

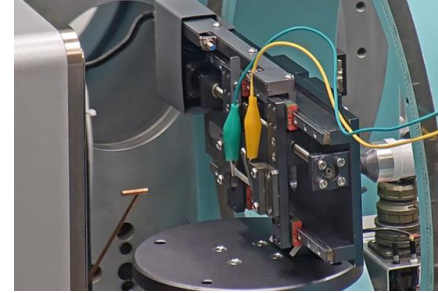
- Thin film studies: XRD, resistivity, roughness, under ultra-high purity gas



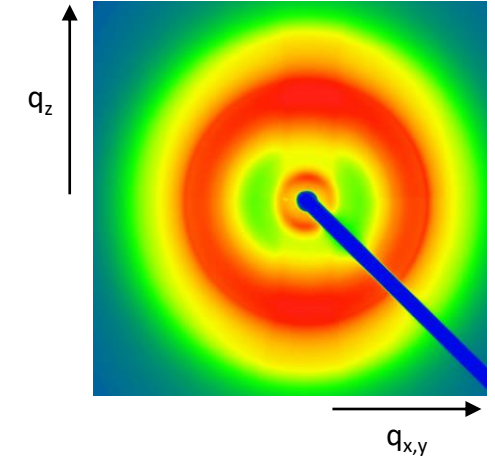
S. Dey et al., J. Vac. Sci. Technol. A 35, 03E109 (2017)

# WAXS (XRD with area detector)

Biopolymers for bandages  
Youchao Teng, Yimin Wu, U. Waterloo



Measure WAXS, voltage and current while stretching (0 – 100 N)



- Texture on surfaces
- Degree of orientation
- % crystallinity
- Speed/in-situ

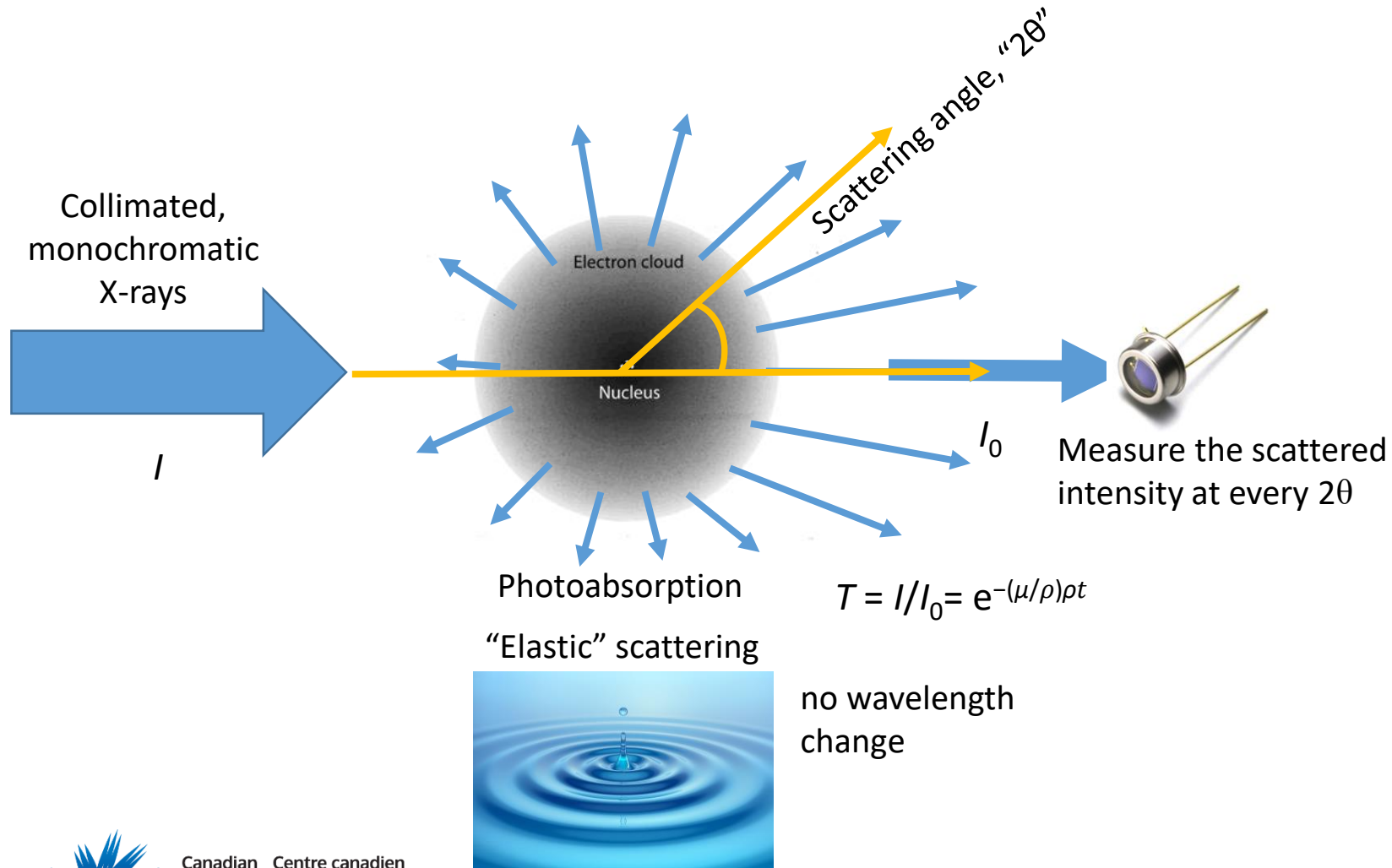


SAXS/WAXS

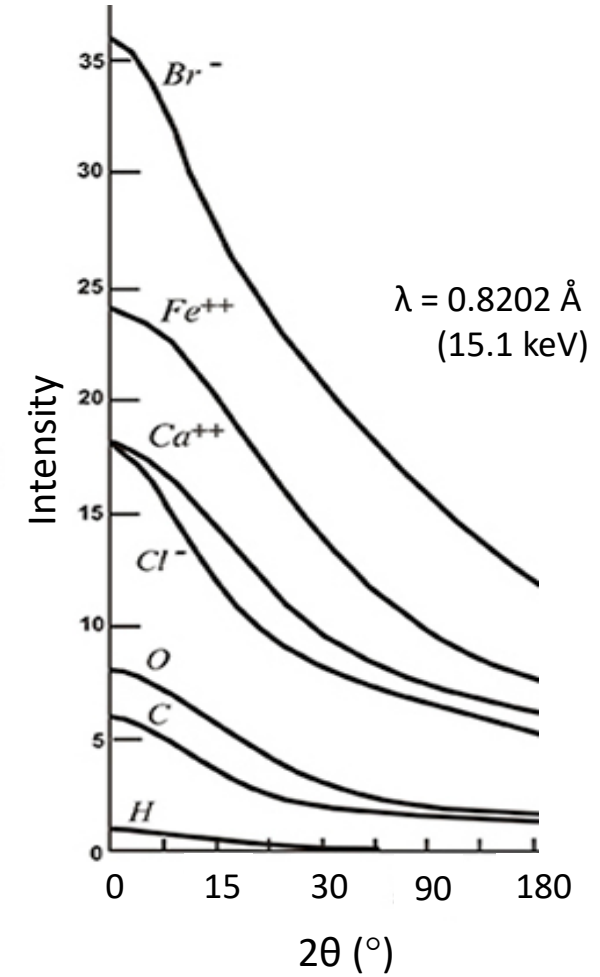
SAXS has been running since February 2021

*No dedicated SAXS beamline at CLS*

# X-ray scattering



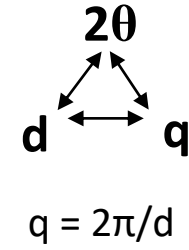
Atomic form factors  
(or atomic scattering factors)



# From scattering to diffraction

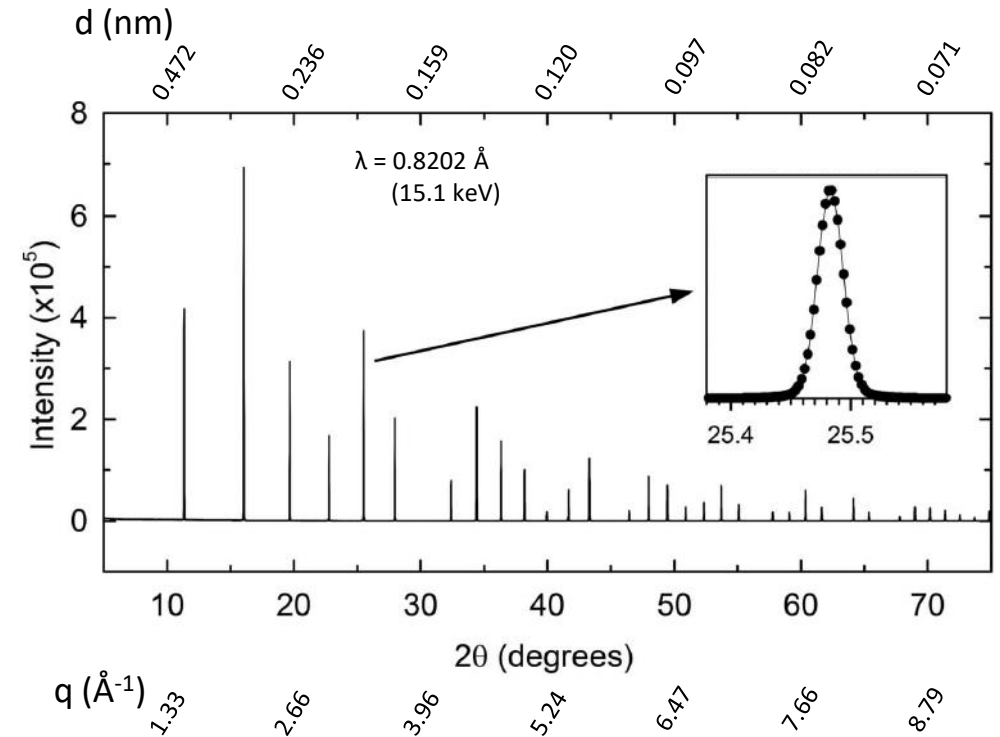
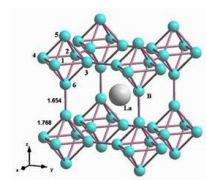
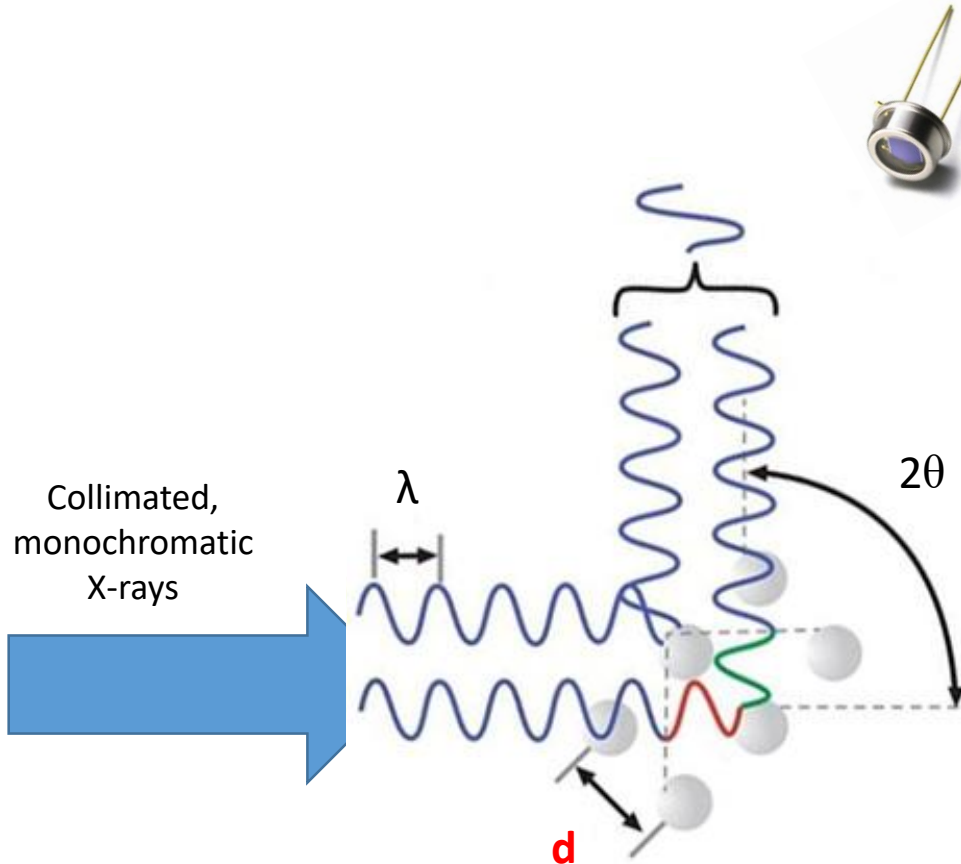
“Bragg” diffraction

$$n\lambda = 2d\sin\theta$$



$$q = (4\pi\sin\theta)/\lambda$$

Constructive interference channels  
filtered intensity into specific directions

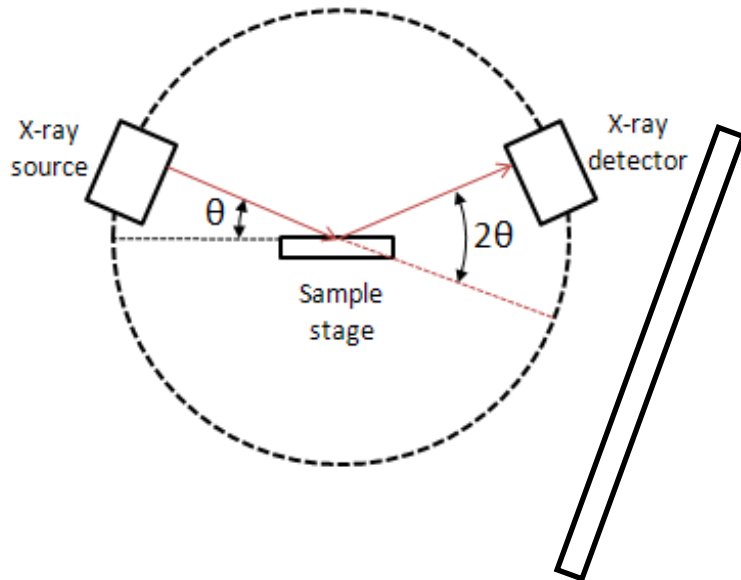


X-ray scattering and diffraction reveals structural order within materials  
on the atomic to 100s of nanometer length scale

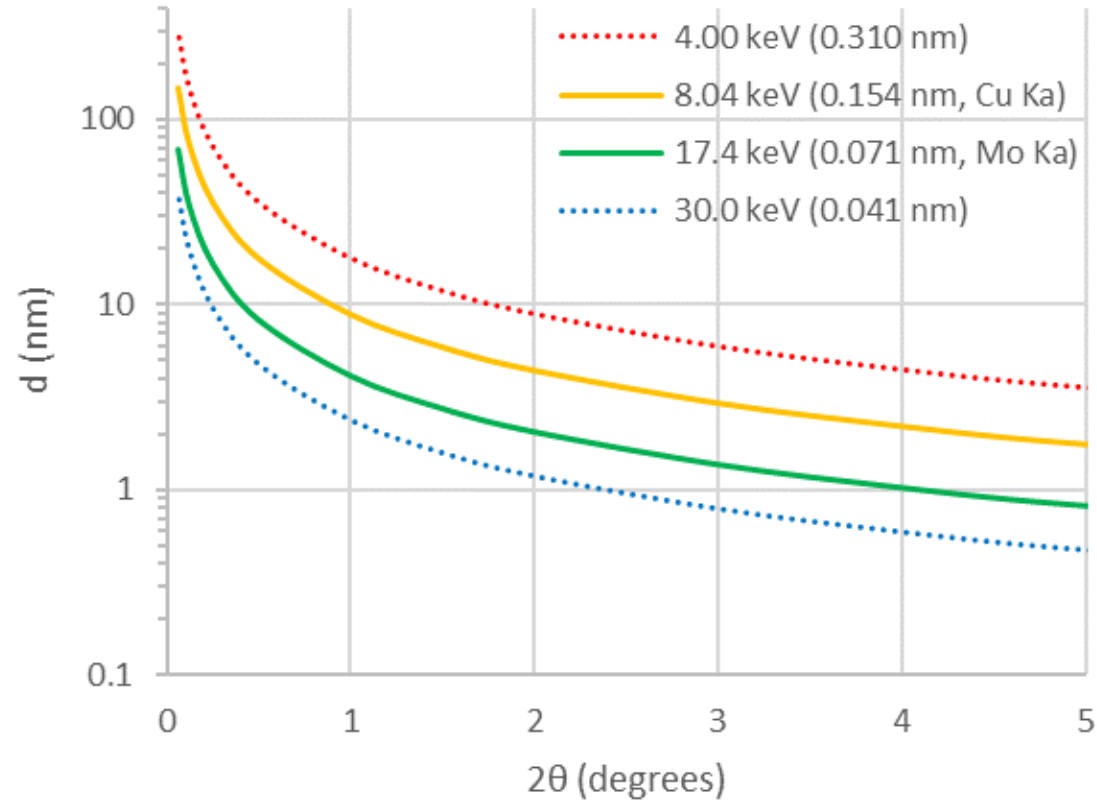


# What is SAXS all about?

Starting from powder diffraction...



$$\lambda = 2d\sin\theta$$



The nano world

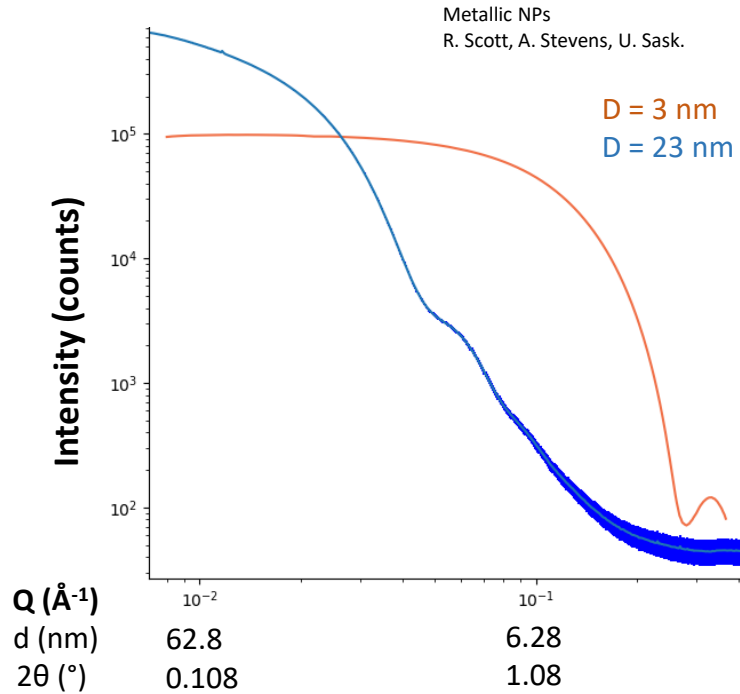
Atomic distances, spacing between planes of atoms, bond lengths

SAXS is generally  $2\theta = 0.05 - 5^\circ$

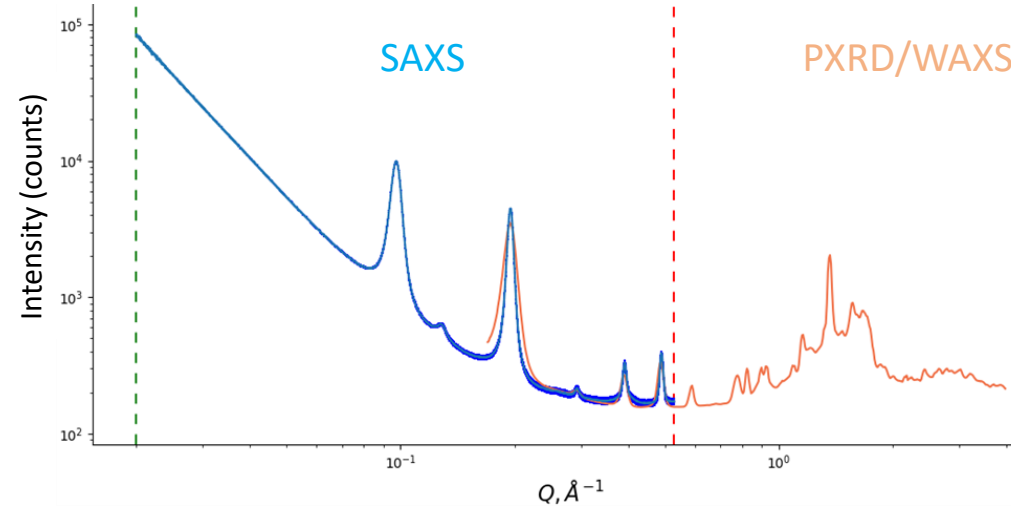
Probe relatively big things (1 - 150 nm) by measuring elastic X-ray scattering at small angles



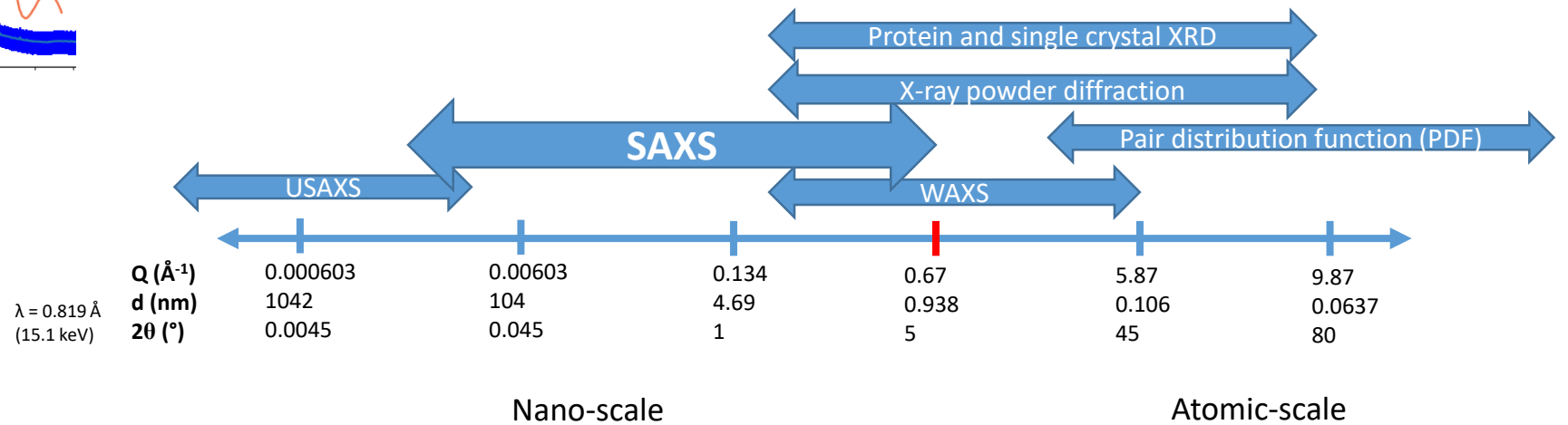
# How is SAXS related to other X-ray diffraction techniques?



- Intensity vs.  $Q$
- log-log plot



Chocolate  
A.G. Marangoni, U. Guelph



Inter-molecular distances and packing arrangements, particle size and shape

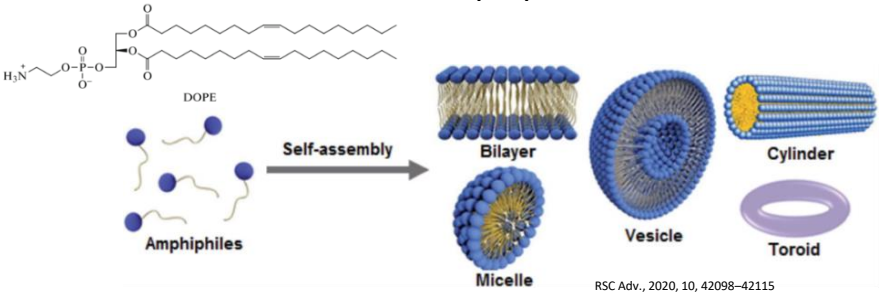
Inter-atomic distances, bond lengths, nearest neighbor atoms  
Crystal structures with atomic resolution





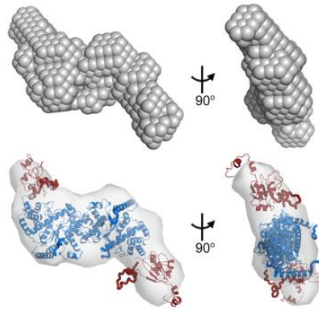
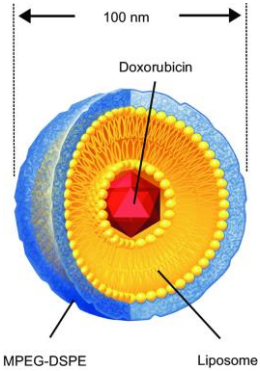
# Applications

## Amphiphiles

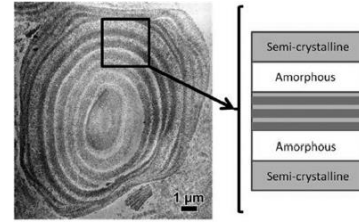


Drug delivery,  
Pharmaceuticals

Micelles,  
emulsions,  
surfactants...



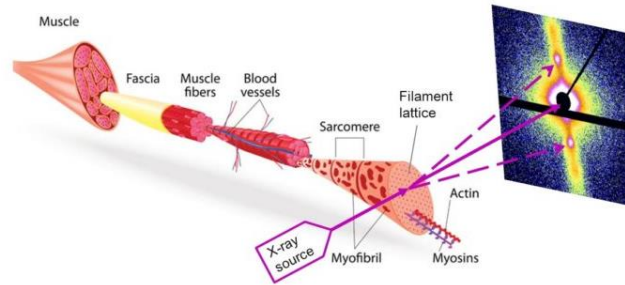
## Food science



S. Perez and E. Bertoft, *Starch* 2010, 62, 389 - 420

## Biomacromolecules

Cellulose  
Collagen  
Chitosan  
  
Proteins

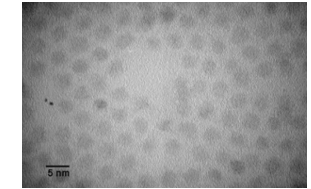
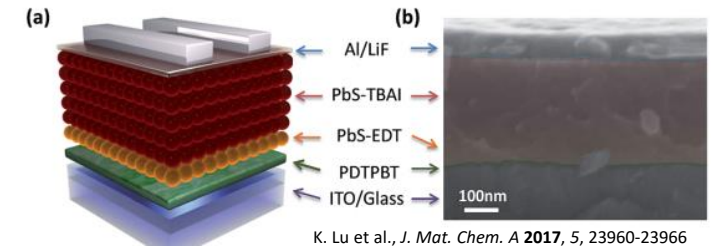


<https://www.techexplorist.com/scientists-investigated-pork-fillet-x-ray-light/11572/>

- 1) Packing, ordering
- 2) Size, size distribution
- 3) Shape



## All kinds of nanoparticles

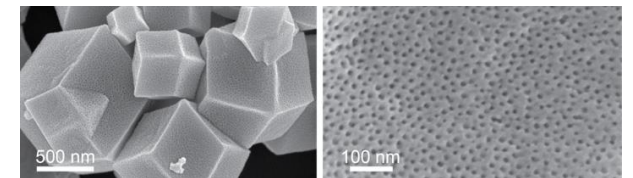


M. Yuan et al., *Adv. Mater.* 2014, 26, 3513-3519

## Nano-porous materials



Geology  
  
Batteries  
  
Zeolites

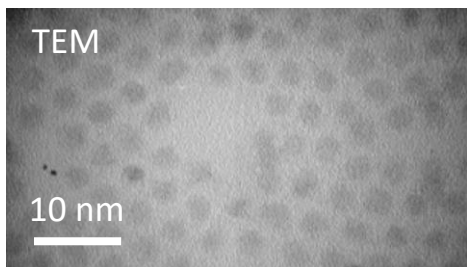


M. Liu, Z.M Hudson, UBC

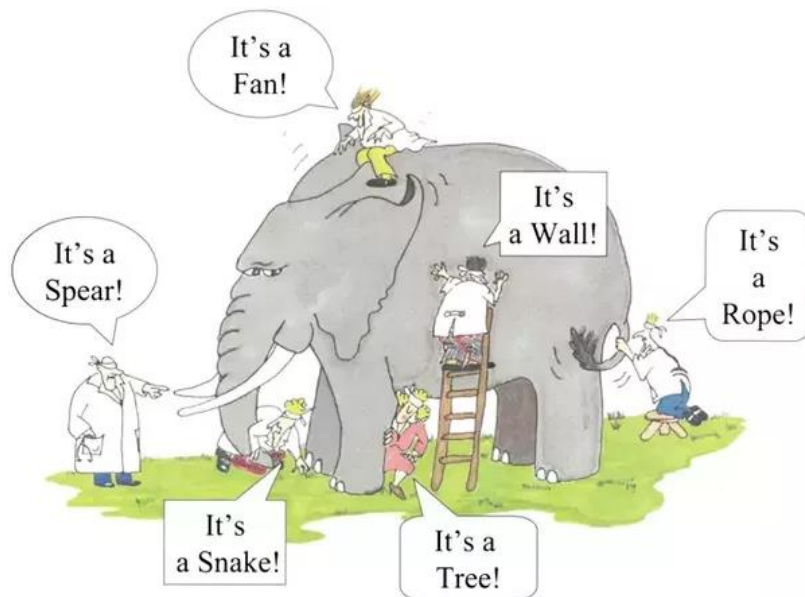


Canadian  
Light  
Source  
  
Centre canadien  
de rayonnement  
synchrotron

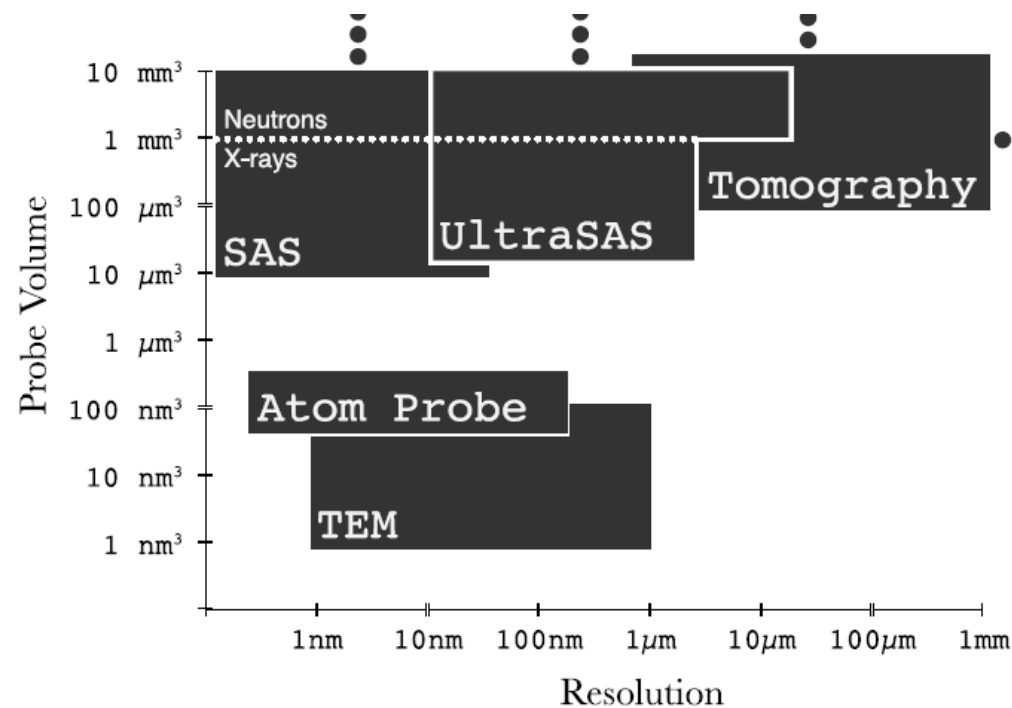
# SAXS is highly complementary to microscopy



M. Yuan et al., Adv. Mater. 2014, 26, 3513–3519



"Blind men and an elephant"



B.R. Pauw, J. Phys.: Condens. Matter 25 (2013) 383201

- FOV decreases as resolution increases
- Observe a small fraction of the complete sample at a time
- Challenging sample prep at smaller size

- SAXS provides nanoscale information **averaged** over the beam volume (~mm<sup>3</sup>)
- Complementary info: shape, folding/unfolding, assembled state in solution



# Instrumentation, sample prep, data collection and reduction

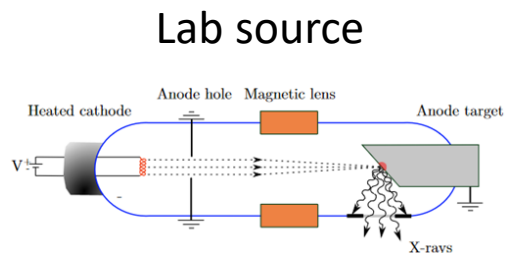


# How to measure small angle X-ray scattering?

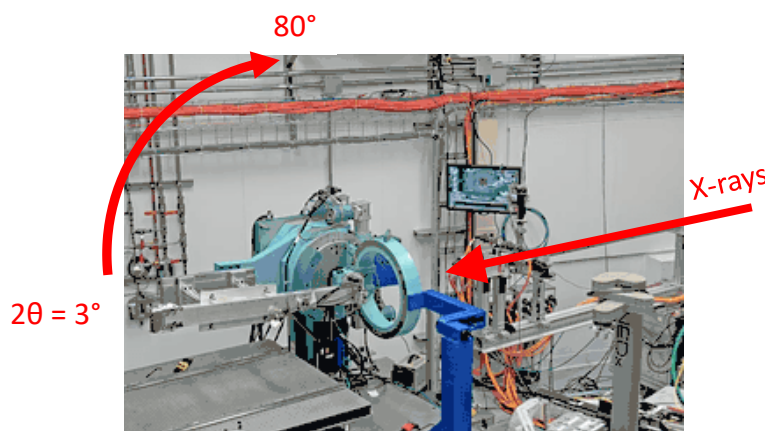
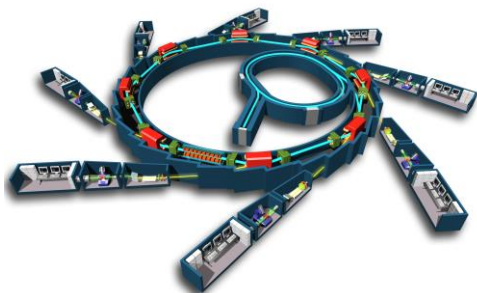
PXRD



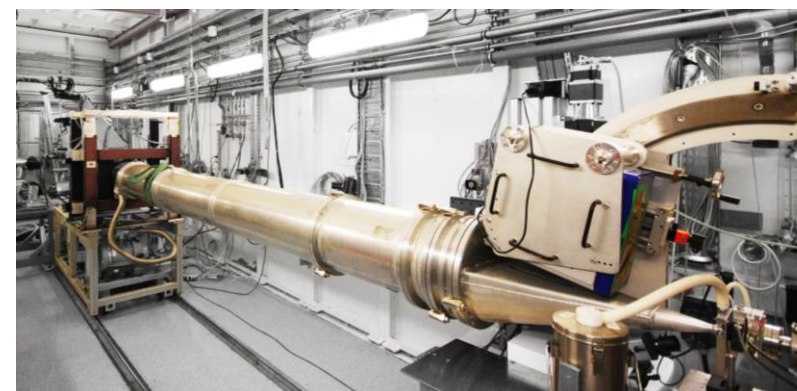
SAXS



Synchrotron

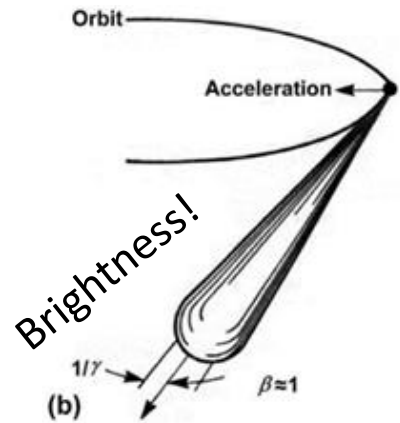
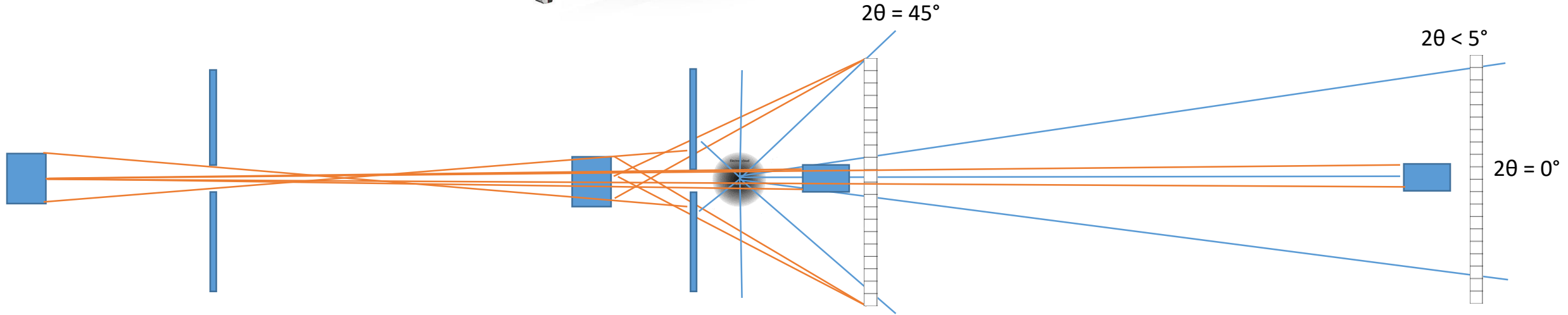


ESRF - BM26



- 1) Special instrumentation required for SAXS
- 2) SAXS instruments are long!

# SAXS instrumentation basics



- More flux (better SNR), weakly scattering objects (organics/biology)
- Better  $q$  resolution
- Time resolved, SAXS mapping
- Choice of many wavelengths

- Highly collimated hard X-ray beam (multiple slits)
- Large area detector with beamstop
- Space for >2 m sample to detector distance
- Some ability to change detector distance (SAXS/WAXS)
- All or most components in vacuum

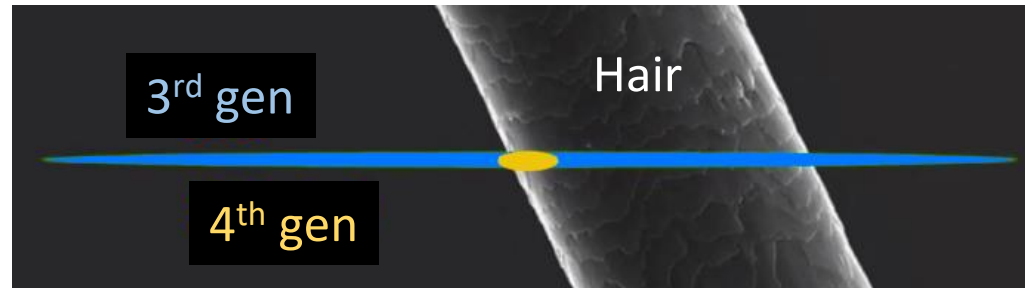


# State of the art instrumentation

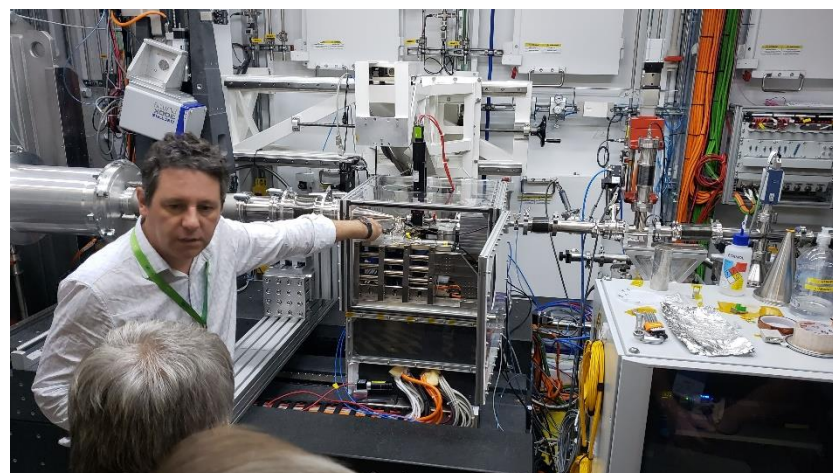
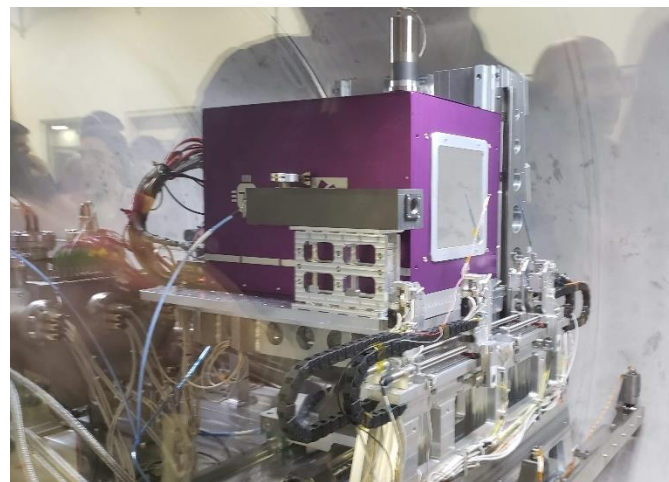
CATERETÊ beamline @ SIRIUS  
Campinas, Brazil

Dedicated SAXS beamline, new for 2022

- 4<sup>th</sup> gen synchrotron, 88 m source to sample
- Modern large area detector with beamstop
- 0 - 28 m sample to detector distance
- All components in vacuum, no windows from source to detector

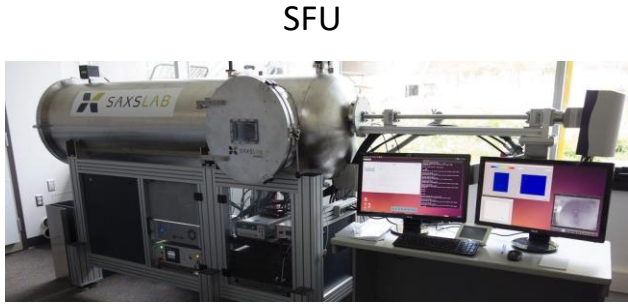


P. R. Willmott, 2022



Canadian Light Source  
Centre canadien de rayonnement synchrotron

# SAXS user facilities in Canada



SFU

SAXSLAB, Ganesha 300XL+

<https://www.4dlabs.ca/our-capabilities/equipment/small-angle-x-ray-scattering-system.html>



McGill

Anton Paar, SAXSpoint 2.0

<https://www.mcgill.ca/mc2/instrumentation/x-ray-diffraction/saxs>

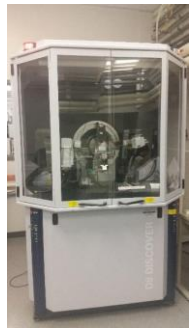


U. Montreal

SAXSLAB, Ganesha

[http://airen.bcm.umontreal.ca/biostruct/scheduling\\_saxs/](http://airen.bcm.umontreal.ca/biostruct/scheduling_saxs/)

U. Alberta



Bruker D8 DISCOVER Plus

<https://www.nanofab.ualberta.ca/2023/news/saxs-gi-saxs-waxs-are-available-on-bruker-d8d-plus/>

U. Toronto

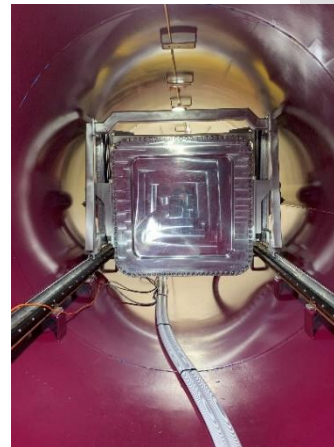


Anton Paar, SAXSpace

<https://lab.research.sickkids.ca/sbc-facility/saxs/>

McMaster

MacSANS - Commissioning begins mid-July 2024



MacSANS Overview	
Source-to-Sample	14 m
Sample-to-Detector	2 – 10 m continuous
Peak Thermal Flux on Sample	$10^4 - 10^5$ neutrons/cm <sup>2</sup> /s
Neutron Wavelength	2.6 - 23 Å at $\Delta\lambda/\lambda \approx 17\%$
Detector	100 cm <sup>2</sup> at 0.9 cm
Accessible Q-range	$0.005 < Q < 1.25 \text{ \AA}^{-1}$
Structure Scale	0.6 – 125 nm

<https://nuclear.mcmaster.ca/neutron-beams/>

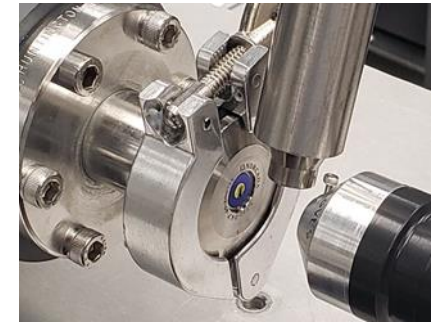
# CLS SAXS instrument

1) Beam defining slit  
+6.1 m  
(1.00 mm x 1.00 mm)

2) Anti-scatter slit  
+0.9 m  
(0.25 mm V x 0.50 mm H)

200 nm SiN window      3) Guard slit (0.80 mm)

~9 m long



Small air gap for sample (~35 mm)

3 pinhole layout

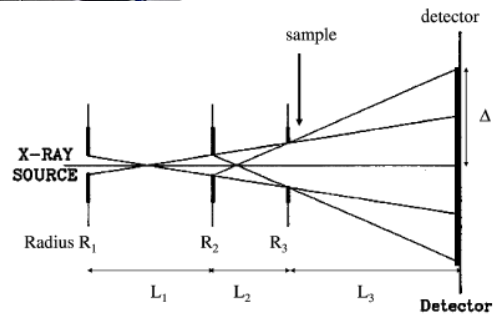
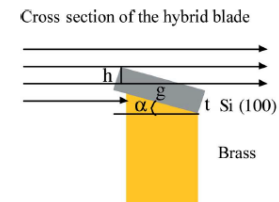
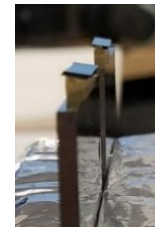
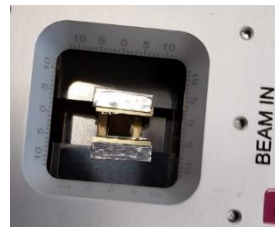


Figure 4  
Schematic layout of a SAXS pinhole camera.

J. Appl. Cryst. (2004). 37, 369-380

low parasitic scatter



Y. Li et al., J. Appl. Cryst. (2008). 41, 1134-1139

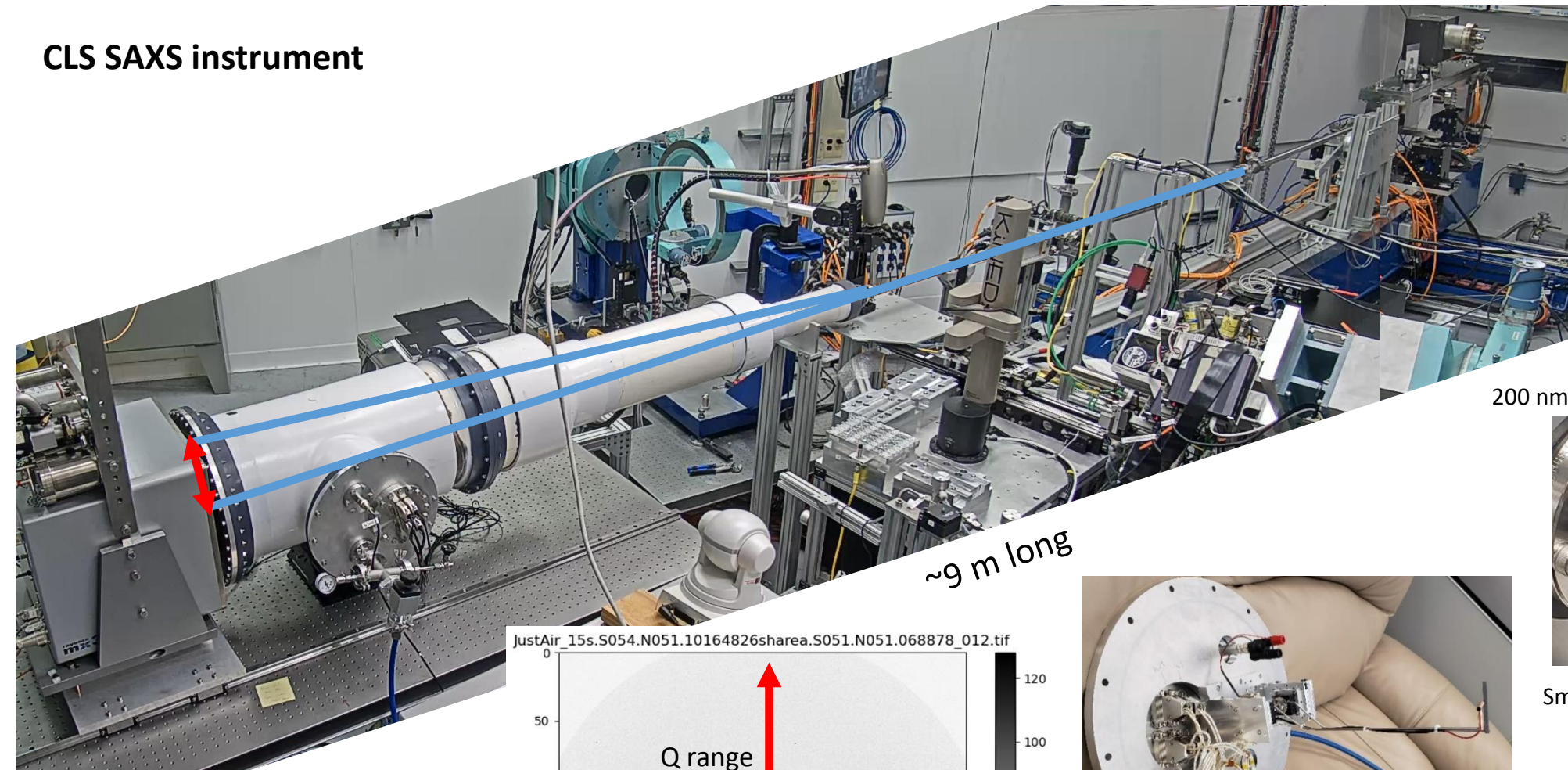
N.M. Kirby et al., J. Appl. Cryst. 46, 1670-1680 (2013)



Canadian Light Source  
Centre canadien de rayonnement synchrotron

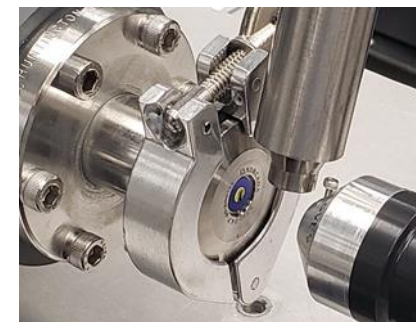


# CLS SAXS instrument



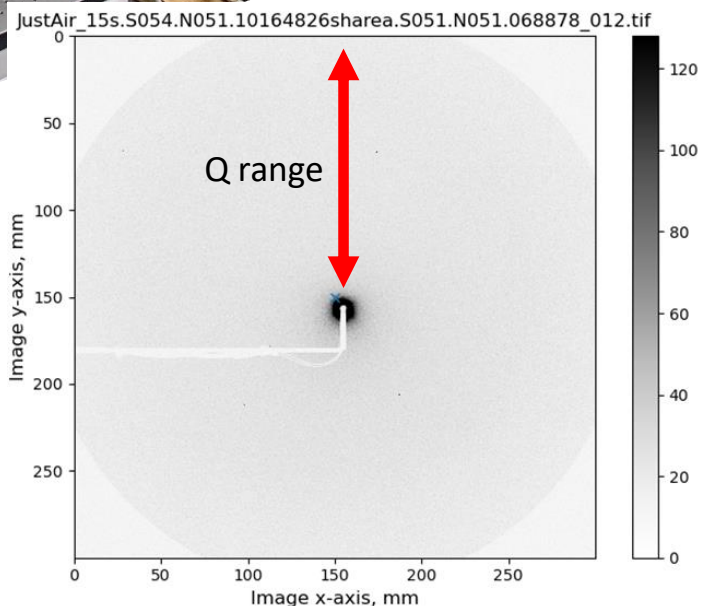
~9 m long

200 nm SiN window      3) Guard slit (0.80 mm)



Small air gap for sample (~35 mm)

$\lambda = 1.26 \text{ \AA} \text{ (9.85 keV)}$   
 $0.658 \text{ \AA} \text{ (18.8 keV)}$   
 $Q \sim 0.005 - 0.5 \text{ \AA}^{-1}$   
 $d \sim 125 - 1.25 \text{ nm}$   
 $2\theta \sim 0.05 - 3.7^\circ \text{ (see website)}$



Vacuum tube ( $\leq 10^{-2}$  Torr)  
 Beamstop with photodiode  
 -2.0 m  
 (4.0 mm)



13.5" Kapton window (125  $\mu\text{m}$ )  
 ~2100 pounds

# CLS SAXS instrument

New for next cycle (Jan-June 2025)

New table

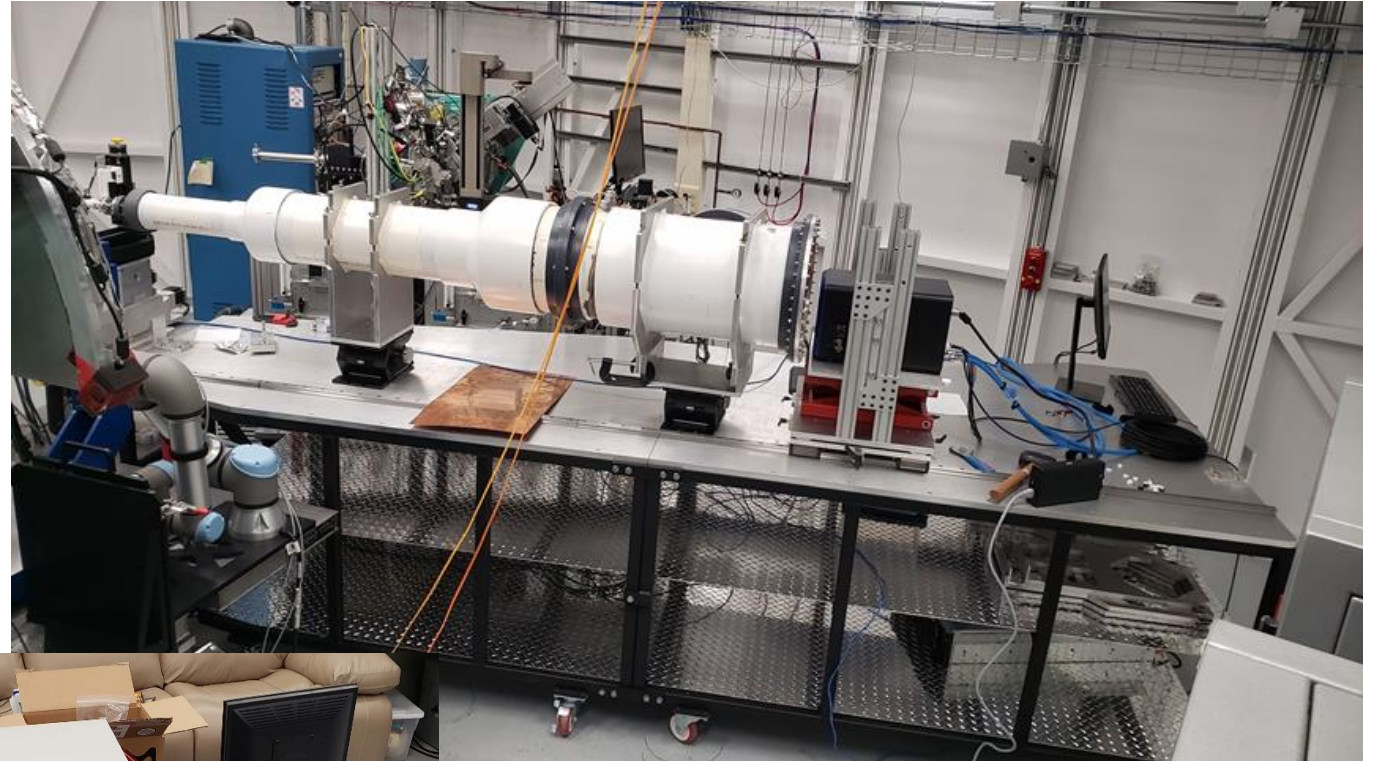
- Easily go between SAXS and WAXS
- $q_{\min}$  from  $0.005 \text{ \AA}^{-1}$  to  $0.003 \text{ \AA}^{-1}$

Optional Eiger 2 detector

- Strongly scattering samples
- Dynamic processes (sub-second)

HPLC system

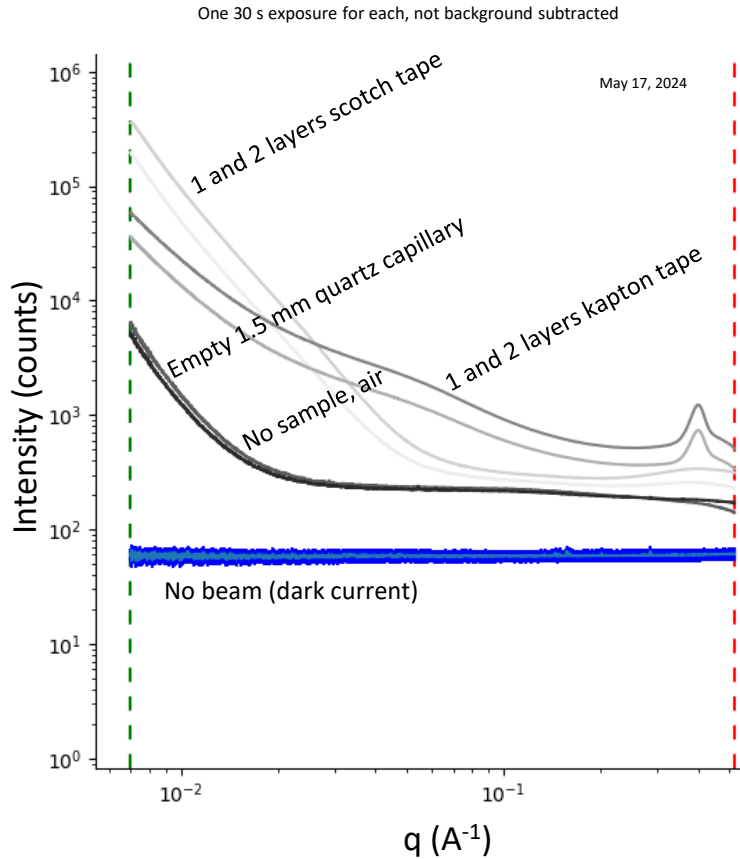
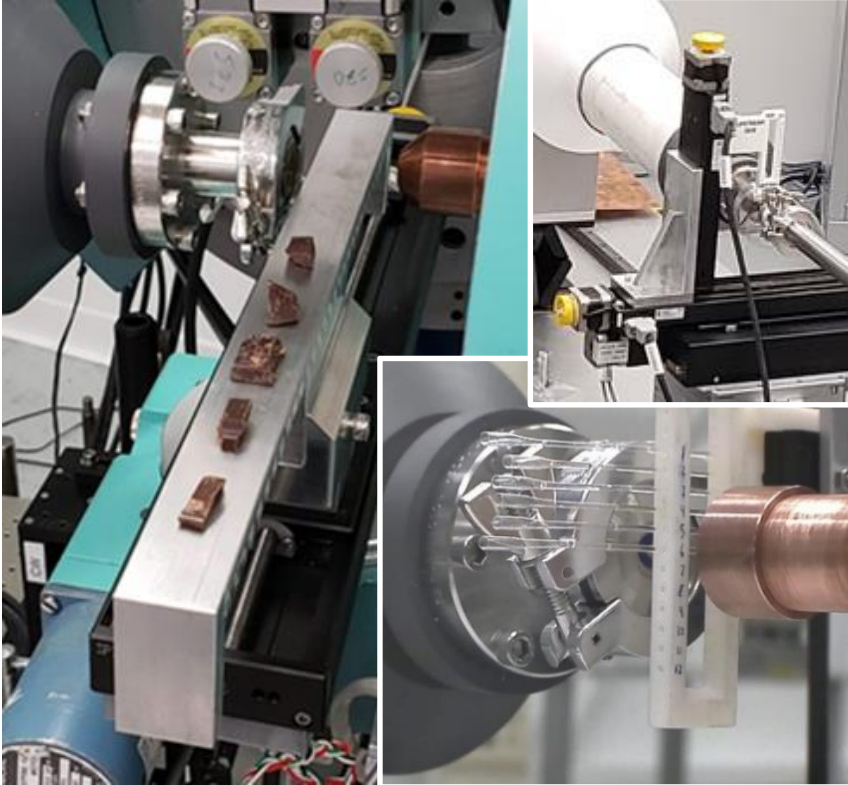
- Size exclusion chromatography-coupled SAXS



# Sample preparation and mounting

## Transmission

Freestanding, quartz capillaries, or on tape



## Grazing incidence

Thin *flat* films on Si wafers, or glass slides  
Min  $\theta$  step =  $0.01^\circ$



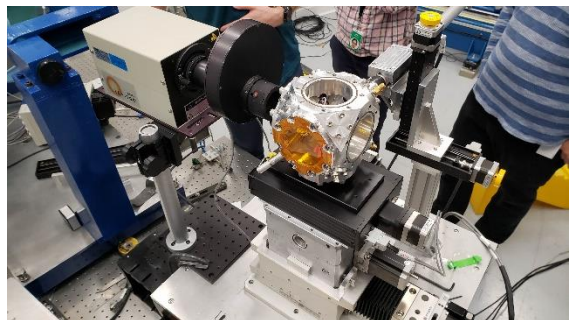
Calculate the transmission of your sample...  $1/e$  is ideal  
Bring blanks, Bring a range of concentrations  
Bring a sample where you know what to expect  
**MUST** buy your own capillaries



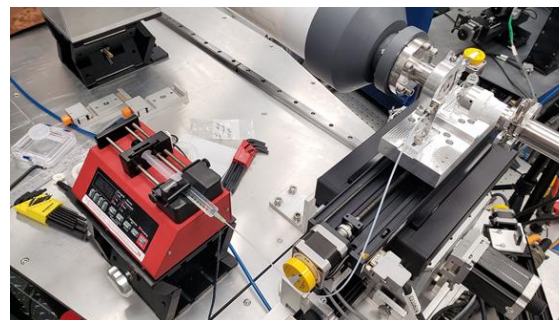
# Sample preparation and mounting

We are flexible!

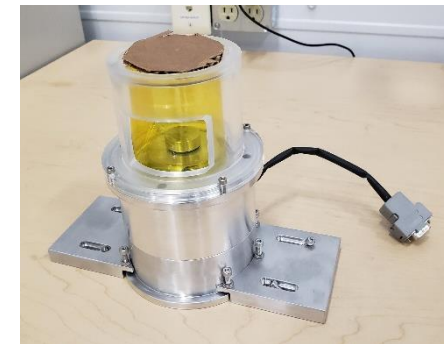
Operando solar cells  
(sunlight, humidity, temperature)



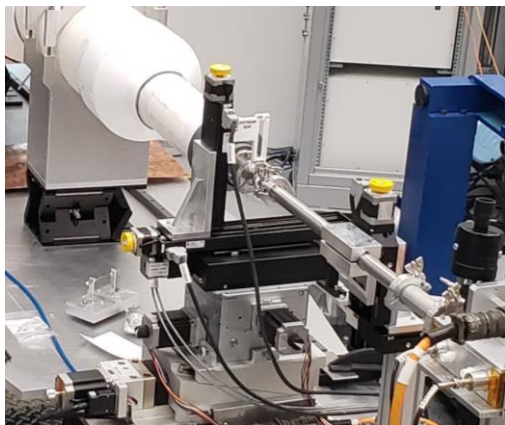
Liquid flow cell and syringe pump



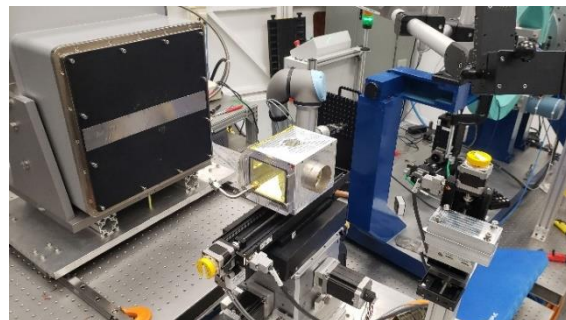
In-situ spin coater  
(soon with heating)



Multi-sample transmission



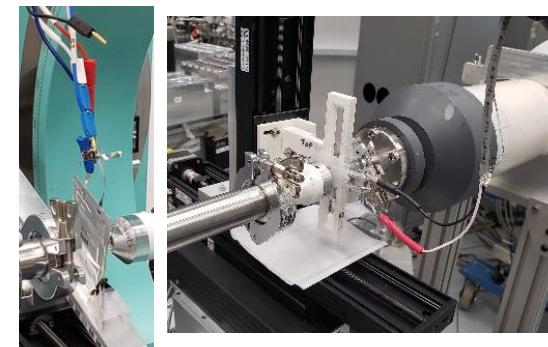
Helium box with Linkam heating and cooling  
(-193 - +350°C)



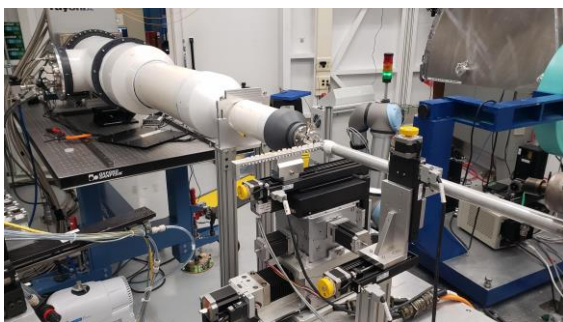
Oxford cryostream (-193 - +226°C)



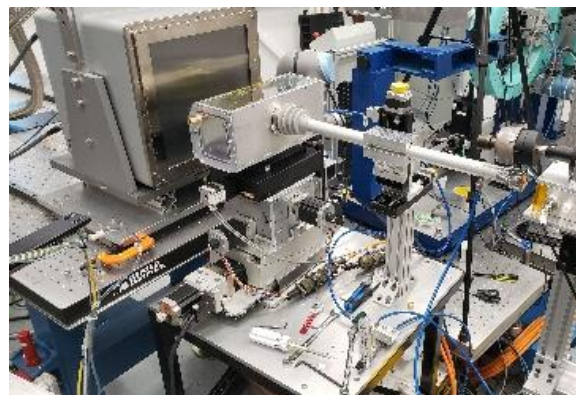
Electric field/Electrochem SAXS



Automated multi-sample grazing



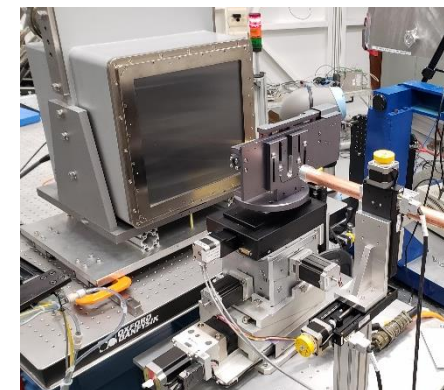
Automated multisample Helium box  
for GIWAXS, also with mild heating



1000°C furnace with gas flow

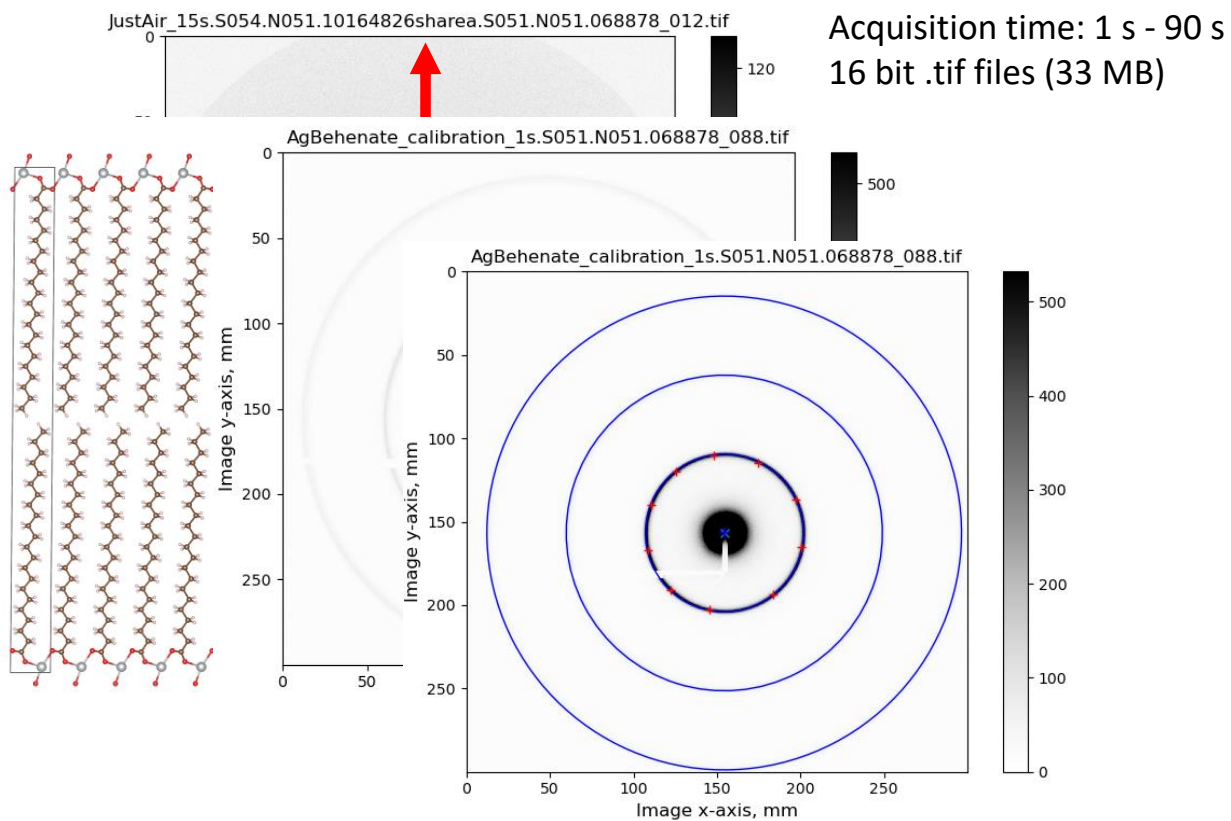


Stretching



# Detector acquisition and calibration

During your beamtime, note the wavelength, the approximate detector distance, and detector details



Acquisition time: 1 s - 90 s  
16 bit .tif files (33 MB)

## GSAS-II demo, "Calibration of an area detector"

GSAS-II project: <unnamed project>

File Data Calculate Import Export | Calibration Integration Parms | Help

Loaded Data:

- Notebook
- Controls
- Covariance
- Constraints
- Restrains
- Rigid bodies
- IMG JustAir\_15s.S054.N051.10164826sharea.S051.N051.068878\_012.tif
  - Comments
  - Image Controls
  - Masks
  - Stress/Strain
- IMG AgBehenate\_calibration\_1s.S051.N051.068878\_088.tif
  - Comments
  - Image Controls
  - Masks
  - Stress/Strain

Image Controls:

Type of image data: SASD - small angle scattering data | Color bar: Greys | Azimuth offset: 0.0

Max intensity: 532 | Min intensity: 0 | Auto scaler: ? | Show line scan:

Calibration coefficients:

- Beam center X: 154.481
- Beam center Y: 156.736
- Wavelength\*: 1.18178
- Distance: 2335.831
- Tilt angle\*: -0.873
- Tilt rotation\*: 242.48

Integration coefficients:

- Bin style: Constant step bins in log(q) | Pink beam source?:
- Inner/Outer Q: 0.464 | 4.494
- Start/End azimuth: 0.0 | 360.0
- No. 2-theta/azimuth bins: 2500 | 1
- Do full integration?
- Use for all new images?
- Azimuth at bin center?
- Apply sample absorption?
- Apply polarization? Value (0.001-0.999): 0.99

Dark image: [ ] multiplier: -1.0

Background image: [ ] multiplier: -1.0

Gain map: [ ]

Calibration controls:

Calibrant: Ag behenate | Calib lines to skin: [ ] | Min calib denoising: 16.0

Min ring I/lb: 1.0

Calibration Integration **Parms** | Help

Controls:

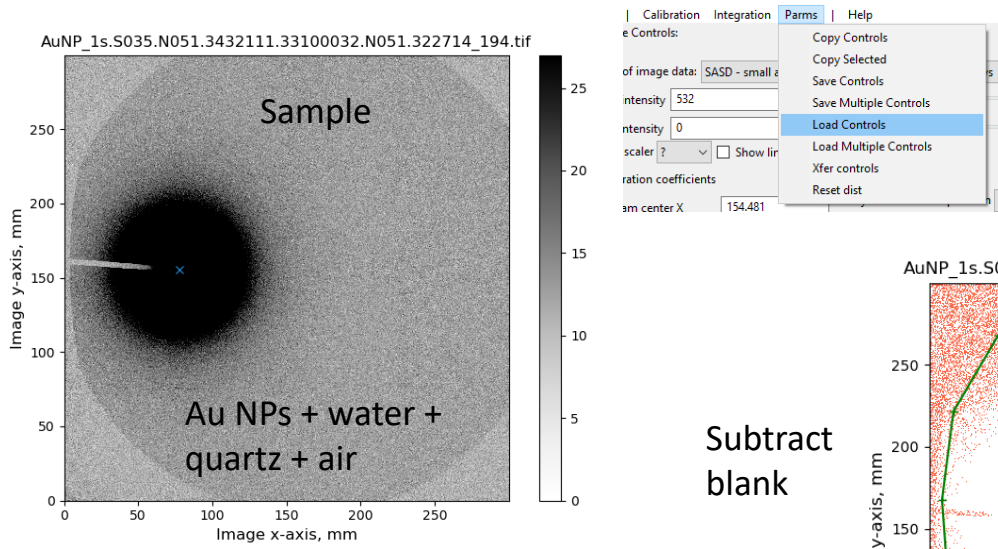
- Copy Controls
- Copy Selected
- Save Controls
- Save Multiple Controls

Image data: SASD - small angle scattering data | Max intensity: 532

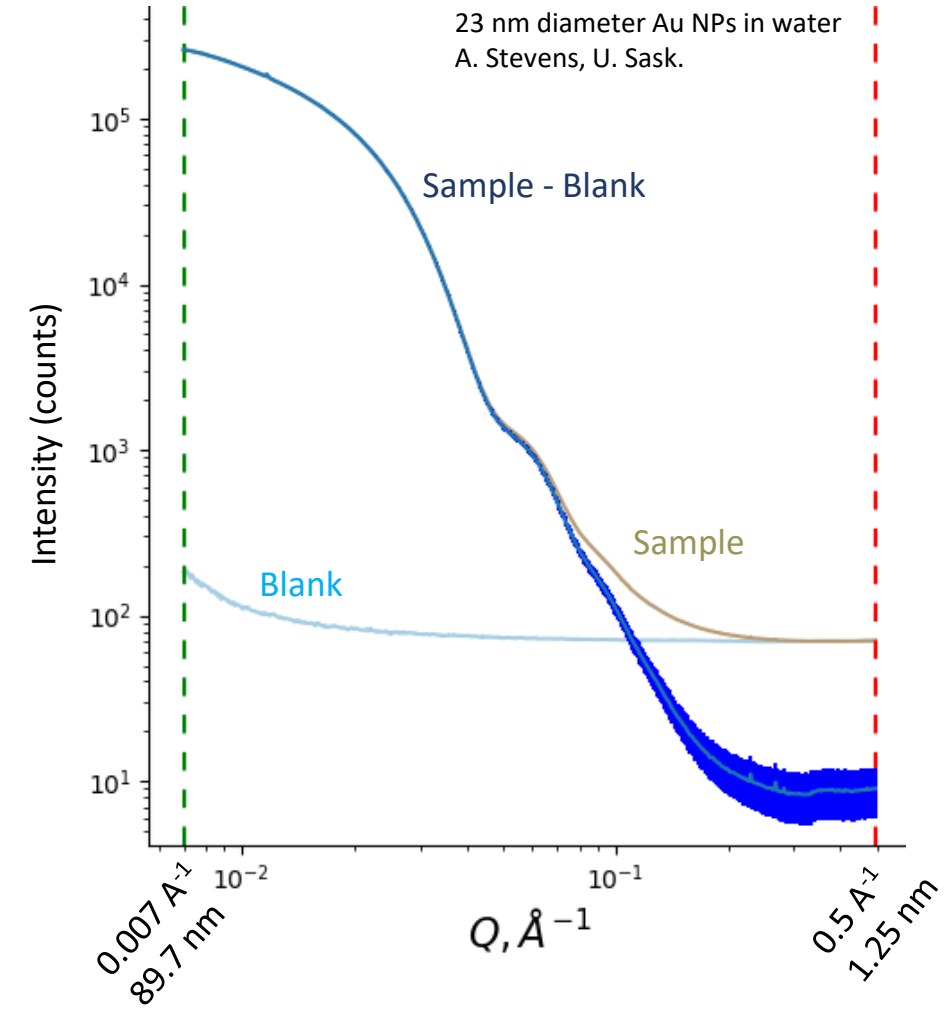
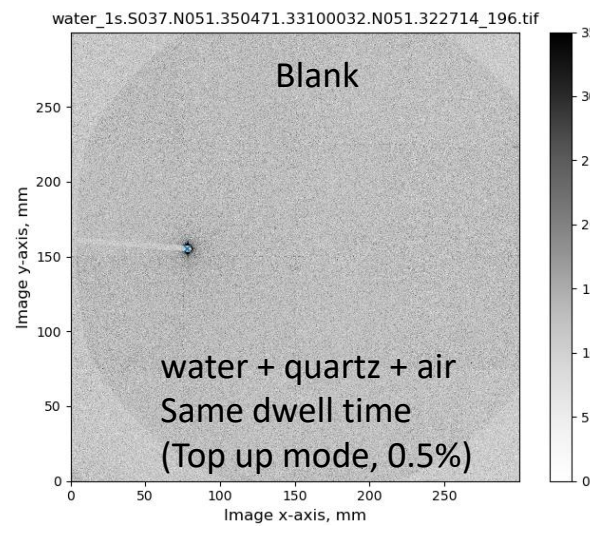
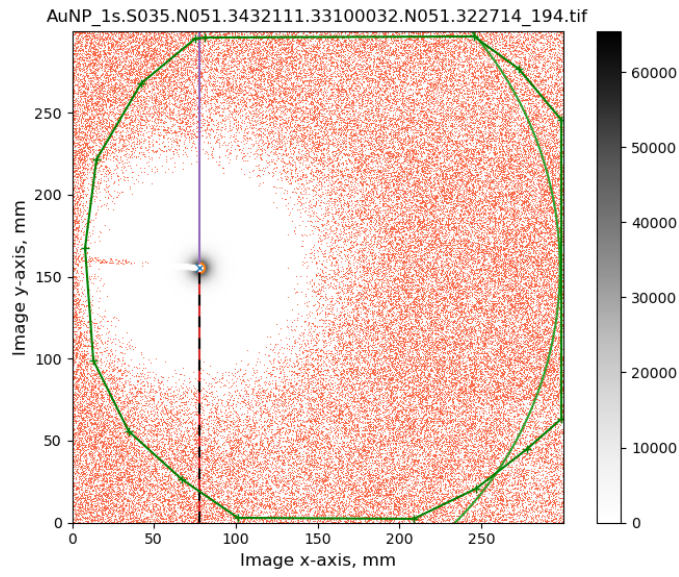
Ring	q (Å <sup>-1</sup> )	d (nm)
1	0.1076	5.839
2	0.2152	2.920
3	0.3228	1.946
4	0.4304	1.460
5	0.5380	1.168



# Data correction, and reduction to 1D plot



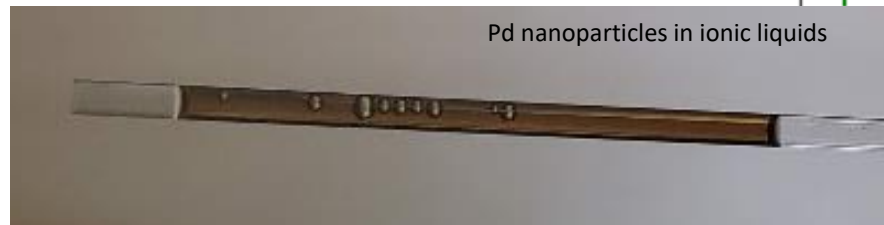
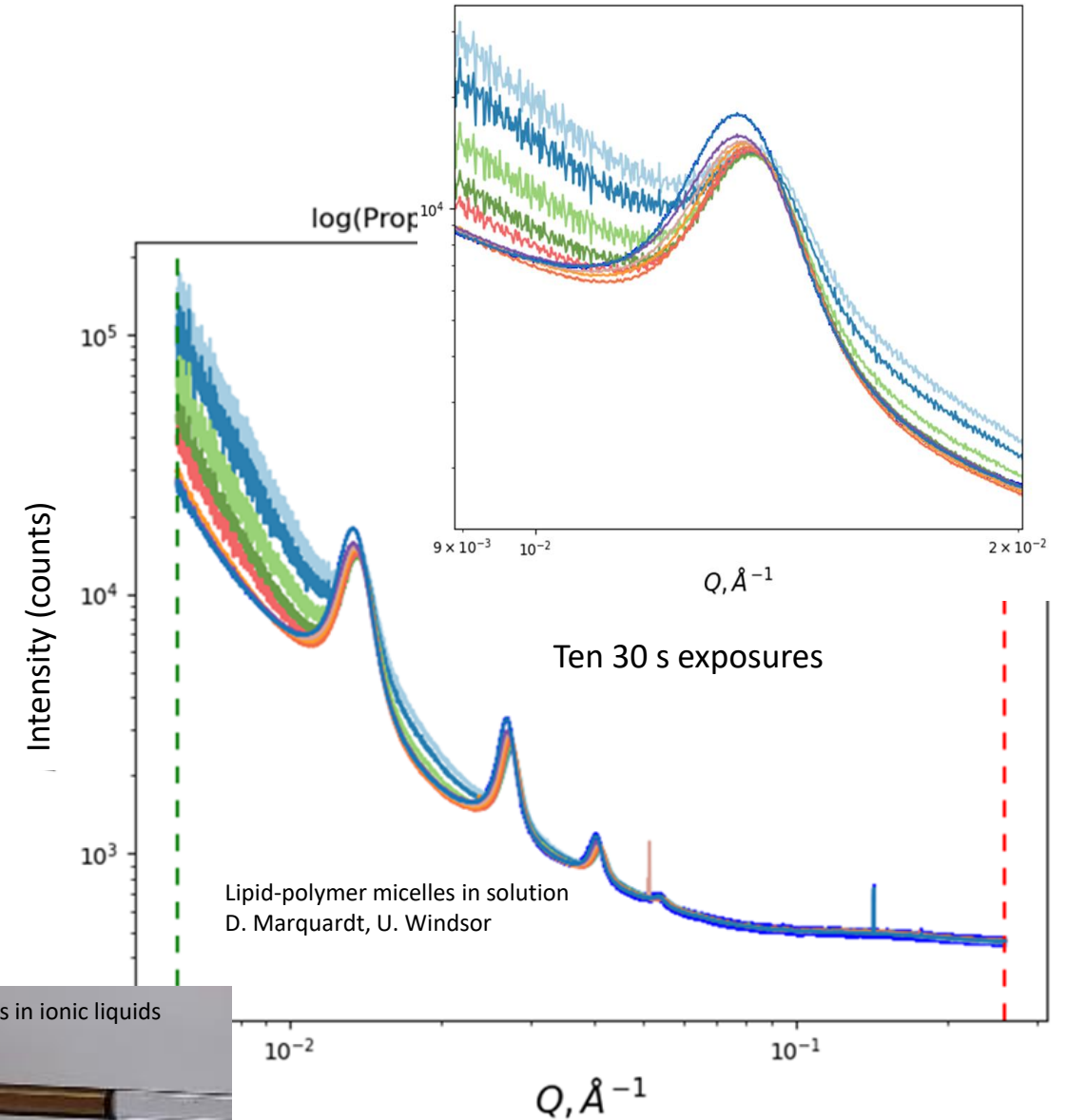
Subtract blank



Proper units of intensity are  $\text{cm}^{-1}$   
IF data are scaled to absolute intensity

## 5 tips for a great data collection experience

- 1) Know what  $q$  range you want, and include it in your proposal. We can tune the endstation to your problem.
- 2) Know what your data should look like. Find published examples, and bring a known good sample.
- 3) Get in contact  $\sim 2$  weeks in advance.
- 4) Choose quality over quantity: Work up data as you go along, check for radiation damage at the start, many blank measurements.
- 5) It's your experiment.



# Data analysis with examples





# Origins of the SAXS signal

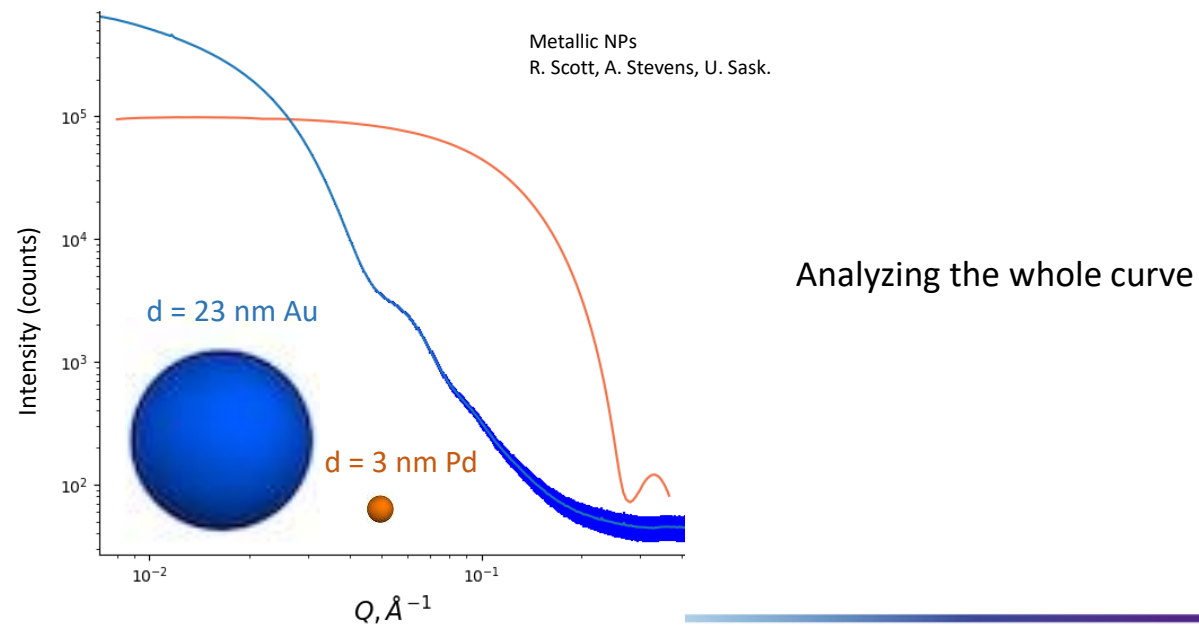
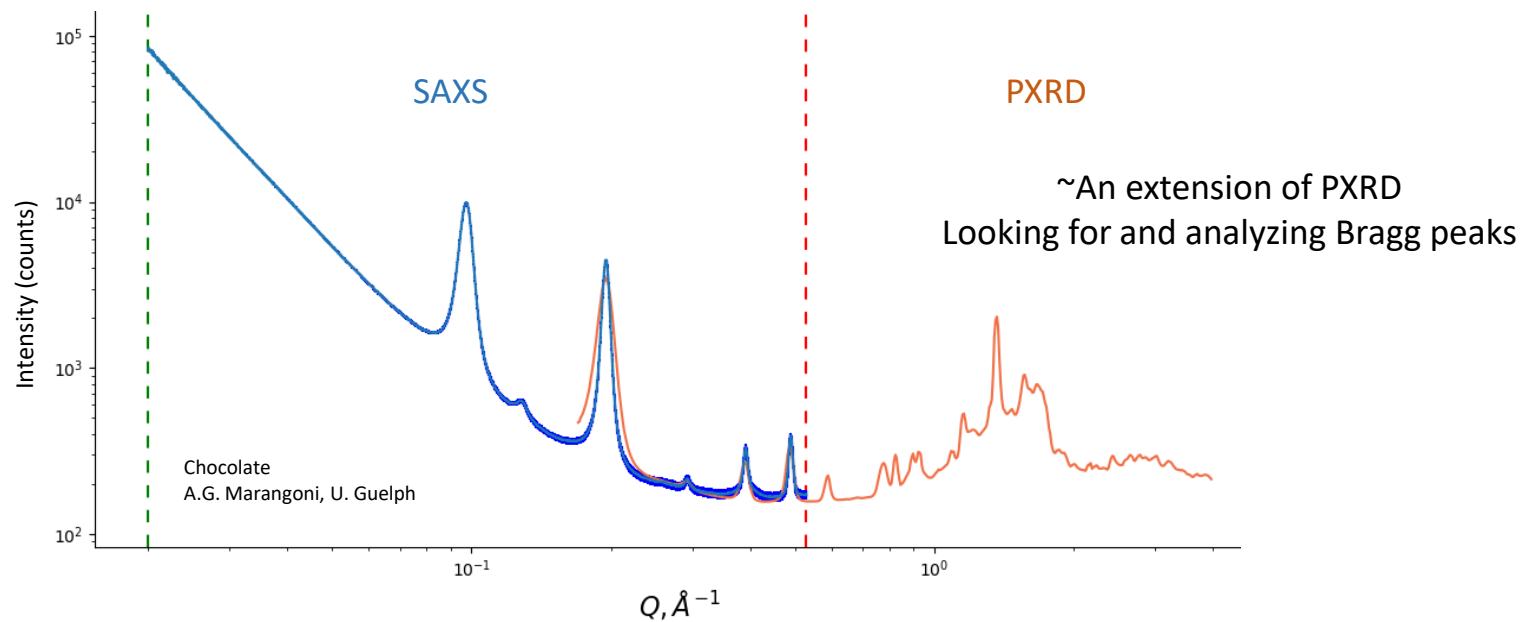
$$I(Q) = S(Q) \times P(Q)$$

**S(Q)** is the **structure** factor

- *Inter*-particle interferences
- High concentrations (>5% vol.)
- **Ordering/packing of particles**

**P(Q)** is the **form** factor

- *Intra*-particle interferences
- Low concentration (dilute limit)
- **Size/polydispersity/shape of particles**



# Phospholipid Self-Assembly in Cocoa Butter Provides a Crystallizing Surface for Seeding the Form V Polymorph in Chocolate

J.A. Stobbs, Alejandro G. Marangoni et al., *Cryst. Growth Des.* **24**, 7, 2685–2699 (2024)

Marangoni group  
Dept. Food Science  
University of Guelph

The nanostructure of chocolate is manipulated using time and energy-intensive “tempering”

Guides crystallization of cocoa butter to “polymorph V”

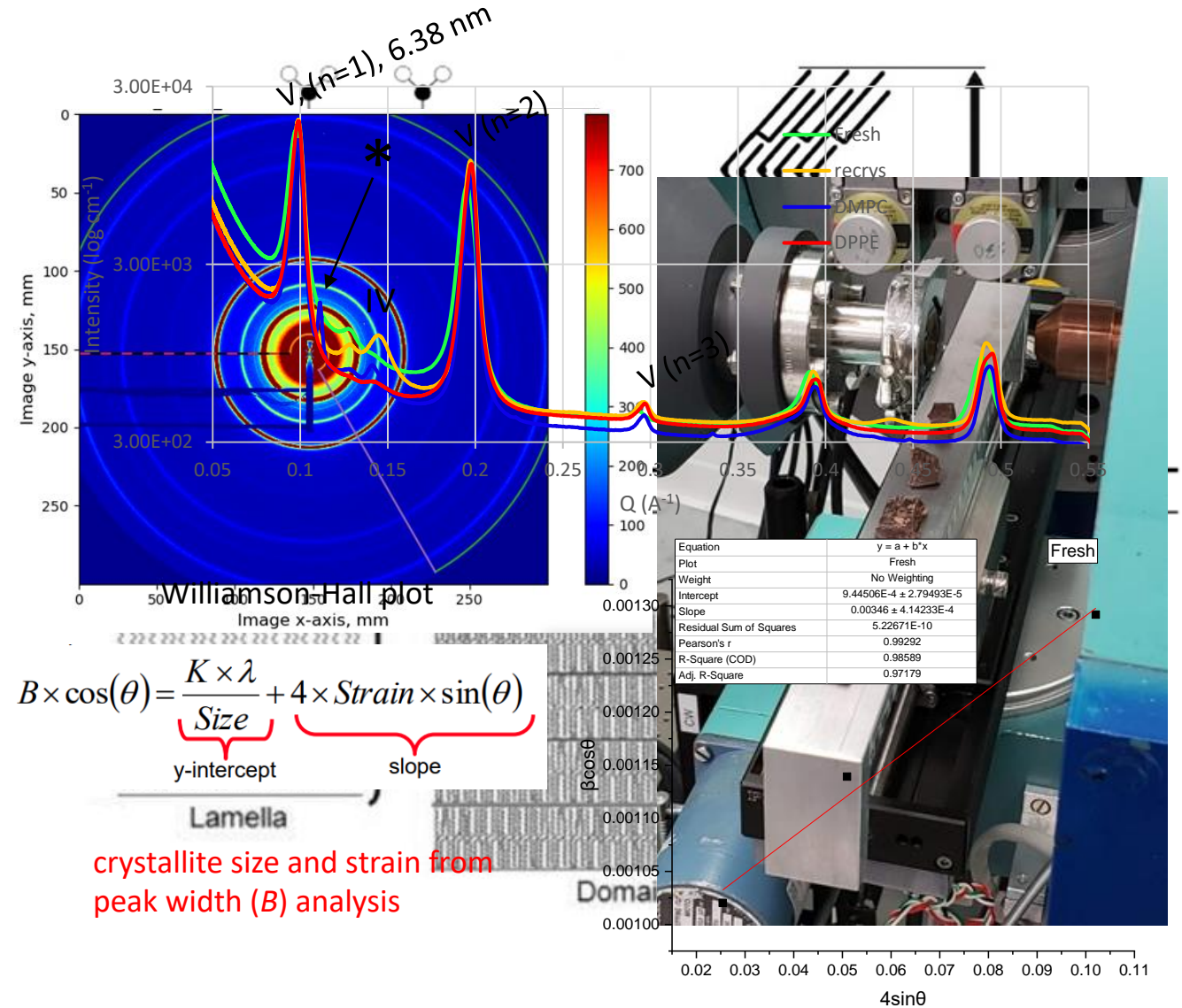


Tempering of cocoa butter and chocolate using minor lipidic components

Jay Chen<sup>1</sup>, Saeed M. Ghazani<sup>1</sup>, Jarvis A. Stobbs<sup>1,2</sup> & Alejandro G. Marangoni<sup>1,2\*</sup>

NATURE COMMUNICATIONS | (2021)12:5018

Why does it work?



crystallite size and strain from peak width (B) analysis

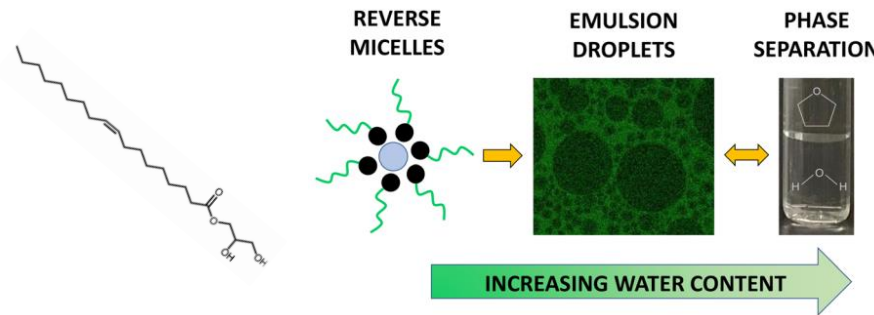


Canadian Light Source  
Centre canadien de rayonnement synchrotron

# Environmental science: Waste water treatment

*Scientific Reports* **12**, 15832 (2022)  
*J. Molecular Liquids* **367**, 120551 (2022)  
*Physics of Fluids* **34**, 097119 (2022)

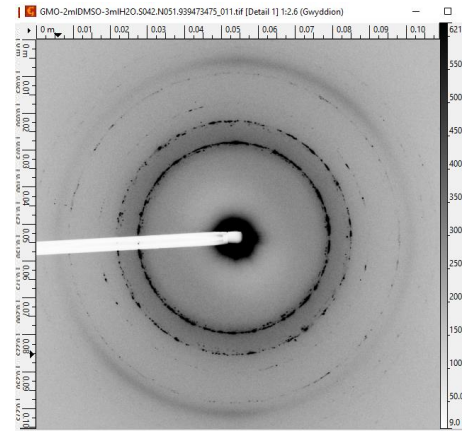
Separating solvents (THF, DMSO, DMF and acetonitrile) and metals from waste water using amphiphiles, emulsifiers and surfactants



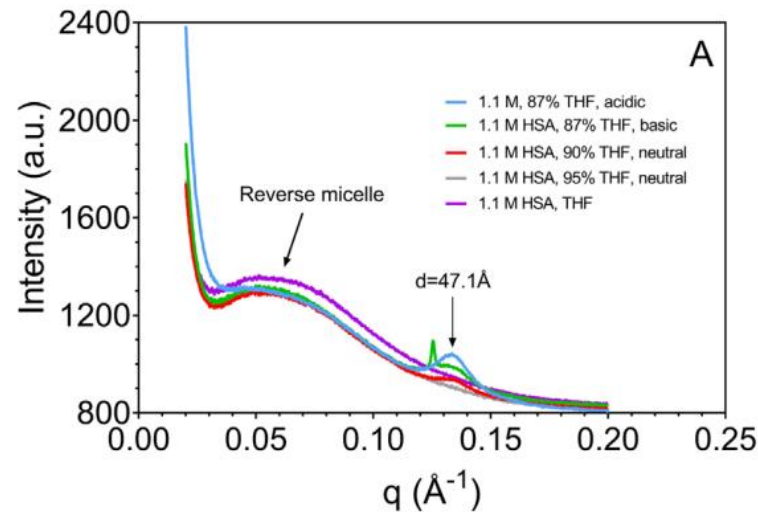
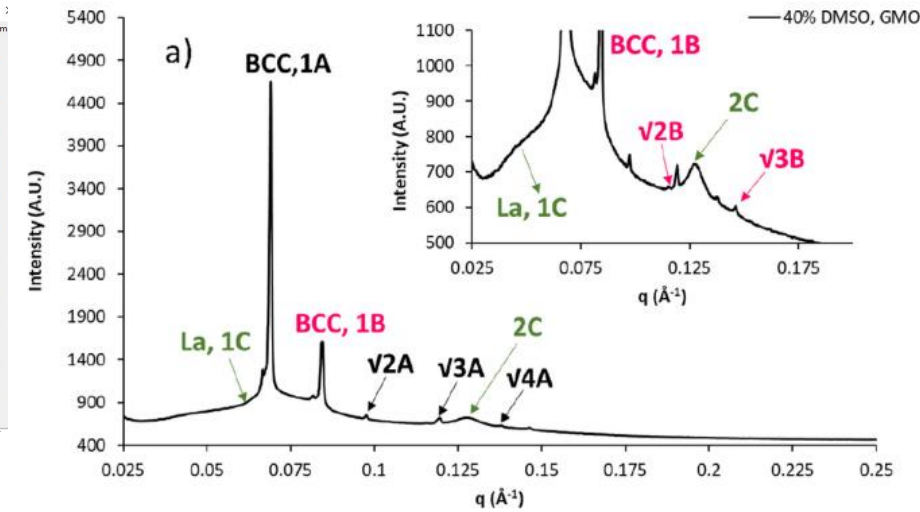
Mechanism of solvent separation?

1.1 M HSA, 87% THF, acidic pH				
		$\xi$ (Å)	d (Å)	d/x
$a_2$	1.158	19.5	86.0	4.4
$c_1$	-98.92			
$c_2$	18.245			

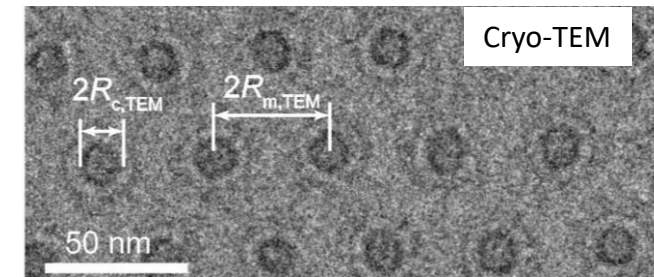
Powder diffraction ... ??



Symmetry	Peak position ratio
BCC (Im3m)	v2, v3, v4 ...



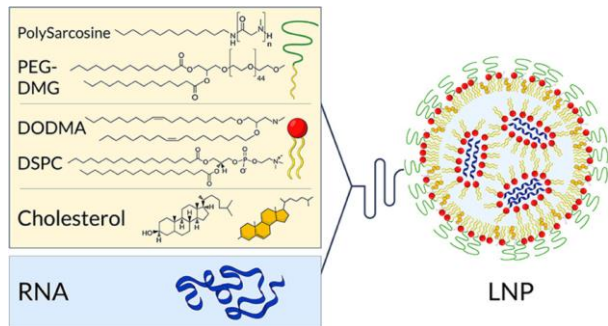
Reverse micelle lattice



L. Chen et al., *PNAS* **115**, 7218-7223 (2018)

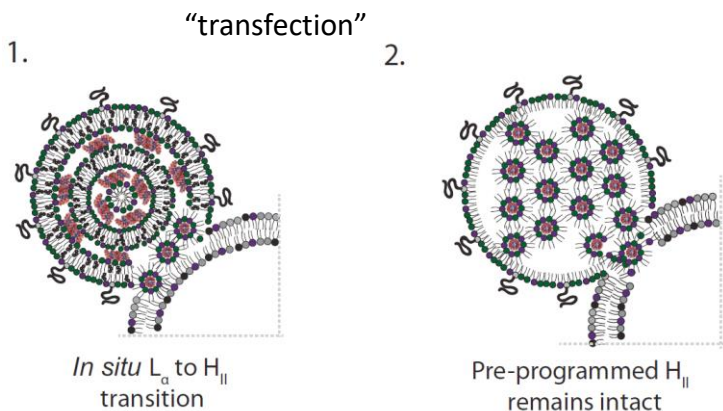
# Pharmacy: Liposomes for RNA delivery

Pure mRNA degraded by enzymes in tissues  
Hide it inside a liposome NP to get it into cells



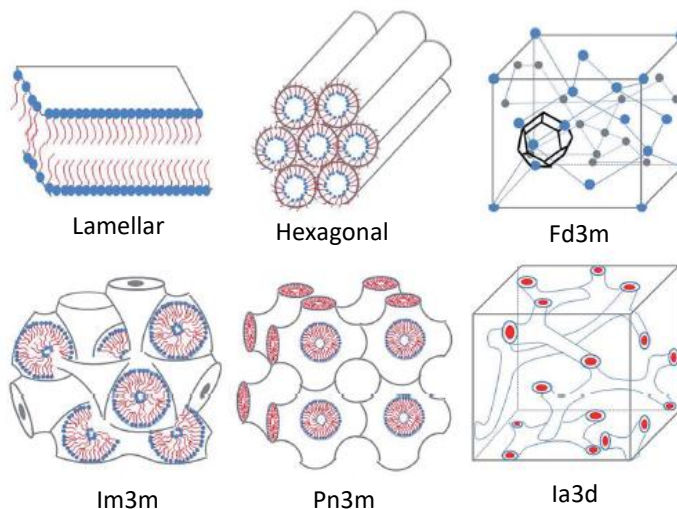
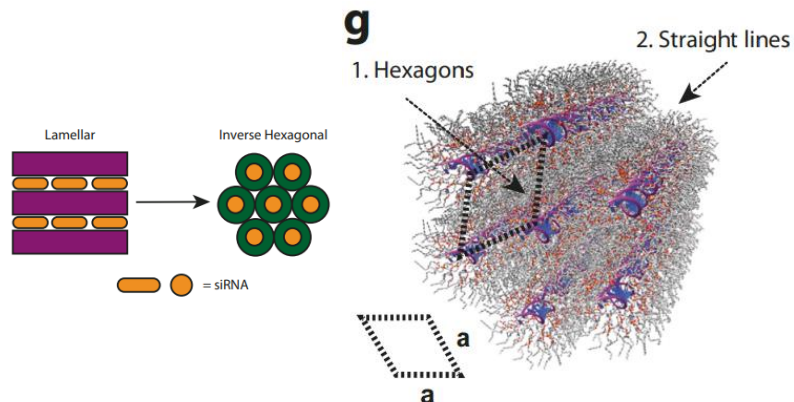
ACS Appl. Nano Mater. 2020, 3, 11, 10634–10645

Pfizer, Moderna COVID-19 mRNA vaccines



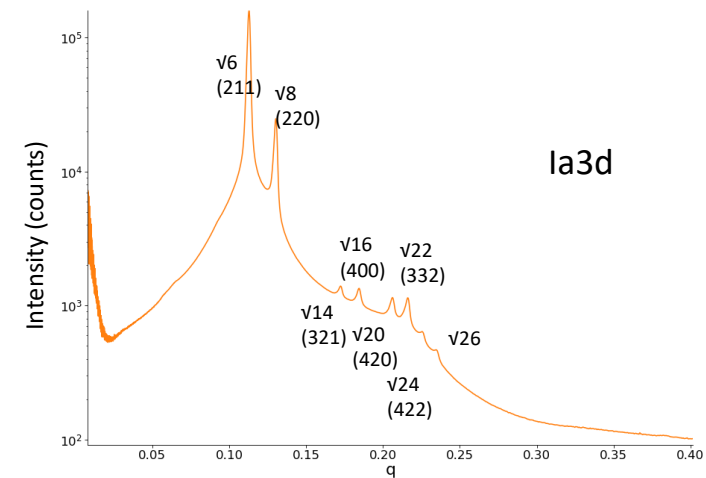
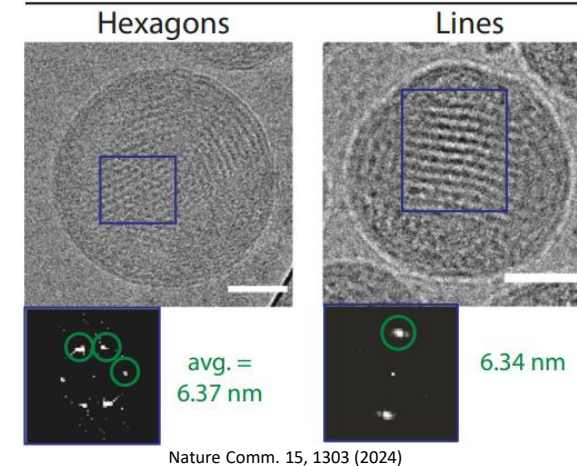
Nature Comm. 15, 1303 (2024)

## Lipid phase affects transfection efficiency



Y. Huang, S. Gui, RSC Adv., 2018, 8, 6978–6987

## Cryo-TEM

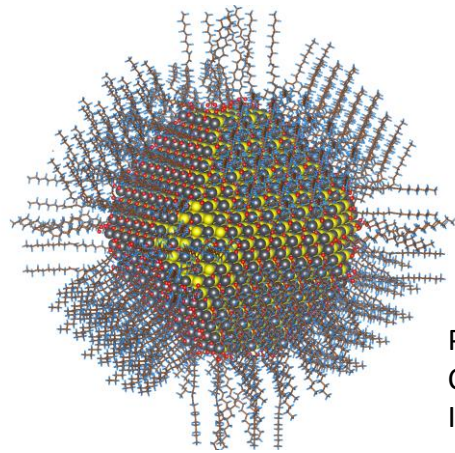


Types of LLC phase	SAXS
Lamellar phase	1 : 2 : 3 : 4
<i>Im3m</i>	$\sqrt{2} : \sqrt{4} : \sqrt{6} : \sqrt{8}$
<i>Pn3m</i>	$\sqrt{2} : \sqrt{3} : \sqrt{4} : \sqrt{6} : \sqrt{8} : \sqrt{9}$
<i>Ia3d</i>	$\sqrt{6} : \sqrt{8} : \sqrt{14} : \sqrt{16} : \sqrt{20} : \sqrt{22}$
<i>Fd3m</i>	$\sqrt{3} : \sqrt{8} : \sqrt{11} : \sqrt{16} : \sqrt{19}$
Reverse hexagonal	1 : $\sqrt{3}$ : 2 : $\sqrt{7}$



Canadian Light Source  
Centre canadien de rayonnement synchrotron

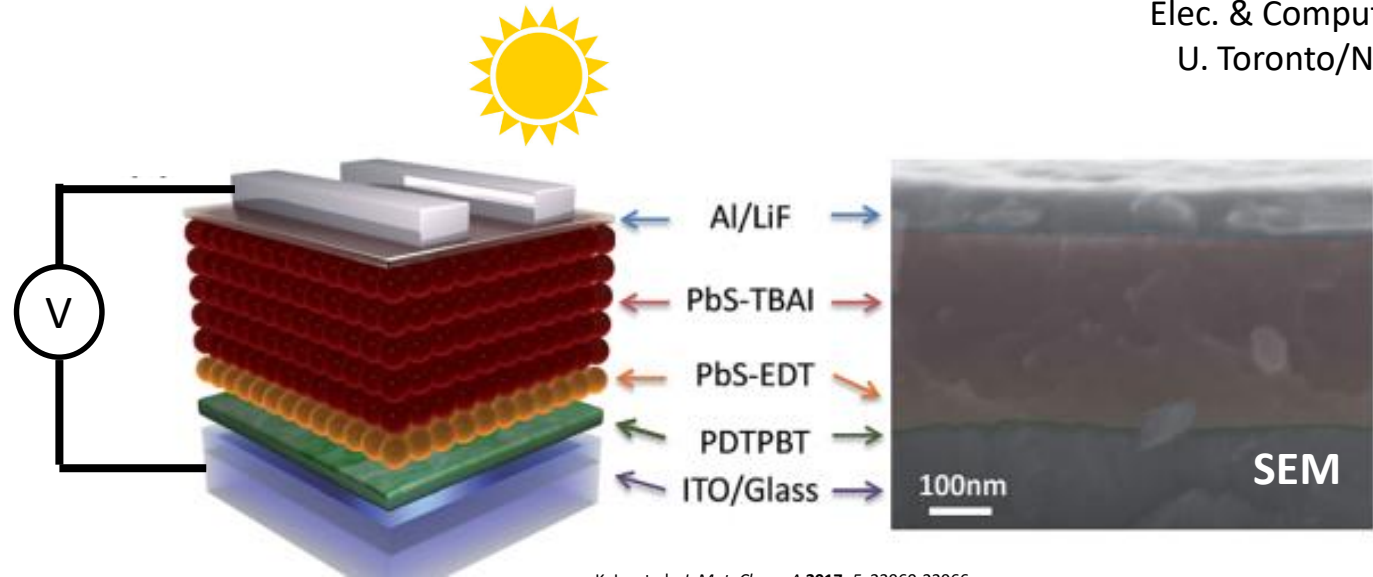
# Energy materials: Quantum dot solar cells



PbS  
CsPb(Br/I)<sub>3</sub>  
InAs

<https://www.olcf.ornl.gov/2015/05/05/demystifying-quantum-dot-conundrums/>

Efficient exciton generation  
Size-based, tune-able band gap

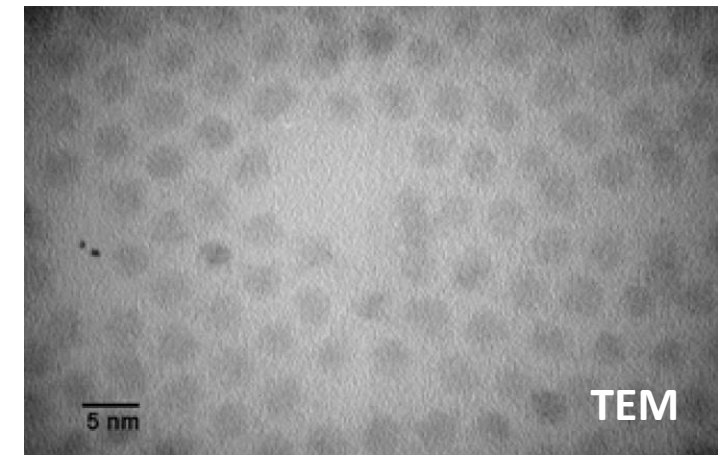


K. Lu et al., *J. Mat. Chem. A* 2017, 5, 23960-23966

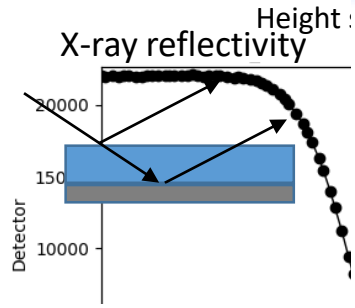
QD films deposited layer-by-layer using spin coating  
Top electrodes deposited using thermal evaporation

- 18.1% record efficiency in 2022  
Performance improved by tuning:
- Particle size distribution or polydispersity
  - How particles pack and the interparticle distance

- Control the size and packing by:
- Ligand exchanges cycles, centrifuge to remove largest QDs
  - Rinsing steps during buildup
  - Spin speeds, solvents, additives

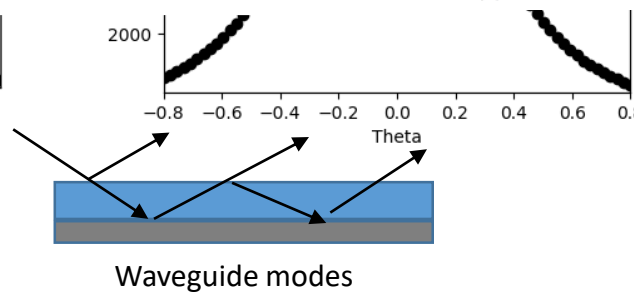
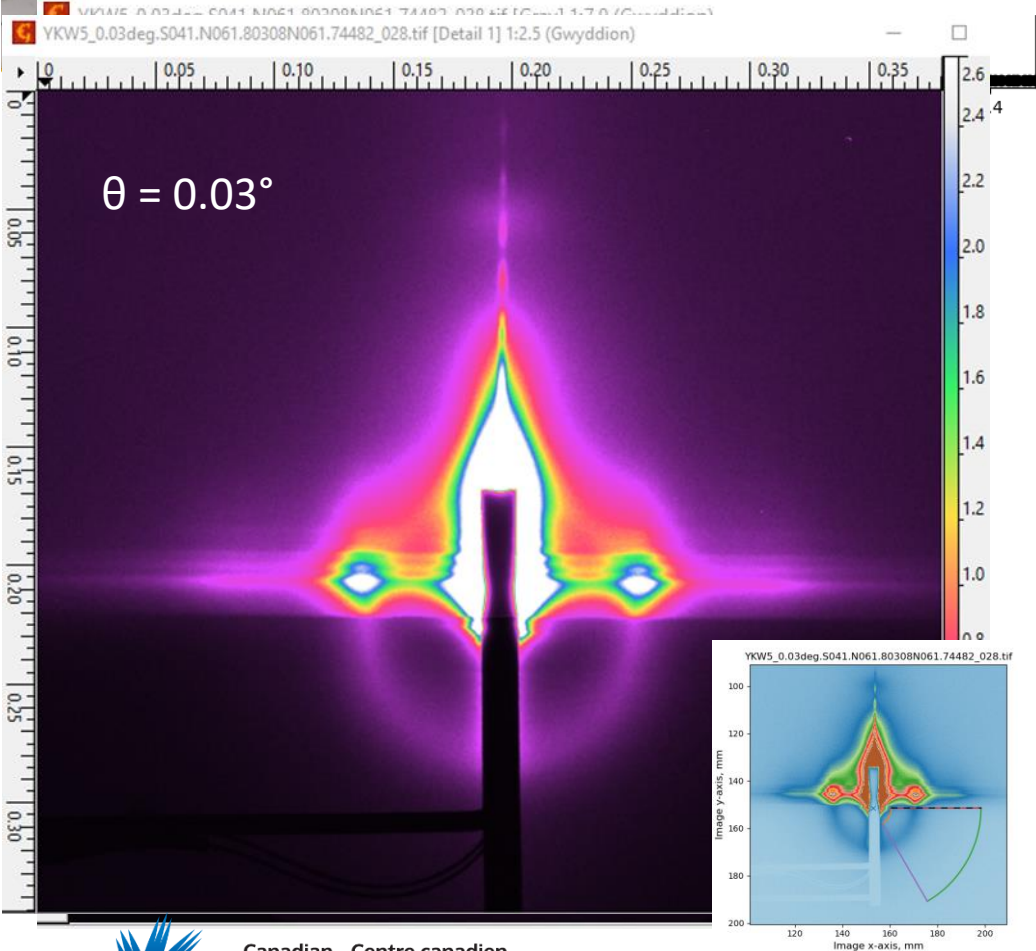
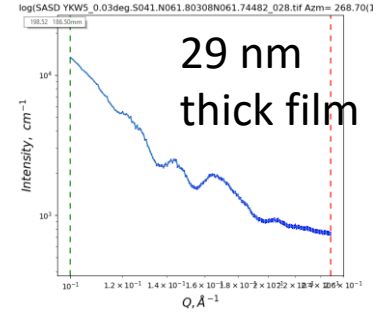
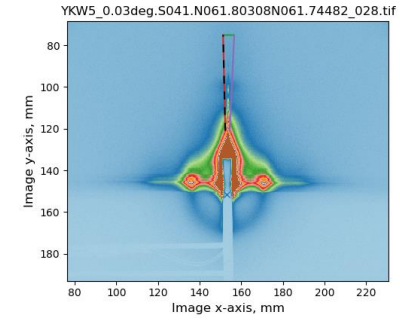
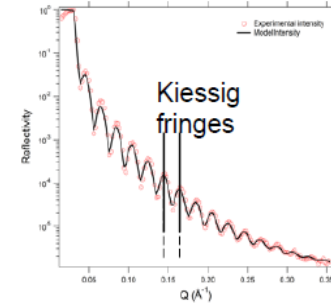


M. Yuan et al., *Adv. Mater.* 2014, 26, 3513-3519



### Simple estimation of film thickness

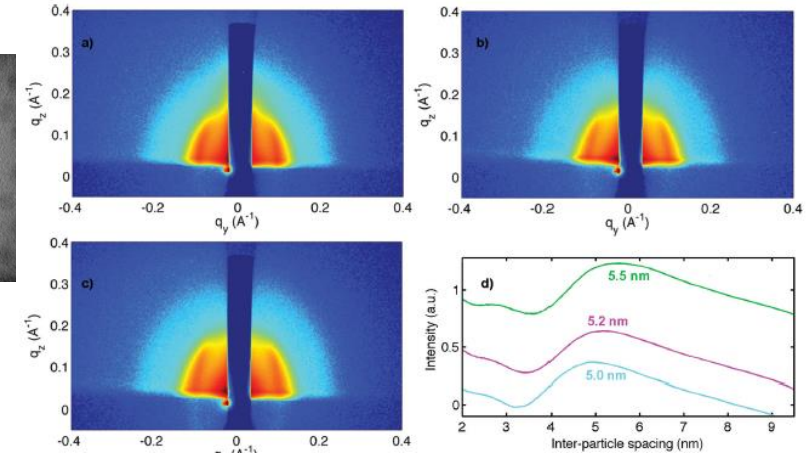
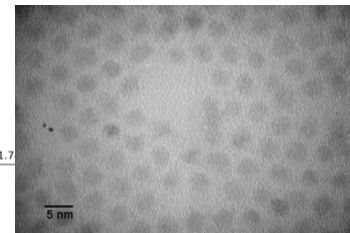
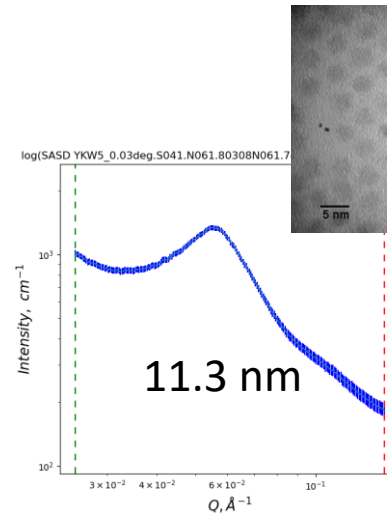
Fringes with uniform spacing  
Thickness of the layer :

$$t = \frac{2\pi}{\Delta q_z}$$


- Indicates very flat, high quality film
- Adjusting grazing angle can give depth information

- Reflectivity gives film thickness

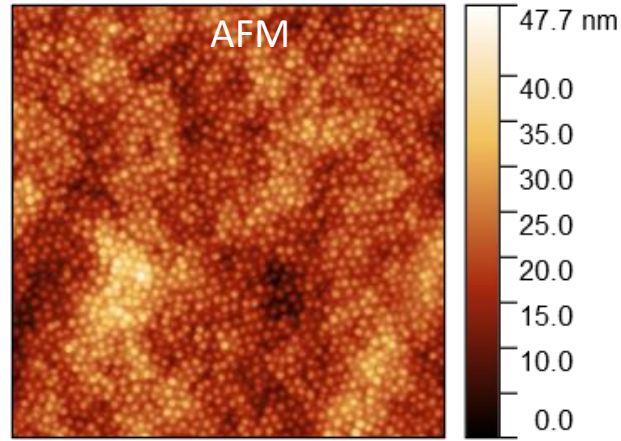
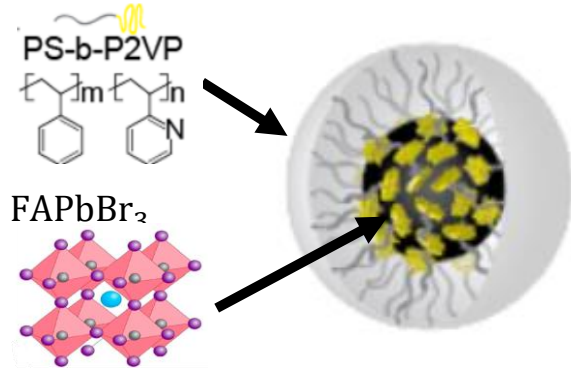
- Position gives interparticle distance
- Peak width gives size distribution
- Average over relatively huge area



M. Yuan et al., Adv. Mater. 2014, 26, 3513–3519

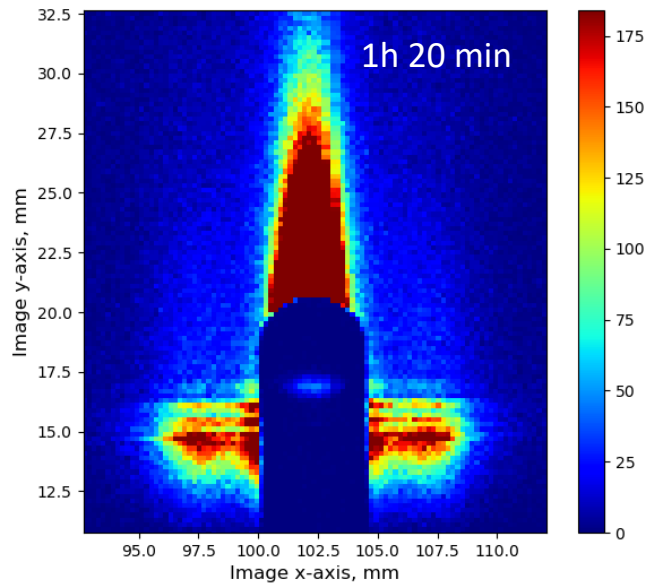
# GISAXS - Lab X-ray source vs. Synchrotron

TURAK FUNCTIONAL NANOMATERIALS RESEARCH GROUP  
ayse.turak@concordia.ca

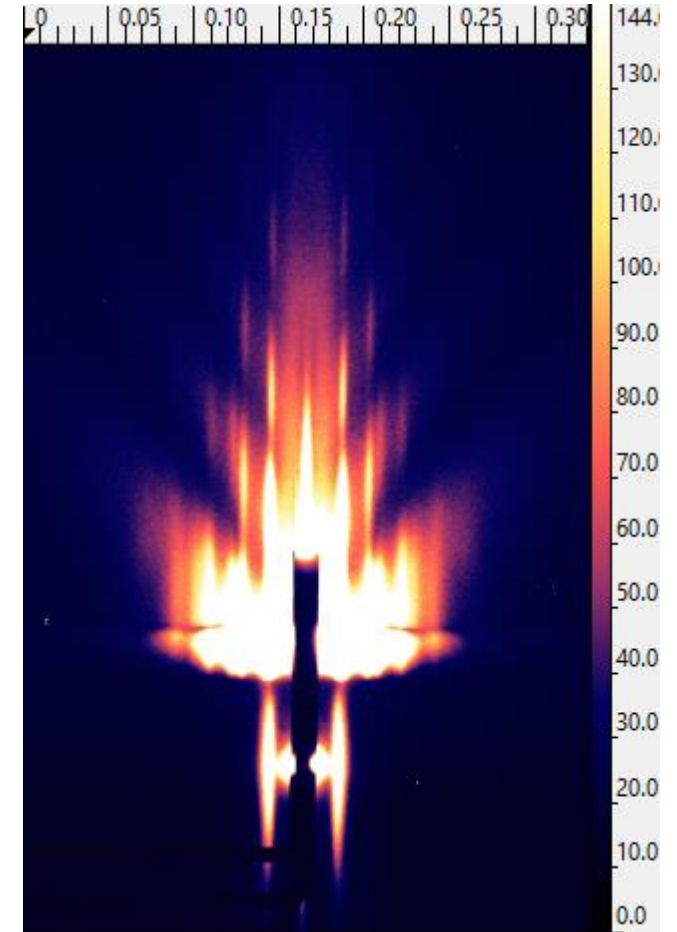
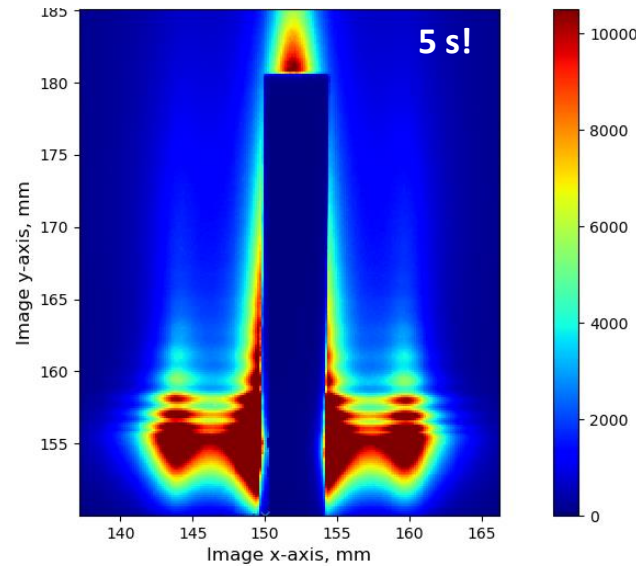


- Horizontal stripes indicate stacked layers of NPs, layer spacing.
- Vertical stripes indicate lateral ordering of the NPs, lateral distance.
- Peak width indicates size distribution and disorder in the system.

Lab (McGill, Anton Parr)



Synchrotron (WLE)

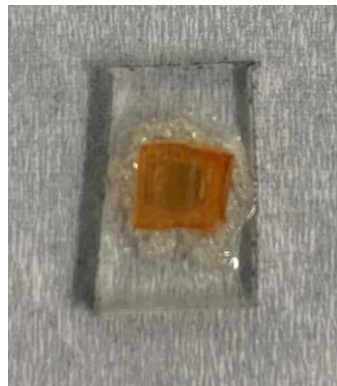


$$I(Q) = S(Q) \times P(Q)$$

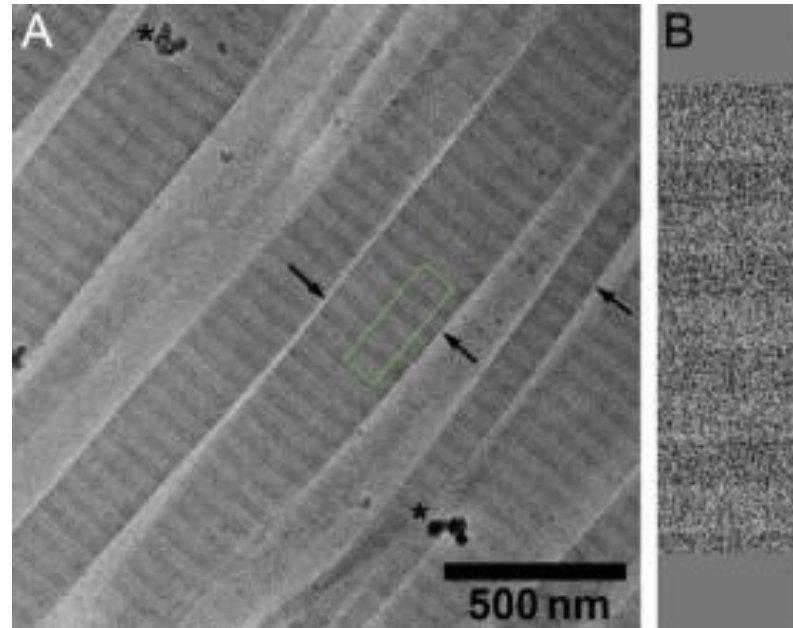
**S(Q)** is the **structure** factor

- *Inter*-particle interferences
- High concentrations (>5% vol.)
- Ordering/packing of particles

Type 1 collagen fibril gaps

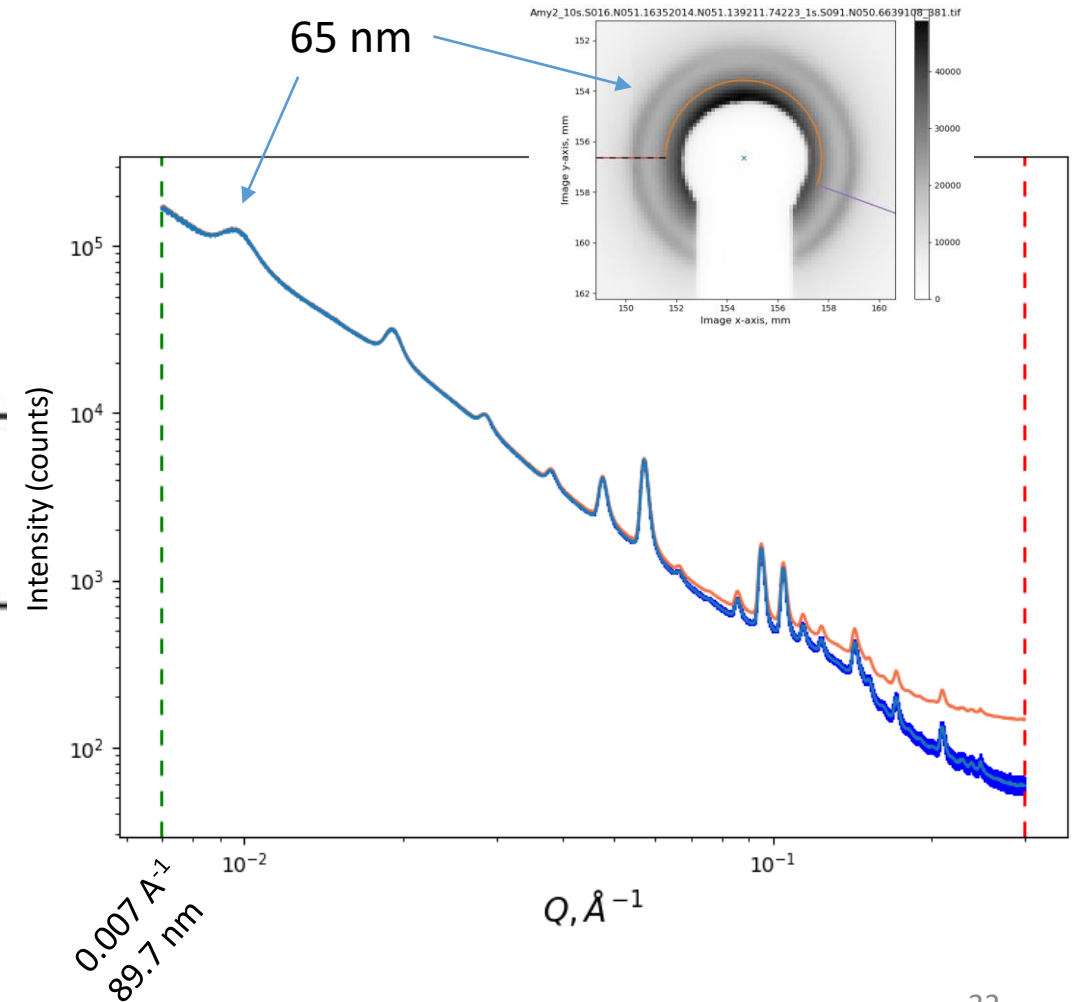


Sea cucumber dermis  
M. Harrington, McGill



<https://doi.org/10.1016/B978-0-12-416617-2.00009-6>

~An extension of PXR  
Looking for and analyzing Bragg peaks



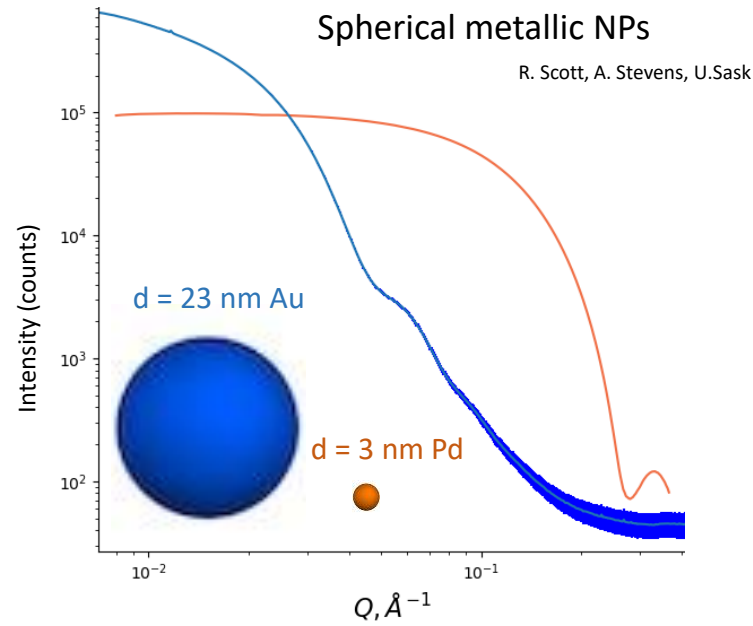


# Origins of the SAXS signal

$$I(Q) = S(Q) \times P(Q)$$

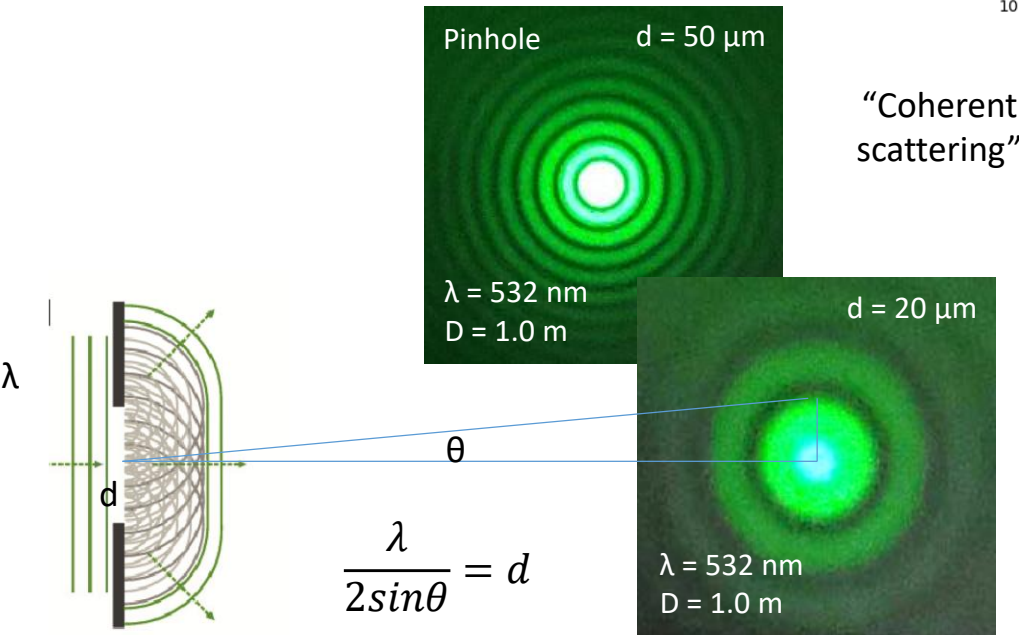
**P(Q)** is the **form factor**

- *Intra*-particle interferences
- Low concentration (dilute limit)
- **Size, shape, polydispersity of particles**

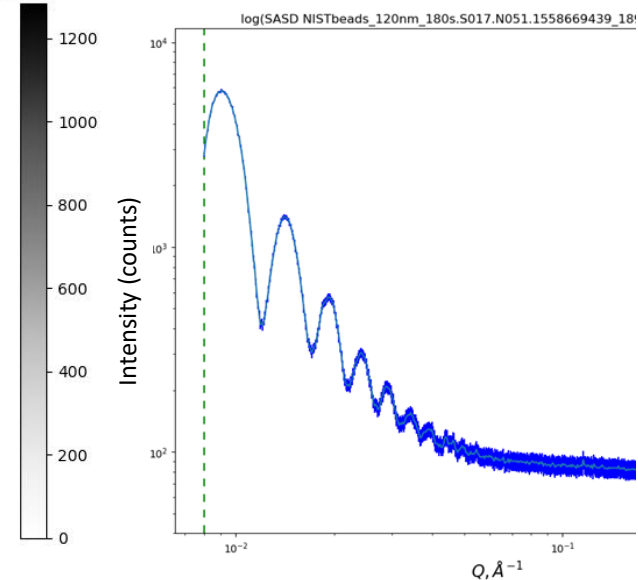
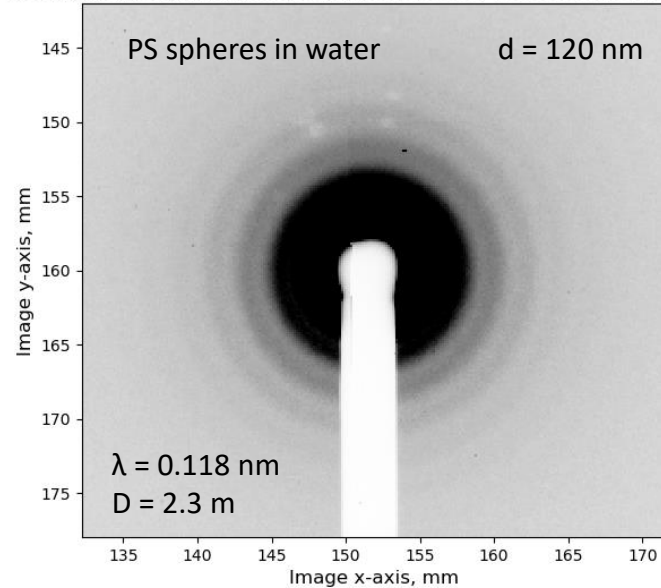


Sample prep is critical !!!

1. Dilute (~1 – 10 mg/mL)  
no interparticle interactions
2. Pure
3. Monodisperse

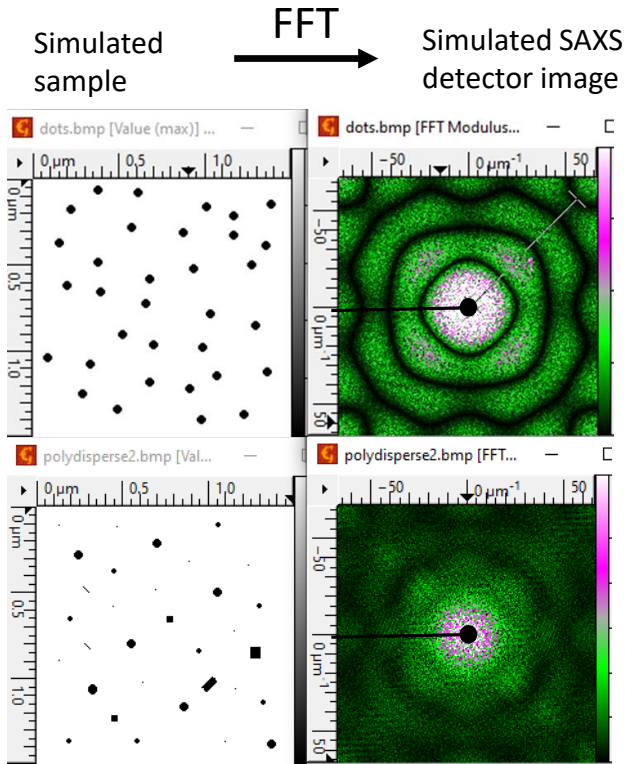


120nm\_NIST\_std\_quartz\_300s.S014.N051.4056907486906665264173\_365.tif

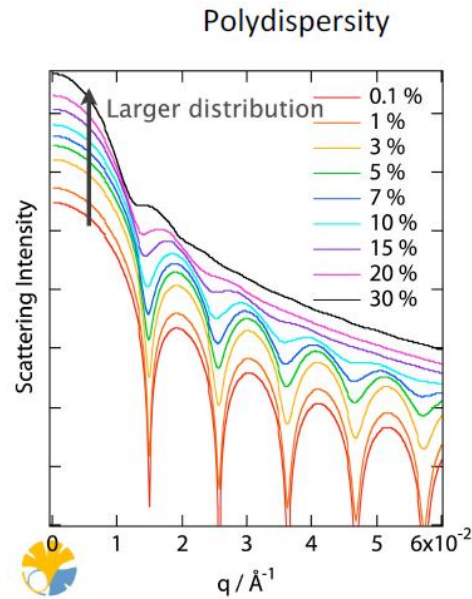
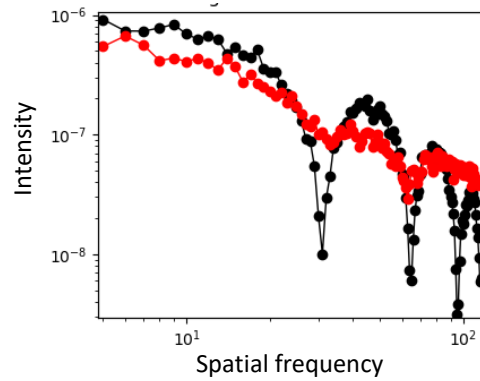


# But there are limits!

Solutions must be monodisperse, and pure

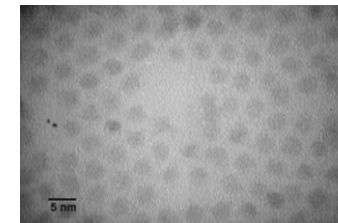
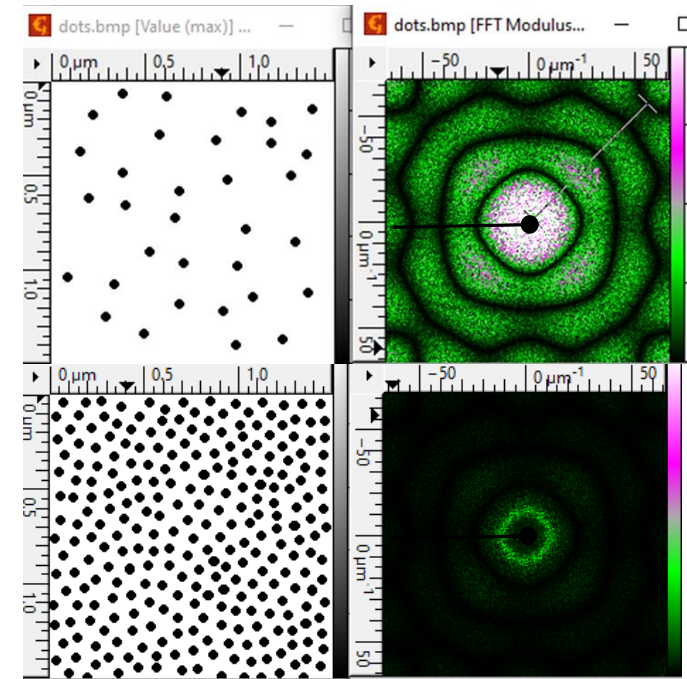


Form factor oscillations smear out when size polydispersity is  $\geq 15\%$

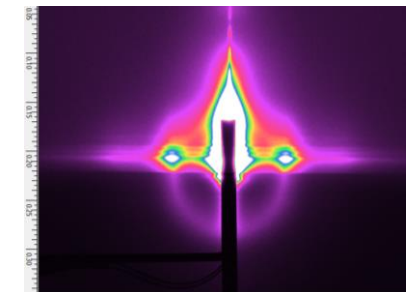


Solutions must be dilute: No interparticle effects

$\sim 1 - 10$  mg/mL... Bring many concentrations



M. Yuan et al., Adv. Mater. 2014, 26, 3513–3519



# Form factor data analysis: Catalytic nanoparticles

Guinier's approximation:

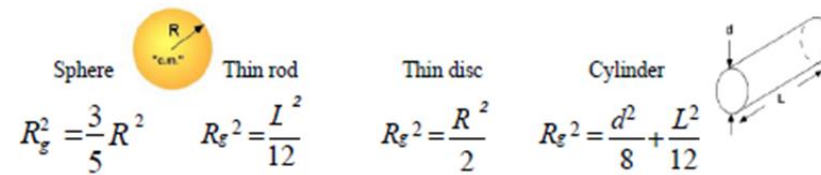
$$I(q) \approx I(0)e^{-q^2 R_g^2/3}$$

$R_g$ : Radius of gyration  
 $I(0)$ : Intensity at  $q=0$

$R_g$  is the square root of the average squared distance of each scatterer from the particle center

RMS distance of the objects parts from its center of mass

➤ Particle radius



$q$  range required

If  $q_{min} = 0.005 \text{ \AA}^{-1}$

Globular, disc:  $q_{min} * R_g < 1.0$  to  $\sim 1.3$

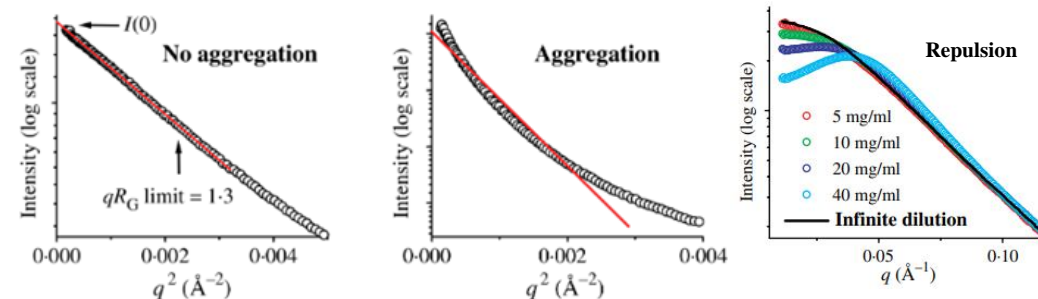
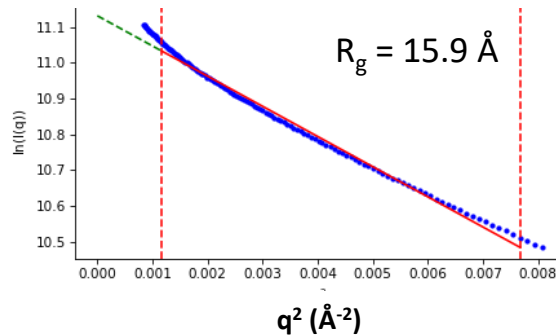
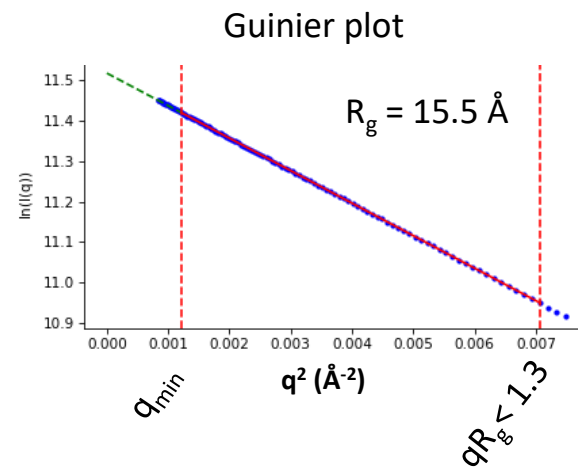
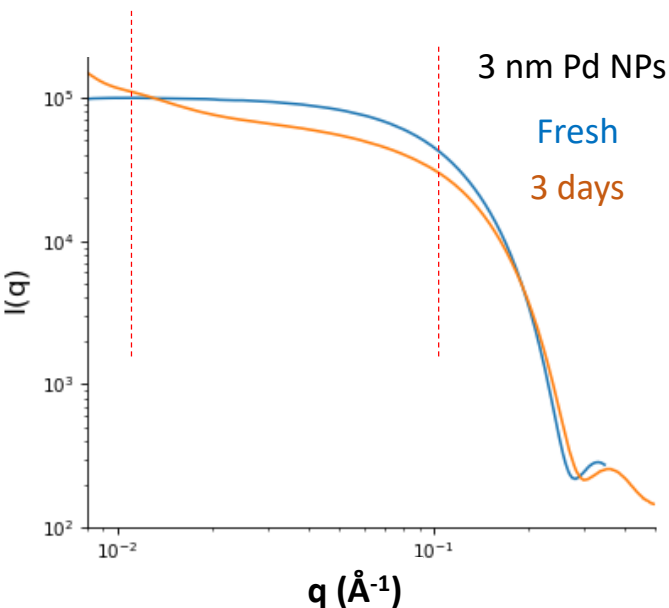
$R_{g,max} = 20 \text{ nm}$

Linear:  $q_{min} * R_g < 0.65$  to  $\sim 1.0$

$R_{g,max} = 13 \text{ nm}$

[https://bioxtas-raw.readthedocs.io/en/latest/saxs/saxs\\_guinier.html](https://bioxtas-raw.readthedocs.io/en/latest/saxs/saxs_guinier.html)  
 D. Putnam et al. Quart. Rev. Biophys. 40, 191-285 (2007)

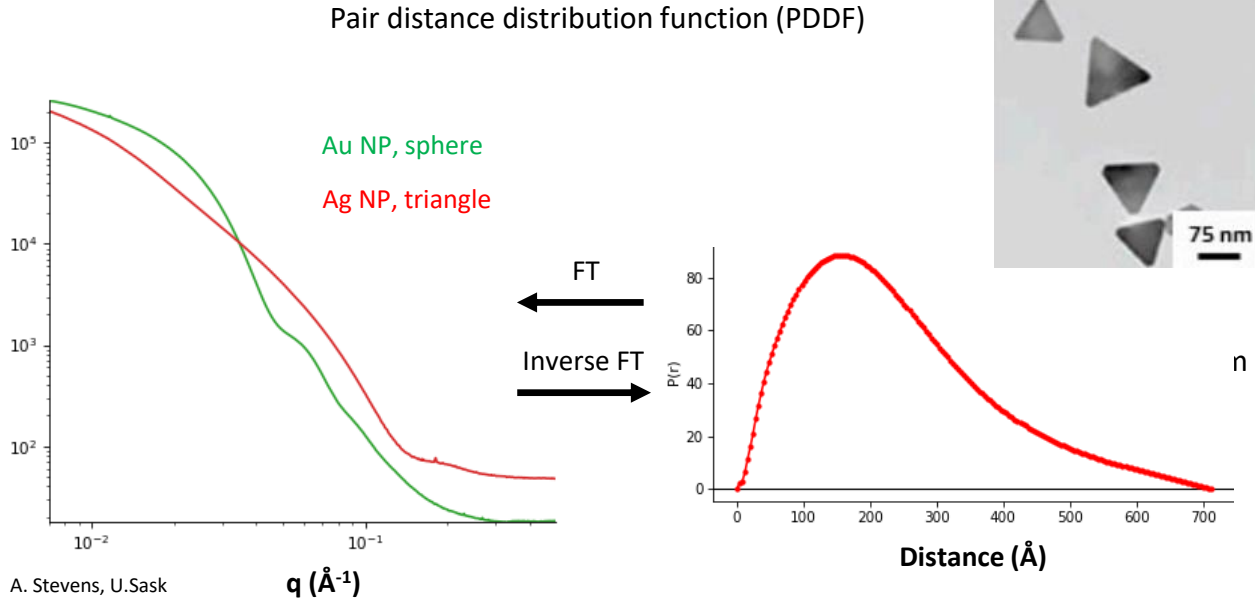
➤ Sample quality: non-linearity can indicate aggregation, repulsion, radiation damage



D. Putnam et al. Quart. Rev. Biophys. 40, 191-285 (2007)



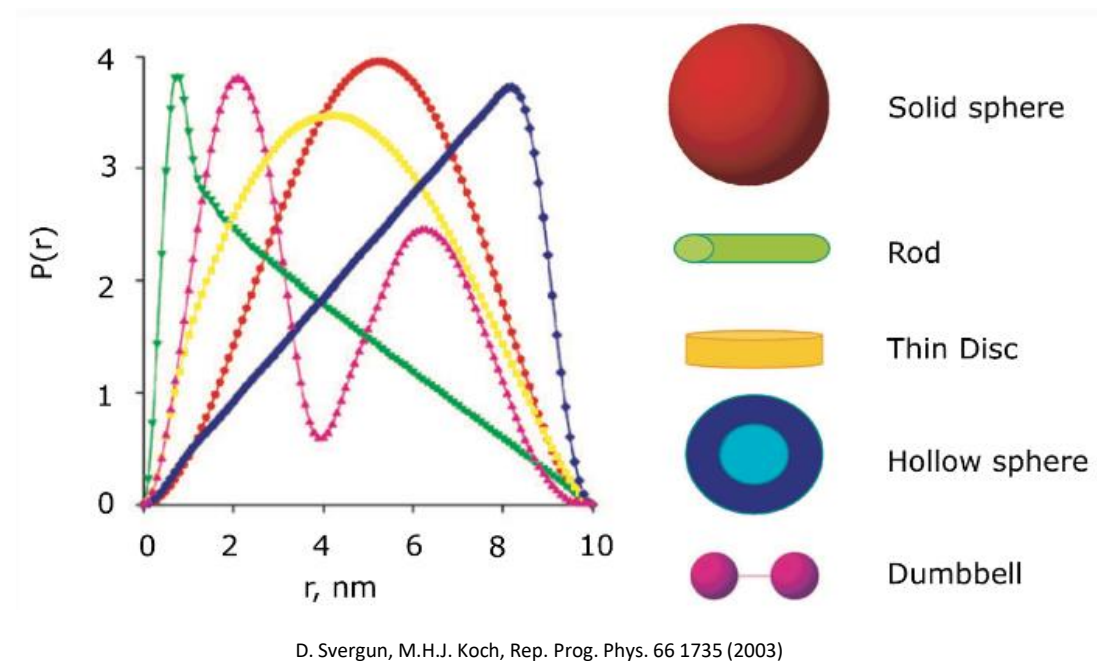
# Form factor data analysis: Catalytic nanoparticles



Reciprocal space intensity

Real space distance frequency

Histogram of atom-atom distances within a single particle, weighted by respective electron densities



$D_{\max}$  – **Largest interatomic distance** in the scattering particle

Shape of the P(r) function is related to the **shape** of the particle, with low spatial resolution

q range required

$$q_{\min} \leq \pi / D_{\max} \quad (\pi / 0.005 \text{ \AA}^{-1} = 62 \text{ nm})$$

$$q_{\max} \geq 2\pi / D_{\max}$$

D. Putnam et al. Quart. Rev. Biophys. 40, 191-285 (2007)

# Optimized Chitosan-Based Nanoemulsion Improves Luteolin Release

C. Diedrich, I. Badea et al., *Pharmaceutics* 15, 1592 (2023)

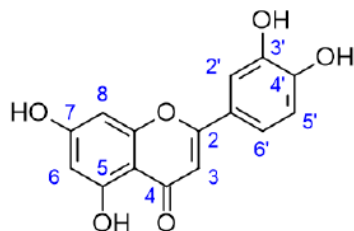


Figure 1. Luteolin structure.

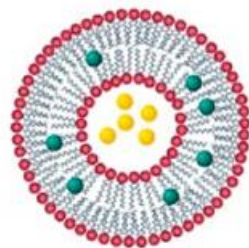
Anti-oxidant, -inflammatory, -tumor, -viral  
Poor absorption after oral administration, limited water solubility

*Nano-encapsulation might improve the solubility of luteolin?*

Diameter: 20 – 200 nm  
Bilayer: ~5 nm



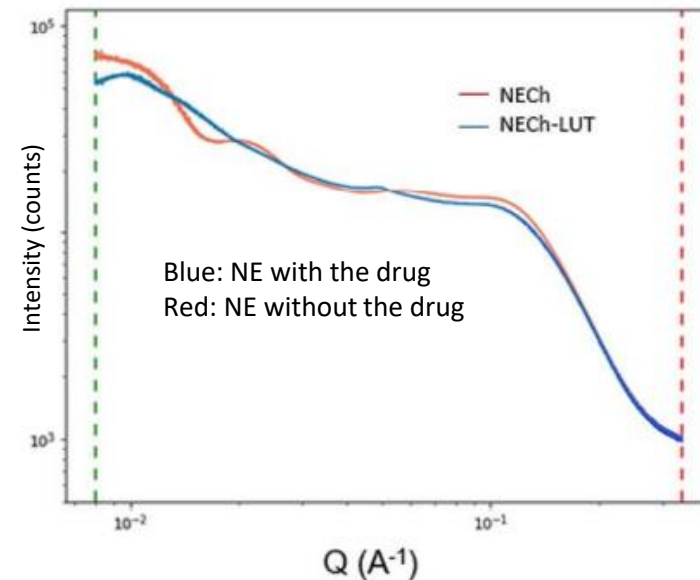
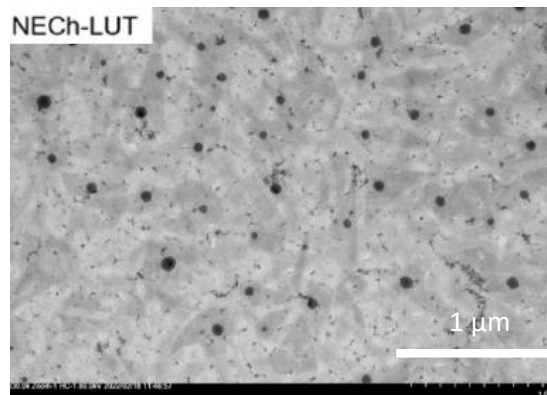
liposome



drug-loaded liposome

Oil phase: Luteolin, oleic acid, with ethylene glycol and Tween 20 as surfactants  
Aqueous: Chitosan solubilized in 0.25% acetic acid.

Drop aqueous into oil while sonicating



The most reasonable fit was obtained modeling two spherical droplets

$d = 27.0 \pm 2.6$  nm

$d = 1.4 \pm 0.26$  nm

Liposomes

Tween 20 micelles

*Where is luteolin?*

1. Dilute (~1 – 10 mg/mL), no interparticle interactions
2. Pure
3. Monodisperse

**Sample prep is critical !!!**



# Proteins in solution

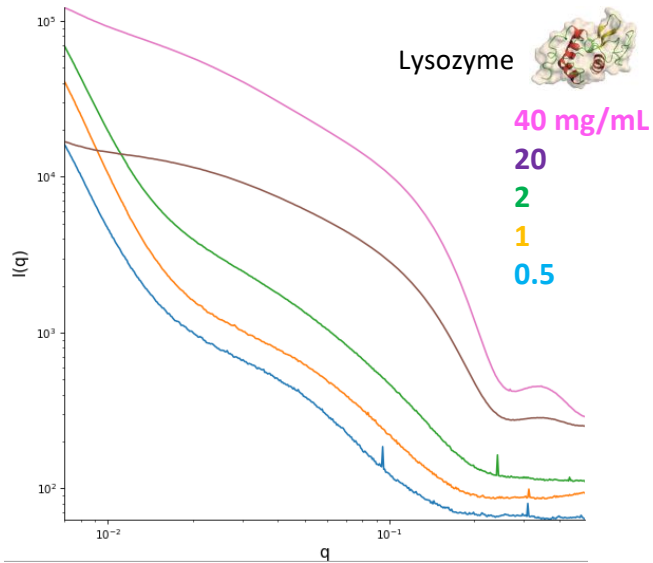
Several dedicated Bio-SAXS beamlines

Aid structure determination

- Size, molecular weight (>10%) to verify the oligomeric state
- Low resolution shape

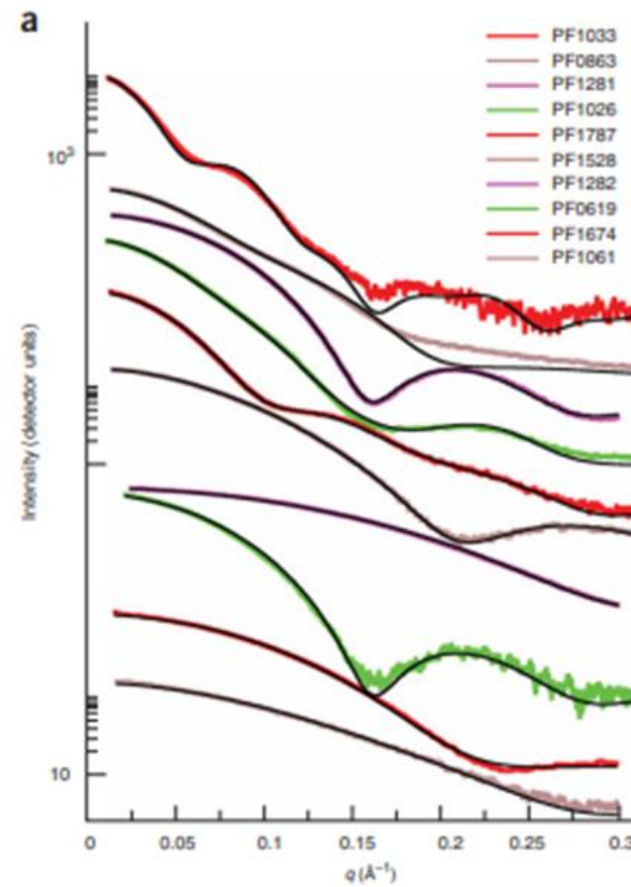
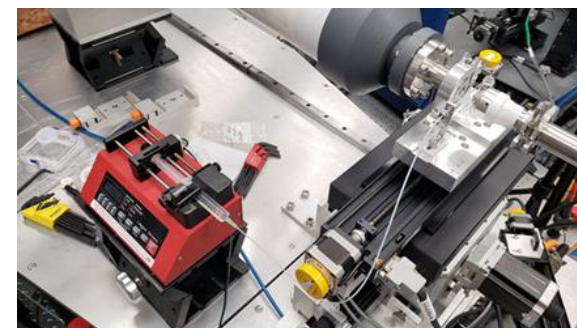
Challenging !!

- Weakly scattering
- Radiation damage
- Limited amounts of material (~30  $\mu$ L)

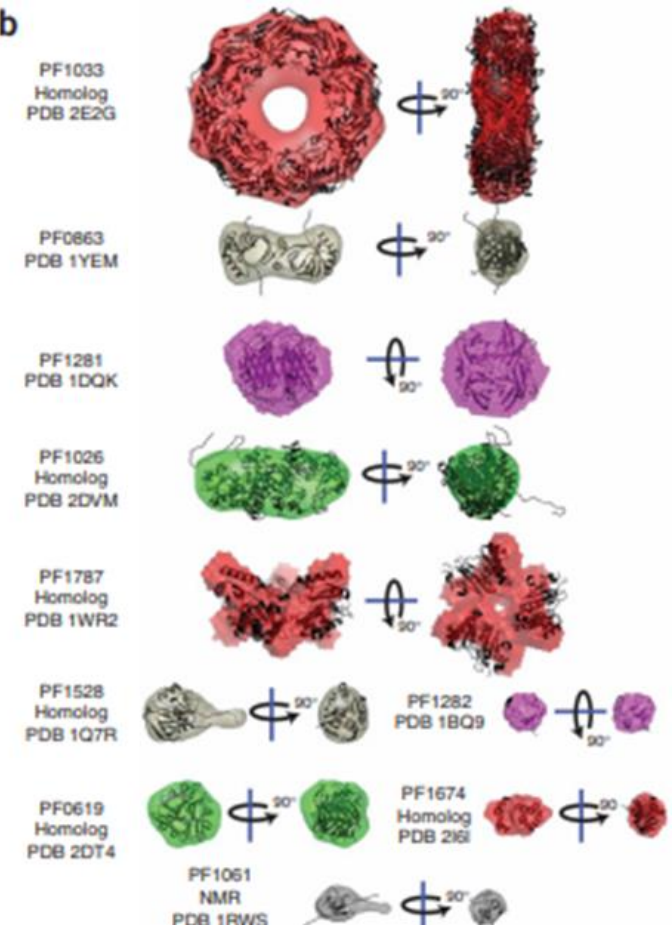


buffer

2 mg/mL



b



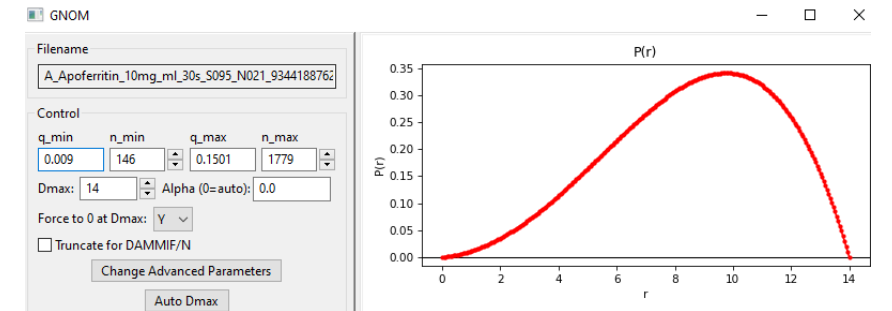
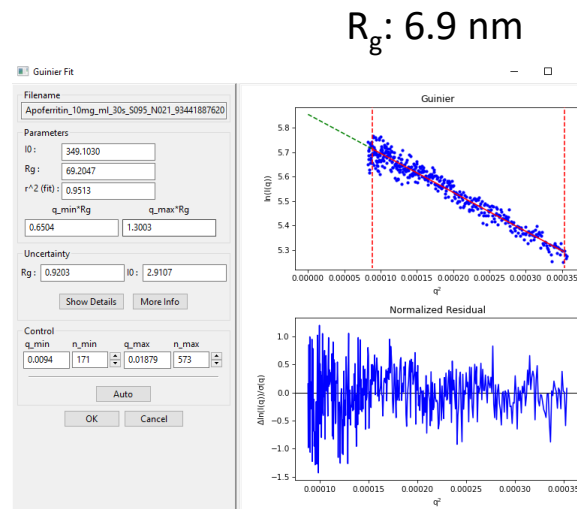
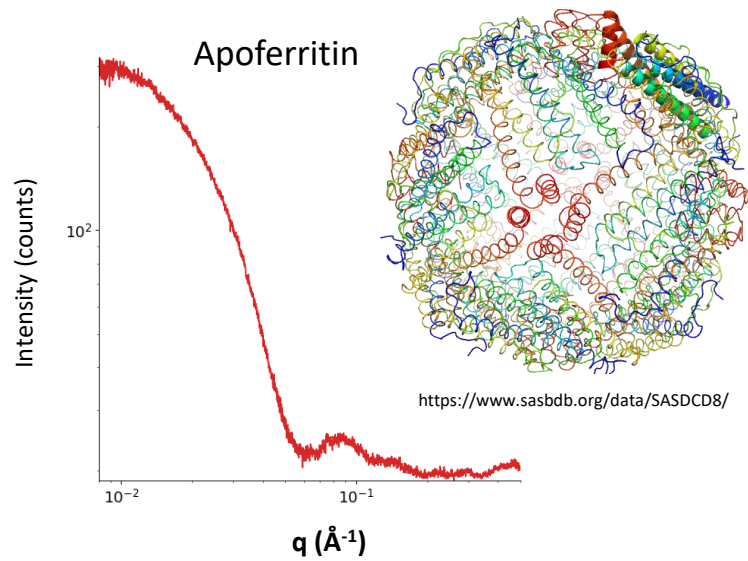
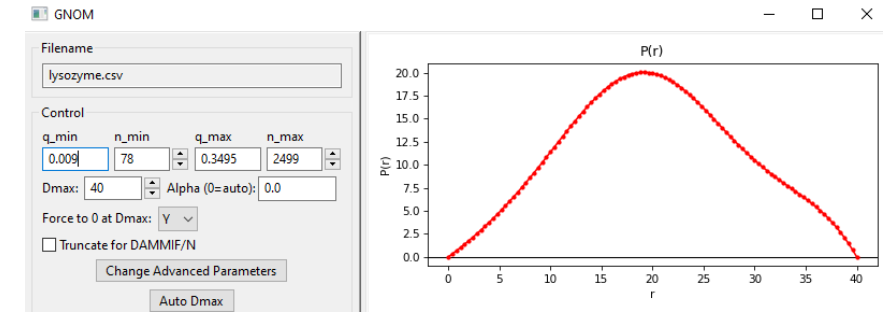
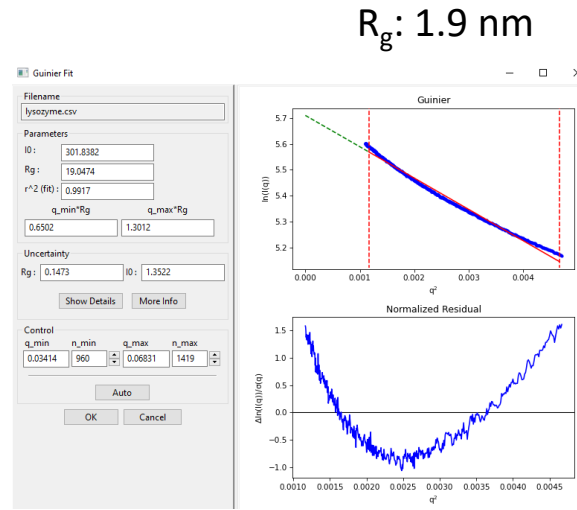
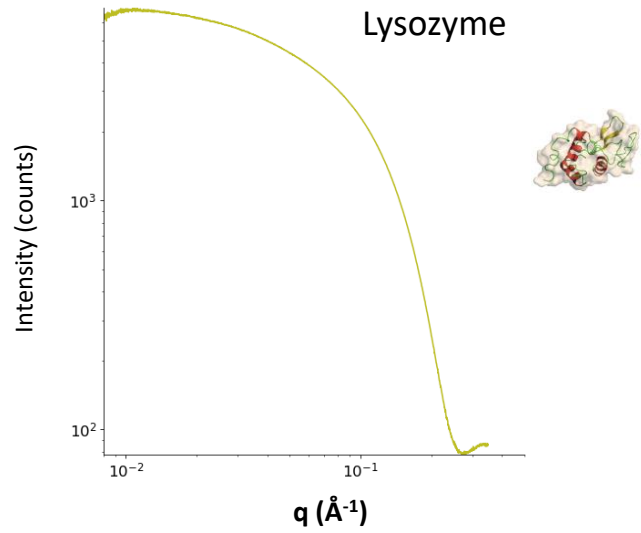
G.L. Hura et al., *Nature Methods* 6(8), 606 (2009)



Canadian Light Source  
Centre canadien de rayonnement synchrotron

# Proteins in solution

D. Svergun EMBL group example



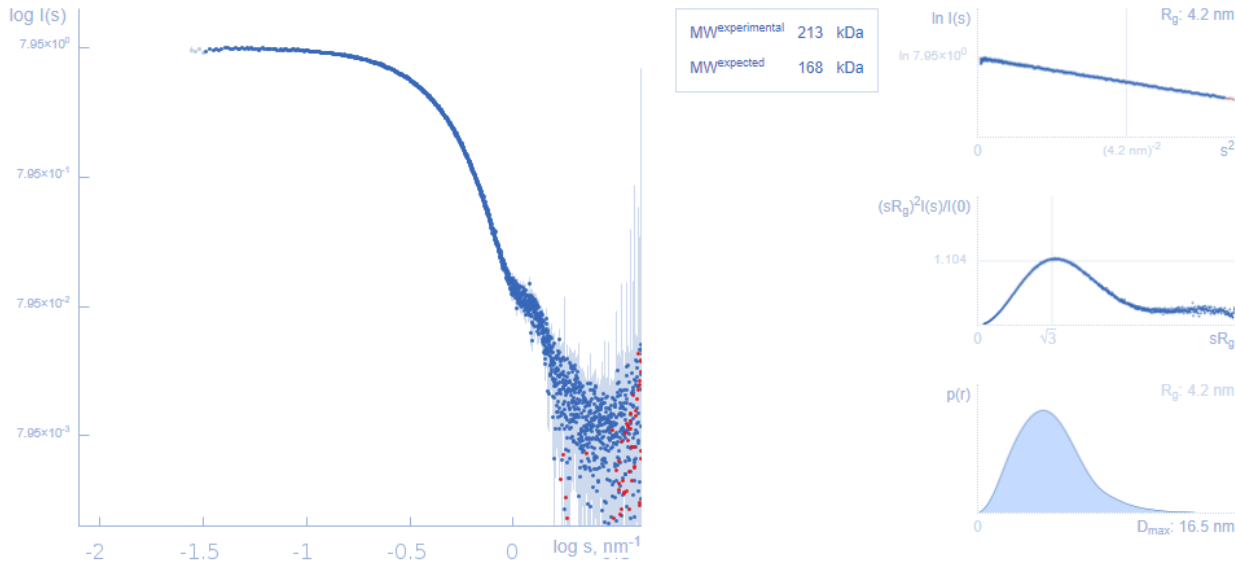
### LARGE1 Processively Polymerizes Matriglycan Using Active Sites on Alternate Protomers

Joseph S, Schnicker N, Xu Z, Yang T, Hopkins J, Watkins M, Chakravarthy S, Davulcu O, Anderson M, Venzke D, Campbell K, SSRN  
Electronic Journal () DOI

[Download files](#)

### SASDNH8 – Xylosyl- and glucuronyltransferase LARGE2 (LARGE2dTM) dimer

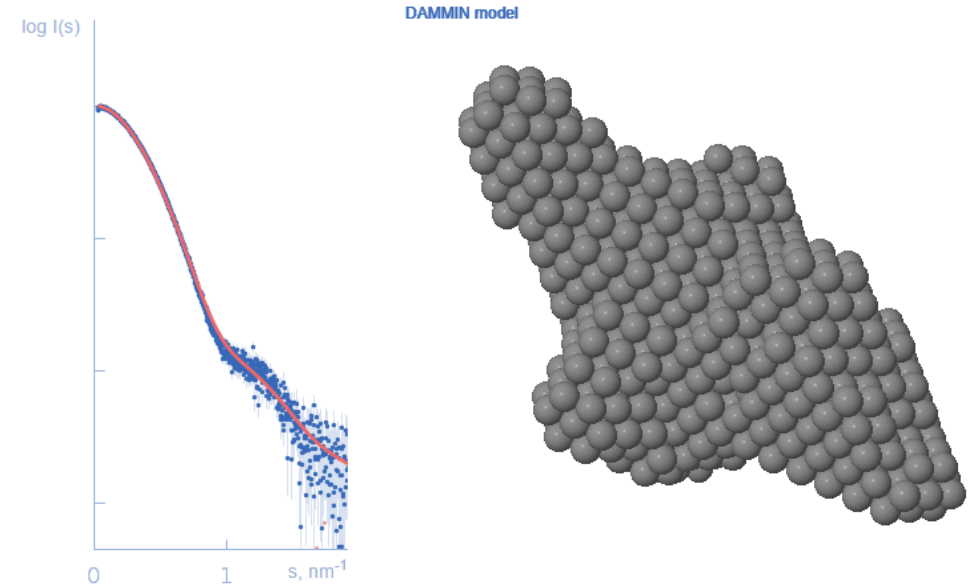
Xylosyl- and glucuronyltransferase LARGE2



Synchrotron SAXS data from solutions of a secreted form that lacks the transmembrane domain of mouse LARGE xylosyl- and glucuronyltransferase 2 (LARGE2dTM) in buffer (20 mM HEPES pH 7.4, 150 mM NaCl) were collected on the BioCAT 18-ID-D beamline at the Advanced Photon Source (APS) (Chicago, IL, USA) using a Eiger2 XE 9M detector at a sample-detector distance of 3.67 m and at a wavelength of  $\lambda = 0.1033$  nm ( $I(s)$  vs  $s$ , where  $s = 4\pi\sin\theta/\lambda$ , and  $2\theta$  is the scattering angle). In-line size-exclusion chromatography (SEC)-MALS-SAXS was employed. The SEC parameters were as follows: A 300  $\mu$ l sample at 4 mg/ml was injected at a 0.6 ml/min flow rate onto a Superdex 200 increase 10/300 GL column (GE healthcare) at 23°C. 2500 successive 1 second frames were collected. The data were normalized to the intensity of the transmitted beam and radially averaged; the scattering of the solvent-blank was subtracted.

boundary conditions for bead modelling programs

#### Fits and models



ATSAS package  
DAMMIF/DAMMIN  
GASBOR

Low resolution protein structure





## Summary

The CLS has a SAXS endstation available and our user community is growing!

Probe nano-scale dimensions and ordering (1 - 150 nm) with X-rays by measuring elastic scattering at small angles

Can learn about 1) packing, 2) particle size, and 3) particle shape, but generally not all 3 at the same time

Limited structural information

Complementary information, that should be supported with supplementary techniques such as microscopy



## Further reading

### Basic SAXS concepts

H. Schnablegger, Y. Singh, “The SAXS Guide: Getting Acquainted With the Principles”, Anton Parr, Austria. (2017)

### Making a good measurement

B.R. Pauw, “Everything SAXS: small-angle scattering pattern collection and correction” J. Phys.: Condens. Matter 25 (2013) 383201

### Basic data work up

J.B. Hopkins, R.E. Gillilan, S. Skou  
[https://bioxtas-raw.readthedocs.io/en/latest/saxs\\_tutorial.html](https://bioxtas-raw.readthedocs.io/en/latest/saxs_tutorial.html)

### BioSAXS, data workup, introduction to structure modelling

C.D. Putnam, M. Hammel, G.L. Hura, J.A. Tainer, “X-ray solution scattering (SAXS) combined with crystallography and computation: defining accurate macromolecular structures, conformations and assemblies in solution” Quarterly Reviews of Biophysics 40, 3 (2007), pp. 191–285.

## Tutorials

### Data processing

- Averaging
- Absolute intensity scale
- Data reduction

### Data analysis

- Guinier plots
- Kratky plots
- PDDF
- Amphiphile phase identification

# The Brockhouse Sector Team



Al Rahemtulla



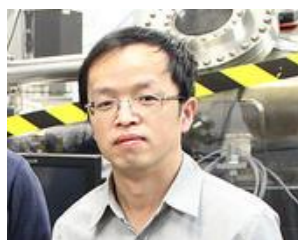
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**Call for General User Proposals opens July 24, closes August 21, 2024**

